



Clinical phenotype of adult-onset systemic histiocytosis harboring BRAF in-frame deletions

by Matthias Papo, Jérôme Razanamahéry, Malik Da Silva, Zofia Hélias-Rodzewicz, Vsevolod Potapenko, Suzanna Bota, Vanessa Leguy-Seguín, Stéphane Dominique, Raphaël Lhote, Quentin Moyon, Dov Taïeb, Tom Abrassart, Marion Campana, Visal Keo, Etienne Rivière, Olivier Lucidarme, Fleur Cohen-Aubart, Zahir Amoura, Julien Haroche, and Jean-François Emile

Received: March 4, 2024.

Accepted: May 15, 2024.

Citation: Matthias Papo, Jérôme Razanamahéry, Malik Da Silva, Zofia Hélias-Rodzewicz, Vsevolod Potapenko, Suzanna Bota, Vanessa Leguy-Seguín, Stéphane Dominique, Raphaël Lhote, Quentin Moyon, Dov Taïeb, Tom Abrassart, Marion Campana, Visal Keo, Etienne Rivière, Olivier Lucidarme, Fleur Cohen-Aubart, Zahir Amoura, Julien Haroche, and Jean-François Emile. Clinical phenotype of adult-onset systemic histiocytosis harboring BRAF in-frame deletions. *Haematologica*. 2024 May 23. doi: 10.3324/haematol.2024.285273 [Epub ahead of print]

Publisher's Disclaimer.

E-publishing ahead of print is increasingly important for the rapid dissemination of science. Haematologica is, therefore, E-publishing PDF files of an early version of manuscripts that have completed a regular peer review and have been accepted for publication.

E-publishing of this PDF file has been approved by the authors.

After having E-published Ahead of Print, manuscripts will then undergo technical and English editing, typesetting, proof correction and be presented for the authors' final approval; the final version of the manuscript will then appear in a regular issue of the journal.

All legal disclaimers that apply to the journal also pertain to this production process.

Clinical phenotype of adult-onset systemic histiocytosis harboring *BRAF* in-frame deletions

Matthias Papo¹, Jérôme Razanamahéry², Malik Da Silva³, Zofia Hélias-Rodzewicz³, Vsevolod Potapenko⁴, Suzanna Bota⁵, Vanessa Leguy-Seguin², Stéphane Dominique⁵, Raphaël Lhote¹, Quentin Moyon¹, Dov Taïeb¹, Tom Abrassart¹, Marion Campana⁶, Visal Keo⁷, Etienne Rivière⁸, Olivier Lucidarme⁹, Fleur Cohen-Aubart¹, Zahir Amoura¹, Julien Haroche¹, Jean-François Emile³

¹Internal Medicine Department 2, French National Referral Center for Rare Systemic Diseases and Histiocytoses, Sorbonne University, Assistance Publique–Hôpitaux de Paris, Pitié-Salpêtrière Hospital, Paris, France

²Department of Internal Medicine and Clinical Immunology, Dijon University Hospital, Dijon, France

³Paris-Saclay University, Versailles SQY University, Assistance Publique-Hôpitaux de Paris, Ambroise-Paré Hospital, Smart Imaging, Service de Pathologie, Boulogne, France

⁴Municipal educational hospital №31, Saint-Petersburg, Russia

⁵Department of Pneumology, CHU ROUEN, F-76000 ROUEN, France

⁶Pneumology Department, Source hospital, Orléans, France

⁷Internal medicine departement, Bayonne hospital, Bayonne France

⁸Department of internal Medicine and Infectious Diseases Department, Haut Leveque Hospital, University Hospital Centre of Bordeaux, F33604 Pessac, France

⁹APHP Sorbonne Universités Pitié-Salpêtrière Hospital and UMR 7371, UMR_S 1146, Laboratoire d'Imagerie Biomédicale, F-75013, Paris, France.

Corresponding author: Dr Matthias Papo, Service de Médecine Interne 2, Institut E3M, Hôpital Pitié Salpêtrière, 47-83, boulevard de l'Hôpital 75013, Paris, France; matthias.papo@aphp.fr

Running title: Clinical features of *BRAF*del mutated histiocytosis

Key words: Histiocytosis; Cholangitis; Mutations

Funding: nothing to disclose

Competing interests: All authors have nothing to disclose.

Patient consent for publication: Not required.

Patient and Public involvement: This research was done without patient involvement. Patients were not invited to comment on the study design and were not consulted to develop patient relevant outcomes or interpret the results. Patients were not invited to contribute to the writing or editing of this document for readability or accuracy.

Data availability statement: All data relevant to the study are included in the article or uploaded as supplementary information.

Contributors: Study conception and design: MP, JH, JFE. Acquisition of data and experiments performance: MP, JR, MDS, ZHR, VP, SB, VLS, SD, RL, QM, DT, TA, MC, VK, ER, OL, FCA, ZA, JH, JFE. Analysis of data, drafting and writing of the manuscript: MP, JH, JFE. All authors contributed to reviewing the manuscript and approved the final version for publication.

L-group histiocytoses (Erdheim-Chester disease (ECD) and Langerhans-cell histiocytosis (LCH)) are multi-system diseases characterized by histiocytes infiltration in several organs¹⁻³. In these diseases, histiocytes frequently display activating somatic mutations of intracellular signaling pathway protein kinases (mostly the MAPKinase pathway)¹⁻³. Many of these mutations seem to be associated with a specific phenotype: *BRAF*^{V600E} mutated ECD patients have more frequently cardiac and retroperitoneal manifestations⁴, *MAP2K1* mutated ECD patients may exhibit overt Rosai-Dorfman Disease (RDD) component⁵, and *ALK*-mutated patients have a high prevalence of neurological manifestations⁶. Therefore, we suspect that each specific mutation or mutation type could be associated with a specific clinical phenotype of histiocytosis. Our objective was to describe characteristics of patients with histiocytosis and in-frame deletion within exon 12 of *BRAF* (*BRAF*^{Δβ3-αC}).

Inclusion criteria were: 1) Diagnosis of histiocytosis confirmed by central review according to most recent published guidelines^{3,7}, 2) presence of *BRAF*^{Δβ3-αC}, and 3) clinical data available. Patients were retrieved from the files of pathology laboratory in Ambroise-Paré Hospital (Boulogne, France). Clinical, biological and morphological data were retrieved, as well as treatment received and follow-up.

DNA was extracted as previously described⁸. Since 2020, DNA extraction has been automated on a Maxwell® RSC Instrument (Promega, France), with extraction performed according to the supplier's recommendations. For formalin-fixed paraffin-embedded (FFPE) and frozen biological materials, the Maxwell® RSC DNA FFPE Kit and Maxwell® RSC Tissue DNA Kit were used, respectively. The DNA-Seq Next Generation Sequencing (NGS) panel included almost 60 genes covering hot spots or all exons previously reported to be mutated in histiocytoses and genes involved in the MAP-kinase pathway and myeloid

neoplasia. The sequencing data were analyzed depending on the applied technique. Mutations detected by DNA sequencing were interpreted according to standards and guidelines as previously described⁹.

We identified patients with $BRAF^{A\beta3-\alpha C}$ and contacted the centers for clinical, morphological, and biological data.

This study was conducted in accordance with the Declaration of Helsinki. Patients provided written informed consent (clinical trial registration NCT04437381 [Molecular Targets for the Treatment of Histiocytosis HISTIO-TARGET]).

In 429 patients with L-group histiocytosis cohort, 189 had a $BRAF^{V600E}$ mutation (46.2%), 25 had a $BRAF^{A\beta3-\alpha C}$ (5.8%) and 9 another $BRAF$ mutation (2.1%). Among patients with $BRAF^{A\beta3-\alpha C}$, data were available for 20 of them. Patients and mutations characteristics are described in **Table 1 and Figure 1**. Most patients (n=18) had LCH, and two had an ECD, one pure and the other mixed with LCH. Median age at diagnosis was 50.5 years (IQR 34-78). The most frequent manifestations were hepatic (n=9, 45%) and vulvar (8/11 female gender patients, 73%). Other localization were cystic interstitial lung disease (n=7), lytic bone lesions (n=8), classical cutaneous manifestations (n=8), diabetes insipidus (n=8), panhypopituitarism (n=3), pachymeningitis (n=2), long bone osteosclerosis (n=1), perirenal infiltration (n=1), salivary gland infiltration (n=1) and digestive track infiltration (n=1). Among patients with LCH, 2 patients had a single-system pulmonary disease, one patient a single-system liver disease, one patient a single-system multifocal bone disease, and all the others had multi-system manifestations.

Hepatic manifestation was sclerosing cholangitis in all patients, and 5/6 patients had histiocytic infiltration in liver biopsy. All patients with sclerosing cholangitis had biological cholestasis, elevated aminotransferases, and hyperbilirubinemia. No patient had cirrhosis.

Hepatic MRI, when performed, always showed cholangitis (6/6). PET-scan showed liver abnormalities in 4/7 patients (heterogenous liver uptake or uptake in biliary ducts). Six patients had additional mutations in tissue biopsy, including *DNMT3A* (n=4), *TET2* (n=2), *ASXL1* (n=1) and *PGDFRA* (n=1). Among 5 patients who had a bone marrow aspiration, 4 of them had additional mutations, including *DNMT3A* (n=2), *TET2* (n=2), *STAG1* (n=1), *PPM1D* (n=1) and *RAD21* (n=1).

First line treatments included vinblastine (n=6), cytarabine (n=1), methotrexate (n=1), cladribine (n=3), lenalinomide (n=1) and cobimetinib (n=1), with various responses depending on the clinical manifestation. Nine patients did not receive any specific treatment for histiocytosis. Four patients with cholangitis received ursodeoxycholic acid without significant improvement. Patients with cholangitis also received vinblastine (n=3 with one disease progression, one stable disease, and one partial remission), cladribine (n=1 with stable disease), cytarabine (n=1 with stable disease) and lenalimode (n=1 with stable disease). Two patients received cobimetinib, that resulted in partial remission in both patients (PERCIST criteria) at 6 months, while liver function testes and bili-MRI remained stable in one patient (**Figure 2**). One patient had a liver transplant, with no further relapse. After a median follow-up of 47 (IQR 13-315) months, one patient had died from coronary heart disease.

Clinical manifestations of L-group histiocytosis may vary from single-organ benign disease to multi-organ life-threatening neoplasm^{2,3}. To date, the cause of the variety of clinical manifestations in these diseases is unknown, and the type of mutation involved could play a role in the clinical phenotype⁴⁻⁶.

Our study is the first to describe the clinical phenotype of histiocytosis patients with a *BRAF*^{*Δβ3-αC*}, and showed a high frequency of sclerosing cholangitis and vulvar manifestations,

which are typical LCH manifestations but usually rarely observed. In previous published cohorts of adults, liver manifestations are described in 10-15% of LCH cases¹⁰, and a study of 14 pediatric patients with LCH and liver involvement showed a 100% prevalence of *BRAF*^{V600E} mutation¹¹. Vulvar manifestations have only been described in some cases series¹². They can present as erythematous plaques, eczema, ulcer or polypoid appearance, which are non-specific and sometimes it can mimic many other diseases, such as squamous cell carcinoma, malignant melanoma, herpes or some inflammatory reaction¹².

BRAF^{Δβ3-αC} were described in pancreatic, lung, ovarian, thyroid cancers and melanoma¹³, and also occur in histiocytoses. These oncogenic deletions are predicted to shorten the β 3/ α C-helix loop, which could favor dimer formation. They are resistant to the BRAF monomer inhibitors, such as vemurafenib but sensitive in vitro to BRAF dimer inhibitors and MEK inhibitors¹⁴. So far only two patients with histiocytosis harboring *BRAF*^{Δβ3-αC} have been reported with targeted therapy, and both had complete remission with either trametinib¹⁵ or cobimetinib¹⁶. Two patients of our series were treated with MEK inhibitors, with partial remission on PERCIST criteria, but no significant improvement in liver function tests and MRI cholangiopancreatography (**Figure 2**). Based on the low response rate of standard chemotherapy in liver locations, those patients may require first line treatment with MEK-inhibitors.

To conclude, *BRAF*-deletions mutations in histiocytoses seem to be associated with a specific LCH pattern with high prevalence of hepatic and vulvar involvements. These manifestations should be carefully screened in these patients. These results also comfort the hypothesis that each specific mutation in histiocytosis correlates with a specific clinical phenotype.

References

1. Emile J-F, Abla O, Fraitag S, et al. Revised classification of histiocytoses and neoplasms of the macrophage-dendritic cell lineages. *Blood*. 2016;127(22):2672-2681.
2. Goyal G, Heaney ML, Collin M, et al. Erdheim-Chester disease: consensus recommendations for evaluation, diagnosis, and treatment in the molecular era. *Blood*. 2020;135(22):1929-1945.
3. Goyal G, Tazi A, Go RS, et al. International expert consensus recommendations for the diagnosis and treatment of Langerhans cell histiocytosis in adults. *Blood*. 2022;139(17):2601-2621.
4. Cohen-Aubart F, Emile J-F, Carrat F, et al. Phenotypes and survival in Erdheim-Chester disease: Results from a 165-patient cohort. *Am J Hematol*. 2018;93(5):E114-E117.
5. Razanamahery J, Diamond EL, Cohen-Aubart F, et al. Erdheim-Chester disease with concomitant Rosai-Dorfman like lesions: a distinct entity mainly driven by MAP2K1. *Haematologica*. 2020;105(1):e5-e8.
6. Kemps PG, Picarsic J, Durham BH, et al. ALK-positive histiocytosis: a new clinicopathologic spectrum highlighting neurologic involvement and responses to ALK inhibition. *Blood*. 2022;139(2):256-280.
7. Diamond EL, Dagna L, Hyman DM, et al. Consensus guidelines for the diagnosis and clinical management of Erdheim-Chester disease. *Blood*. 2014;124(4):483-492.
8. Colomba E, Hélias-Rodzewicz Z, Von Deimling A, et al. Detection of BRAF p.V600E Mutations in Melanomas: Comparison of Four Methods Argues for Sequential Use of Immunohistochemistry and Pyrosequencing. *J Mol Diagn*. 2013;15(1):94-100.
9. Richards S, Aziz N, Bale S, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet Med*. 2015;17(5):405-424.
10. Aricò M, Girschikofsky M, Généreau T, et al. Langerhans cell histiocytosis in adults. Report from the International Registry of the Histiocyte Society. *Eur J Cancer*. 2003;39(16):2341-2348.
11. Carrere X, Pinto N, Gene Olaciregui N, et al. High prevalence of BRAFV600E in patients with cholestasis, sclerosing cholangitis or liver fibrosis secondary to Langerhans cell histiocytosis. *Pediatr Blood Cancer*. 2021;68(7):e29115.
12. Jiang W, Li L, He Y-M, Yang K-X. Langerhans cell histiocytosis of the female genital tract: a literature review with additional three case studies in China. *Arch Gynecol Obstet*. 2012;285(1):99-103.
13. Chen S-H, Zhang Y, Van Horn RD, et al. Oncogenic BRAF Deletions That Function as Homodimers and Are Sensitive to Inhibition by RAF Dimer Inhibitor LY3009120. *Cancer Discov*. 2016;6(3):300-315.
14. Lauinger M, Christen D, Klar RFU, et al. BRAF $\Delta\beta 3$ - αC in-frame deletion mutants differ in their dimerization propensity, HSP90 dependence, and druggability. *Sci Adv*. 2023;9(35):eade7486.
15. Lee LH, Gasilina A, Roychoudhury J, et al. Real-time genomic profiling of histiocytoses identifies early-kinase domain BRAF alterations while improving treatment outcomes. *JCI Insight*. 2017;2(3):e89473.
16. Diamond EL, Durham BH, Ulaner GA, et al. Efficacy of MEK inhibition in patients with histiocytic neoplasms. *Nature*. 2019;567(7749):521-524.

Table 1. Characteristics of L-group histiocytosis patients with *BRAF* in-frame deletion

Patient	Gender	Age at diagnosis (years)	<i>BRAF</i> mutation	Histiocytosis	Clinical manifestations	Treatment & outcome	Follow-up duration (months)
#1	M	78	c.1459_1473del	LCH, ECD	Sclerosing cholangitis, cystic interstitial lung disease, lytic bone lesions	None (Death from coronary heart disease)	14
#2	M	74	c.1457_1471del	LCH	Sclerosing cholangitis	UDCA (stable disease)	11
#3	M	66	c.1457_1471del	LCH	Cystic interstitial lung disease	None	29
#4	F	59	c.1457_1471del	LCH	Vulvar manifestations, cutaneous manifestations, lytic bone lesions, diabetes insipidus	Vinblastine (disease progression), cladribine (disease progression)	98
#5	F	65	c.1457_1471del	LCH	Sclerosing cholangitis, vulvar manifestations, cutaneous manifestations, diabetes insipidus	Liver Transplantation (remission)	55
#6	M	56	c.1471_1476del	ECD	Long bone osteosclerosis, pachymeningitis, perirenal infiltration	Cobimetinib (partial remission)	120
#7	F	61	c.1457_1471del	LCH	Sclerosing cholangitis, vulvar manifestations, lytic bone lesions, diabetes insipidus	UDCA (stable disease) Vinblastine (disease progression), cobimetinib (partial remission)	43
#8	F	33	c.1457_1471del	LCH	Vulvar manifestations, cutaneous manifestations, cystic interstitial lung disease, lytic bone lesions, diabetes insipidus, panhypopituitarism	Methotrexate (stable disease)	315
#9	F	56	c.1457_1471del	LCH	Sclerosing cholangitis, cystic interstitial lung disease	None	0
#10	F	45	c.1457_1471del	LCH	Vulvar manifestations, cutaneous manifestations, lytic bone lesions, diabetes insipidus, pachymeningitis	None	89
#11	M	41	c.1457_1471del	LCH	Sclerosing cholangitis, cutaneous manifestations, salivary glands infiltration	Cytarabine (skin improvement, cholangitis stability), cladribine (skin improvement, cholangitis stability), lenalinomide (skin improvement, cholangitis stability)	44
#12	M	34	c.1459_1473del	LCH	Cystic interstitial lung disease	None	3
#13	F	21	c.1457_1471del	LCH	Vulvar manifestations, cutaneous manifestations, diabetes insipidus, panhypopituitarism	Cladribine (complete remission)	148
#14	M	23	c.1457_1471del	LCH	Sclerosing cholangitis, digestive track infiltration	UDCA (stable disease), Vinblastine (partial remission)	62
#15	M	17	c.1458_1472del	LCH	Lytic bone lesions	None	42
#16	F	39	c.1458_1472del	LCH	Cystic interstitial lung disease	Vinblastine (partial remission)	2
#17	F	58	c.1457_1471del	LCH	Vulvar manifestations, panhypopituitarism	None	10
#18	F	38	c.1457_1471del	LCH	Sclerosing cholangitis, cystic interstitial lung disease	None	50
#19	F	73	c.1457_1471del	LCH	Sclerosing cholangitis, cystic interstitial lung disease, vulvar manifestations, cutaneous manifestations, diabetes insipidus	UDCA (stable disease), Vinblastine (stable disease)	77
#20	M	11	c.1457_1471del	LCH	Cystic interstitial lung disease, peri-anal manifestations, diabetes insipidus, lytic bone lesions	Vinblastine (remission and relapse)	146

M: Male; F: Female; LCH: Langherans Cell Histiocytosis; ECD: Erdheim-Chester Disease UDCA: Ursodeoxycholic acid

Figure 1. Proportion of clinical manifestations in patients with histiocytosis and *BRAF* in-frame deletions

Figure 2. Sclerosing cholangitis in Langerhans cell histiocytosis patients with *BRAF* in-frame deletion

A. Intense and diffuse hypermetabolism of the intrahepatic biliary ducts (SUVmax 9.5) on FDG PET-CT before cobimetinib onset **B.** Partial regression of intense and diffuse hypermetabolism of the intrahepatic biliary ducts (SUVmax 5.9) on FDG PET-CT six months after cobimetinib onset **C-D.** MRI cholangiopancreatography performed at onset (**C**) and after six months (**D**) of cobimetinib treatment. The main bile duct (white arrow) is normal, as is the main pancreatic duct ("empty arrow"). Numerous peripheral bile ducts appear multifocally dilated and suspended in the right (empty arrowhead) or left (arrowhead) liver. The successive examinations showed no change in the number, distribution or dilatation of intrahepatic bile ducts **E-F** Large portal tract with destructive infiltration of biliary duct by numerous mononucleated histiocytes (x50) (**E**) expressing CD1a (**F**).



