

Soluble SLAMF7 is generated by alternative splicing in multiple myeloma cells

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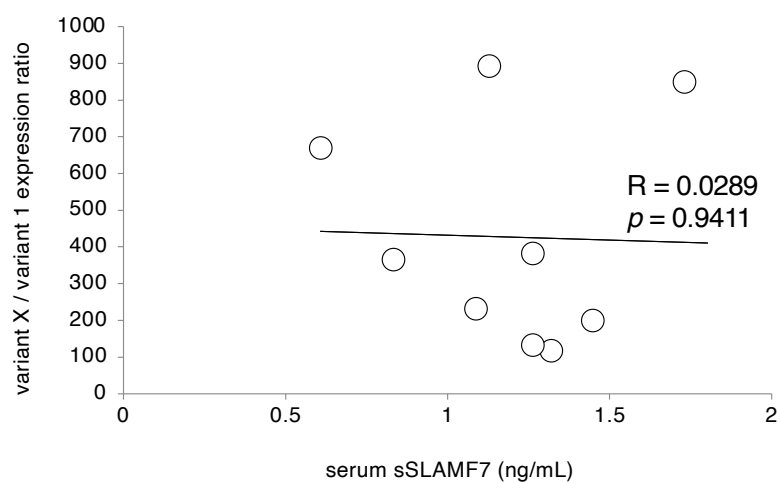
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Figure S1

A



B

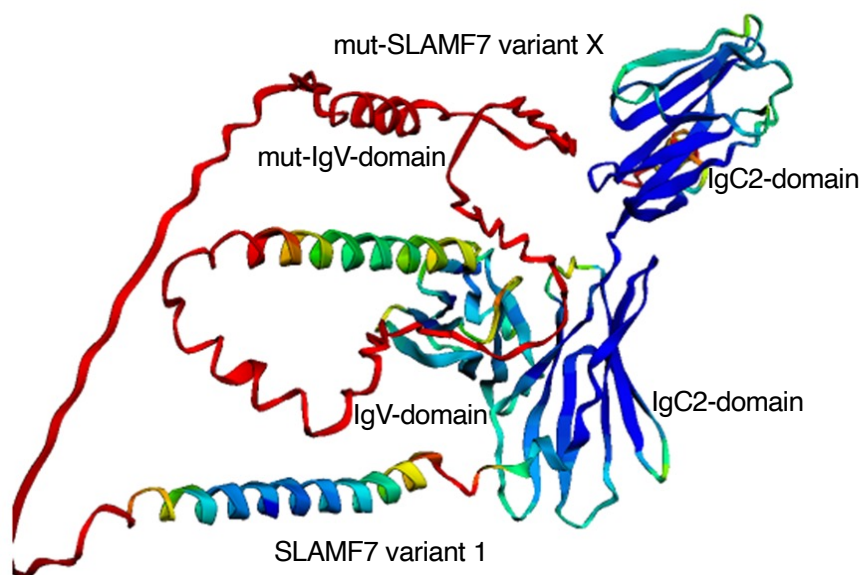


Figure S1. (a) The expression levels of *SLAMF7* variant 1 in 10 MM patients were determined by Q-PCR, normalized to that of *GAPDH*, and quantified using the $2^{-\Delta\Delta C_t}$ method with the values of BM-MNC set at 1.0. Serum levels of soluble *SLAMF7* (s*SLAMF7*) were also measured using the enzyme-linked immunosorbent assay (ELISA). The correlation between *SLAMF7* variant X/variant 1 expression ratio and s*SLAMF7* was evaluated by calculating Pearson's correlation coefficient. (b) Structure-based prediction of *SLAMF7* variant 1-mutant *SLAMF7* variant X (mut-*SLAMF7* variant X) interactions was performed using the AlphaFold2 program. The mutant *SLAMF7* variant X was designed by randomly rearranging the amino acid sequence of the IgV domain.

Figure S2

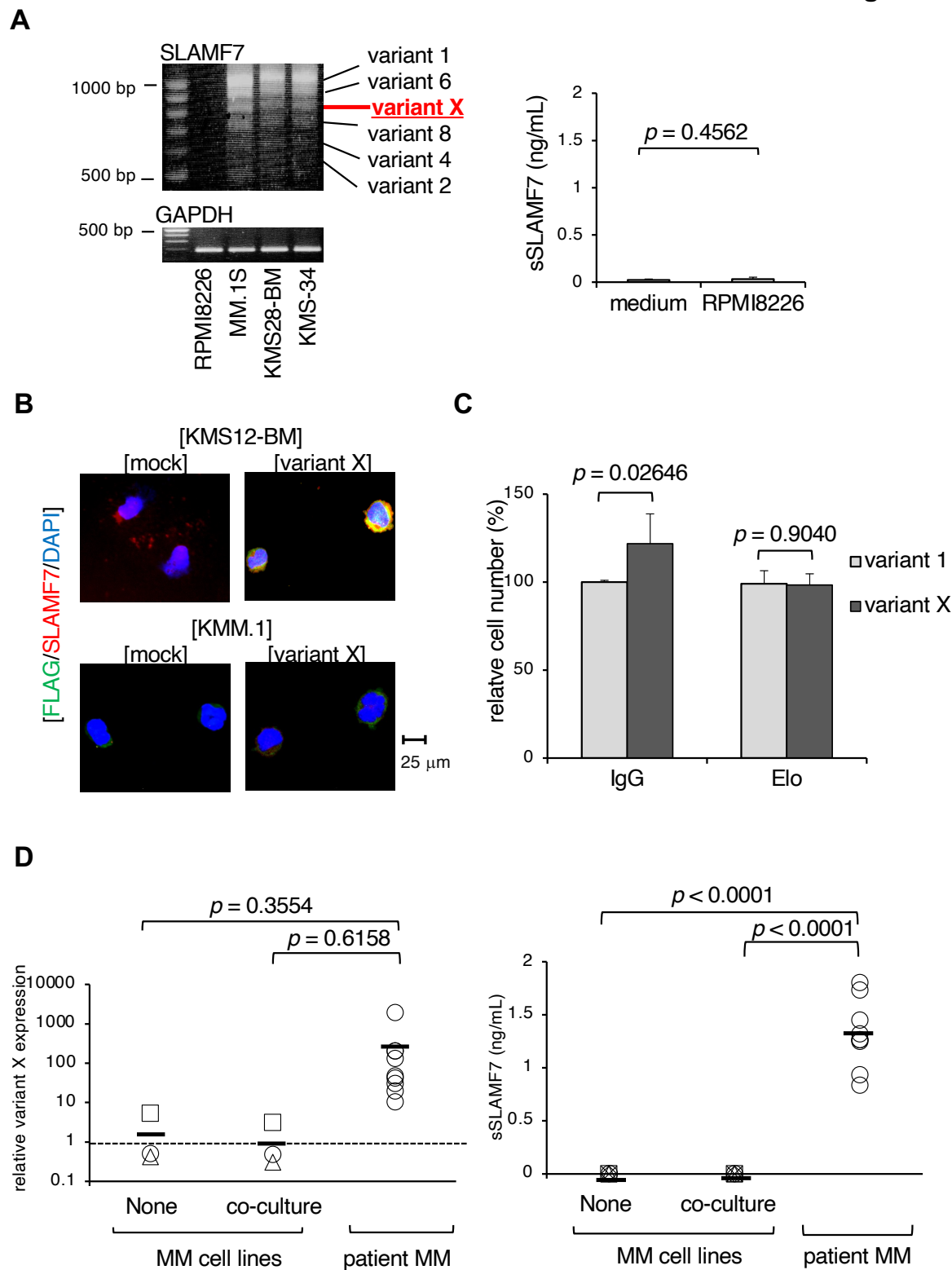


Figure S2. (a) Left panel: The full length *SLAMF7* or *GAPDH* (internal control) was amplified from the cDNA of RPMI8226, MM.1S, KMS28-BM, and KMS-34 cells. PCR was conducted for 40 cycles and the products were analyzed using agarose gel electrophoresis and ethidium bromide staining. Right panel: The supernatants were prepared from 1×10^5 cells of RPMI8226 cells and the concentrations of sSLAMF7 were measured using the enzyme-linked immunosorbent assay (ELISA) (right panel). Bars indicate the means of three independent experiments. *P* value was calculated using a paired Student's *t* test. (b) Cells were mounted onto glass slides using a Cytospin device (Shandon Scientific, Cheshire, England). Immunostaining was performed using PE-conjugated anti-SLAMF7 (Thermo Fisher Scientific) and Alexa Fluor 488-conjugated anti-FLAG (MBL, Tokyo, Japan) antibodies at 1:200 dilution. The nuclei were counterstained with DAPI. Only merged images are shown. Data shown are representative of multiple independent experiments. (c) MM.1S cells were cultured with 5ug/mL of human IgG (IgG) or elotuzumab (Elo) in the supernatant derived from 1×10^5 RPMI8226 cells transduced with *SLAMF7* variant 1 or *SLAMF7* variant X after 24 h of culture. Cell proliferation was assessed after 72 h using the MTT reduction assay and is shown relative to the variant 1 group. Bars indicate the mean of three independent experiments. *P* value was calculated using a paired Student's *t* test. (d) Left panel: KMS12-BM, KMS26, and MM.1S cells were cultured with or without UBE6T-7 cells for 24 hours, respectively. The expression levels of *SLAMF7* variant X in KMS12-BM (Δ), KMS26 (\diamond), and MM.1S (\square), and patient MM cells (\circ) were determined by Q-PCR, normalized to that of *GAPDH*, and quantified using the $2^{-\Delta\Delta Ct}$ method with the values of BM-MNC set at 1.0. Right panel: The supernatants were prepared from 1×10^5 cells of KMS12-BM (Δ), KMS26 (\diamond), and MM.1S (\square) cells co-cultured with or without UBE6T-7 cells for 24 hours, respectively, and the concentrations of sSLAMF7 were measured using ELISA. Serum levels of sSLAMF7 in MM patients (\circ) were also measured using the ELISA. *P* value was determined by one-way ANOVA with Tukey's multiple comparison test.

Supplementary Table S1. PCR primers used in this study

gene	sequence (location)	product size
SLAMF7 variant 1 (full length)	forward: 5'- agggaagtggcttcatttcagtg -3'(exon 1) reverse: 5'- tctcataggcaaatagccttggt -3' (exon 7)	1100 bp
SLAMF7 variant 1	forward: 5'- acagctcatcactccagcagcc-3'(exon 2) reverse: 5'- agtgtgagggattgtgtcgtac-3' (exon 5)	547 bp
SLAMF7 variant X	forward: 5'- aagctctgtgaagagaacaatcc -3' (exon 3/6) reverse: 5'- gcagagacttaggggagtg -3' (exon 7)	162 bp
GAPDH	forward: 5'-gagtcaacggatttggcgt-3' reverse: 5'-gacaagcttcccgttctcag-3'	185 bp