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Letter to the Editor

Validation and molecular integration of the RR6 model to predict survival after 6 months of

therapy with ruxolitinib.

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The datasets used and/or analyzed during the current study are available from the corresponding author on

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Authors have no conflict of interest to declare.

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Authors' contributions

A.M.V., G.Ca., G.Co and P.G. designed the research, interpreted the results and wrote/edited the final

manuscript; A.A., F.M., G.Ca., G.Co., M.B. and M.E. collected data; C.M., E.N. and L.S. assisted in performing the

molecular research; G.Ca and G.Co. performed statistical analysis. M.M., B.M., A.I., M.F., M.C., L.B. and F.P.

contributed to the validation analysis using the original RUXO REL-MF cohort and the critical revision of the

manuscript for important intellectual content. All authors read and approved the final draft of the manuscript.

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Main body

The development of JAK-inhibitors (JAKis) marked a revolutionary break-through in the therapeutic landscape of myelofibrosis (MF)(1, 2). Ruxolitinib (Rux) is associated with consistent responses in terms of spleen volume, symptoms, and quality of life. However, almost half of the patients lose their responses after a median of 3 years, and a minority are primarily refractory(3). In addition, Rux may present dose-limiting toxicities eventually leading to dose reduction and/or discontinuation. Most importantly, Rux failure is associated with evidence of clonal progression and dismal prognosis(4-7), with an estimated overall survival (OS) of less than 18 months(6, 8).

The impact of clinical and/or molecular variables on treatment outcomes in MF patients treated with Rux is still a matter of debate. Recently, the RUXO REL-MF study group developed a clinical prognostic model, named Response to Ruxolitinib After 6 Months (RR6), that allows for the early identification of Rux-treated MF patients with impaired survival(9). The model includes 3 predictor variables collected at baseline, 3 and 6 months, and identifies 3 risk categories with distinct OS. The RR6 model was validated in an independent cohort of 140 patients(10).

In this retrospective, single-center study, we aimed to validate the RR6 model, compare its performance with currently validated prognostic models, and explore the independent contribution of genetic factors. The study was conducted in accordance with European and Italian regulations, and was approved by the ethical committees of each institution.

The study included 105 patients with World Health Organization-defined MF who were treated with Rux at CRIMM (Florence, Italy), fully annotated for clinical and genetic variables, the latter available in 103/105 (98%) patients.

Patient characteristics at Rux start are listed in Table 1. All patients were treated with Rux for at least 6 months, with a median treatment time of 28 (range, 6-130) months. Rux dose was <40 mg daily in 74 (70%), 85 (81%), and 87 (83%) patients at baseline, 12 and 24 weeks, respectively. Transfusion need was reported in 16 (15%) patients at all timepoints and 43 (41%) at 12 and/or 24 weeks. Palpatory spleen reduction \leq 30% at 12 and 24 weeks was observed in 34 (32%) patients. A total of 44 (43%) patients harbored at least 1 high molecular risk mutation (HMR^{mt}; i.e., mutations in *ASXL1*, *EZH2*, *IDH1*, *IDH2*, *SRSF2*, or *U2AF1*)(11, 12), with 12 (12%) having \geq 2 HMR^{mt}. Mutation in RAS pathway genes (RASp^{mt}; i.e., *NRAS*, *KRAS*, *CBL*) were found in 9 (9%) patients.

After a median follow-up of 86 (range, 71-109) months, 38 (36%) patients were still on treatment. Sixty-seven (64%) discontinued Rux, with most frequent reasons for discontinuation including death (27%), resistance (15%), hematological toxicity (13%), and hematopoietic stem cell transplantation (10%). According to the RR6 model, 17 (16%), 50 (48%), and 38 (36%) patients were classified as low (LoR), intermediate (InR), and high risk (HiR), respectively. The estimated median OS from 6 months after Rux start was not reached (NR; 95%CI, 49-NR), 66 (95%CI, 34-135), and 22 (95%CI 21-35) months, respectively in the three risk categories (*P*<.0001; Figure 1). Although HiR patients had a significant higher risk of death compared to both InR (HR, 2.8; 95%CI, 1.6-4.9; *P*=.0003) and LoR (HR, 5; 95%CI, 2-12.2; *P*=.0005) patients, the latter two showed a not significantly different outcome. These findings, while overall validating the RR6 model, raise concerns regarding its capability to effectively discriminate lower risk patients. Blast transformation was reported in 0, 3 (6%), and 10 (26%) patients in the RR6 LoR, InR, and HiR categories, respectively (*P*=.0039).

Next, we investigated whether the RR6 model provided more accurate prognostic information than other currently validated, dynamic prognostic models, such as the Dynamic International Prognostic Scoring System (DIPSS). Overall, RR6 risk categories were broadly represented across the baseline DIPSS (DIPSS^{bl}) ones, especially for LoR and HiR (Supplemental Figure 1A). However, a more heterogeneous composition was observed in DIPSS^{bl} In-1R and In-2R categories, that were enriched in RR6 HiR and LoR patients, respectively. Actuarial survival curves according to DIPSS^{bl} are reported in Supplemental Figure 1B; while the DIPSS^{bl} reliably discriminated lower risk patients, the OS of HiR and In-2R patients did not differ significantly. Aimed to compare the predictive performance of the RR6 vs DIPSS^{bl} models, we computed the respective C-index, Brier score, and time-dependent area under the curve (AUC) (Figure 2A-C). Overall, the RR6 model proved to be superior at all time points. Further, we investigated how the DIPSS prognostic performance changed along Rux treatment by re-computing the score at week 24 (DIPSS^{w24}). Among 104 evaluable patients, 35 (34%) and 18 (17%) switched to a lower and higher risk category, respectively (Supplemental Figure 1C). However, the statistical performance of DIPSS^{w24} did not improve (Figure 2A-C).

Then, we investigated the contribution of genetic variables, in particular conventional cytogenetics (available in 92/105 patients), driver and additional mutations. Median time between cytogenetic/molecular studies and Rux initiation was 5.7 (range, 0.2-68.8) and 4.9 (range, 0-145.4) months, respectively. Univariate Cox proportional hazards analysis identified the following molecular signatures as being associated with inferior OS (Supplemental Table 1): unfavorable karyotype according to the conventional two-tiered cytogenetic risk model(13), $ASXLI^{mt}$, $SRSF2^{mt}$, harboring \geq 1 HMR^{mt}, and having RASp^{mt}. Upon multivariate analysis, RR6 (HiR vs InR: HR 3.1; 95% CI, 1.7-5.9; P=.0004; HiR vs LoR: HR 4.4; 95% CI, 1.7-11.1; P=.0020), unfavorable karyotype (HR 3.2; 95% CI, 1.5-6.7; P=.0019), \geq 1 HMR^{mt} (HR 2.5; 95% CI, 1.4-4.6; P=.0023), and RASp^{mt} (HR 6.1; 95% CI, 2.2-17; P=.0005) remained independent predictors of reduced OS.

Next, we evaluated the prognostic contribution of genetic features by computing the C-index, Brier score, and AUC of the RR6 after its integration with HMR^{mt} and/or RASp^{mt} (Figure 2A, D-E). The highest values for performance and accuracy were achieved by the RR6-HMR^{mt}-RASp^{mt} combination, that showed to be superior at all time points, followed by the RR6-RASp^{mt} and RR6-HMR^{mt} combinations. These findings were validated using the original RUXO REL-MF cohort. Among the 71 molecularly-annotated patients, 23 (32%) harbored an HMR^{mt}, whereas 7 (10%) had a RASp^{mt}. Median time on Rux was 28 (range, 6-93) months. Also in this validation series, the RR6-HMR^{mt}-RASp^{mt} combination had the highest values for performance (C-index and in most instances AUC) (Figure 2F). Accuracy values on the other hand were better for the triple combination than the RR6 alone, but the advantage of the triple versus double (RR6-HMR^{mt}) combination was lost, likely due to the relatively small number of patients and events, especially within the RASp^{mt} group. In addition, we confirmed the superiority of the HMR^{mt}-RASp^{mt}-integrated RR6 in a cohort of 116 transplant-age patients (≤70 years) resulting from the combination of our and the original RUXO REL-MF cohorts (Supplemental Figure 2).

To further explore the role of genetic factors, we investigated clonal evolution among RR6 risk categories. Of 54 (51%) patients with molecular data at baseline and follow-up (median time from Rux start, 22 months; range, 3-67), 22 (41%) acquired at least one mutation, with most frequent acquisitions involving *ASXL1* (6/22), *KRAS* (4/22), and *NRAS* (3/22). Notably, new mutation acquisition was enriched in RR6 HiR patients (8/17, 47%), as opposed to LoR (1/12, 8%) and InR (8/27, 30%) patients. Furthermore, acquisition of >1 mutation was observed in 5 HiR patients, compared to none of LoR and InR patients.

Treatment failure to Rux due to resistance (either primary or secondary) or intolerance is associated with adverse prognosis(6, 8). Therefore, timely identification of MF patients with no or suboptimal response to Rux still represents a major therapeutic caveat. This is even more relevant when considering new available JAKis and the plethora of novel agents in advanced clinical development.

In this study, we validated the RR6 model in a large, single-center cohort of Rux-treated MF patients with extensive clinical and molecular data. The RR6 model effectively identifies Rux-treated patients with dismal survival, providing a greater prognostic performance compared to the DIPSS. However, our data suggest that the RR6 may present inferior performance in discriminating lower risk patients, possibly due to the smaller study cohort. Most importantly, we provided compelling data supporting the role of distinct molecular signatures as additional, independent risk factors. The adverse prognostic role of HMR^{mt} is currently well defined in MF(11, 12, 14). In addition, we recently reported that RASp^{mt} are associated with adverse survival outcomes, and may predict reduced response to JAKis(7). Accordingly, the integration of both HMR^{mt} and RASp^{mt} in the RR6 model remarkably enhanced the performance of the score. We validated these findings in 71 molecularly annotated patients of the original RUXO REL-MF cohort, albeit with some limitations due to the small number. Finally, we showed that clonal evolution is more frequent in patients with RR6-defined HiR disease, thus corroborating the role of genomic instability in Rux response/resistance and disease outcome. Notably, the observation that new mutation acquisition mostly involved ASXL1, KRAS, and NRAS further underscores their significance as key biological drivers in MF.

In conclusion, our findings suggest that 1) the RR6 model effectively allows the identification of HiR patients, but suffers from inferior performance in discriminating lower risk patients; 2) integration with HMR^{mt} and RASp^{mt} improves the performance of the score; and 3) in RR6 higher risk patients, inferior survival is pathogenetically associated with clonal evolution.

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Table 1. Characteristics of patients at Rux initiation in the	whole cohort and acco	rding to the RR6 mo	del		
	All patients	RR6	RR6	RR6	
Variable	n=105	low risk	intermediate risk	high risk	P
		n=17 (16%)	n=50 (48%)	n=38 (36%)	
Clinical and demopraphics WHO 2016 diagnosis; n (%)					.7
Pre-PMF; n (%)	7 (7)	2 (1)	4 (8)	1 (3)	.,
Overt PMF; n (%)	43 (41)	7 (41)	19 (38)	17 (45)	
Post-PV/ET MF; n (%)	55 (52)	8 (47)	27 (54)	20 (53)	
Male sex; n (%)	50 (48)	7 (41)	22 (44)	21 (55)	.5
Age, years; median (range)	66 (36-88)	58 (38-77)	66 (36-82)	67 (49-88)	.1
Age >65 years; n (%)	55 (52)	6 (35)	27 (54)	22 (58)	.3
Leukocytes, x10 ⁹ /l; median (range)	12.5 (2.5-80)	11.2 (4.3-37.4)	13.2 (2.6-80)	12.1 (2.5-46.1)	0.9
Leukocytes >25x10 ⁹ /l; n (%)	25 (24)	6 (35)	10 (20)	9 (24)	.4
Hemoglobin, g/dl; median (range)	11.1 (6.5-16.7)	12.6 (10.3-14.4)	11.2 (8.1-15.2)	9.1 (6.5-16.7)	.0002
Hemoglobin <10 g/dl; n (%) RBC transfusion dependence; n (%)	38 (36) 18 (17)	0 (0)	13 (26) 0 (0)	25 (66) 18 (47)	<.0001 <.0001
Platelets, x10 ⁹ /l; median (range)	193 (38-1114)	304 (165-530)	183 (38-729)	170 (53-1114)	.0016
Platelets < 100x10 ⁹ /l; median (range)	20 (19)	0 (0)	11 (22)	9 (24)	0.1
PB blasts, %; median (range)	0 (0-10)	0 (0-2)	0 (0-8)	1 (0-10)	0.1
PB blast s ≥1%; n (%)	41 (40)	5 (29)	16 (33)	20 (53)	.1
PB blast s ≥2%; n (%)	31 (30)	2 (12)	14 (29)	15 (39)	.1
BM fibrosis grade ≥2 ; n (%)	96 (91)	15 (88)	45 (90)	36 (95)	.6
Palpable spleen below the LCM; cm (range)	15 (1-33)	13 (1-29)	15 (4-26)	16 (5-33)	.1
Constitutional symptoms; n (%)	77 (73)	14 (82)	37 (74)	26 (68)	.6
Prognostic stratification					0000
DIPSS risk stratification	2 (2)	2 (42)	4 (2)	0 (0)	.0003
Low risk; n (%)	3 (3)	2 (12)	1 (2)	0 (0)	
Intermediate-1 risk; n (%)	40 (38) 51 (49)	4 (24) 11 (65)	15 (30) 31 (62)	21 (55)	
Intermediate-2 risk; n (%) High risk; n (%)	11 (10)	0 (0)	31 (62)	9 (24) 8 (21)	
MPN drivers	11 (10)	0 (0)	3 (0)	3 (21)	
JAK2 mutated; n (%)	81 (77)	14 (82)	40 (80)	27 (71)	.5
JAK2 ^{V617F} AB; median (range); evaluable=81	71 (12-100)	58 (31-90)	77 (32-100)	72 (12-99)	.3
CALR mutated; n (%)	19 (19)	2 (13)	7 (15)	10 (28)	.3
MPL mutated; n (%)	4 (4)	2 (13)	1 (2)	1 (3)	.2
Triple negative; n (%)	2 (2)	0 (0)	2 (4)	0 (0)	.3
Myeloid neoplasm-associated genes					
ASXL1 mutated; n (%); evaluable=103	37 (36)	3 (18)	20 (41)	14 (38)	.2
CBL mutated; n (%); evaluable=100	2 (2)	0 (0)	0 (0)	2 (6)	.1
CSF3R mutated; n (%); evaluable=86	0 (0)	0 (0)	0 (0)	0 (0)	-
CUX1 mutated; n (%); evaluable=79	0 (0)	0 (0)	0 (0)	0 (0)	- 1
DNMT3A mutated; n (%); evaluable=99	3 (3)	0 (0)	3 (6)	0 (0)	.1
EZH2 mutated; n (%); evaluable=103 IDH1/2 mutated; n (%); evaluable=103	10 (10)	1 (6) 2 (12)	5 (10) 1 (2)	4 (11) 1 (3)	.8
KIT mutated; n (%); evaluable=105	0 (0)	0 (0)	0 (0)	0 (0)	. 4
KRAS mutated; n (%); evaluable=97	7 (7)	2 (13)	3 (7)	2 (6)	.7
NF- E2 mutated; n (%); evaluable=90	11 (12)	0 (0)	6 (13)	5 (17)	.3
NRAS mutated; n (%); evaluable=97	11 (11)	2 (13)	4 (9)	5 (14)	.7
PTPN1 mutated; n (%); evaluable=86	3 (3)	1 (8)	1(2)	1 (3)	.7
RUNX1 mutated; n (%); evaluable=98	6 (6)	1 (6)	2 (4)	3 (9)	.7
SETBP1 mutated; n (%); evaluable=86	1 (1)	0 (0)	1 (2)	0 (0)	.6
SF3B1 mutated; n (%); evaluable=99	5 (5)	2 (13)	1(2)	2 (6)	.3
SH2B3/LNK mutated; n (%); evaluable=99	10 (10)	2 (13)	4 (9)	4 (11)	.9
SRSF2 mutated; n (%); evaluable=103	6 (6)	1 (6)	2 (4)	3 (8)	.7
TET2 mutated; n (%); evaluable=100	25 (25)	4 (25)	9 (19)	12 (33)	.3
TP53 mutated; n (%); evaluable=98	1(1)	0 (0)	1 (2)	0 (0)	.6
U2AF1 mutated; n (%); evaluable=103	1(1)	0 (0)	0 (0)	1 (3)	.4
ZRSR2 mutated; n (%); evaluable=86 HMR mutations [†] ; n (%); evaluable=103	9 (10) 44 (43)	1 (8) 5 (29)	3 (7) 23 (47)	5 (16) 16 (43)	.4
≥2 HMR mutations; n (%); evaluable=103	12 (12)	2 (12)	5 (10)	5 (14)	.9
>1 RASp mutation [‡] ; n (%); evaluable=99	9 (9)	2 (13)	3 (6)	4 (11)	.7
Cytogenetics	3 (3)	2 (20)	5 (5)	. (11)	.,
Conventional two-tiered cytogenetic; evaluable=92					.1
Favorable karyotype; n (%)	76 (83)	13 (93)	39 (89)	24 (71)	
Unfavorable karyotype; n (%)	16 (17)	1 (7)	5 (11)	10 (29)	
Revised three-tiered cytogenetic; evaluable=92					.3
Favorable karyotype; n (%)	66 (72)	11 (79)	34 (77)	21 (62)	
Unfavorable karyotype; n (%)	21 (23)	3 (21)	9 (20)	9 (26)	
Very high risk karyotype; n (%)	5 (5)	0 (0)		4 (12)	

RR6 model					
Rux dose <40 mg daily at all time points; n (%)	69 (66)	0 (0)	32 (65)	37 (97)	<.0001
Rux dose <40 mg daily at baseline; n (%)	74 (70)	0 (0)	36 (72)	38 (100)	<.0001
Rux dose <40 mg daily at 12 weeks; n (%)	85 (81)	5 (29)	42 (84)	38 (100)	<.0001
Rux dose <40 mg daily at 24 weeks; n (%)	87 (83)	8 (47)	42 (84)	37 (97)	<.0001
RBC transfusion need all time points; n (%)	16 (15)	0 (0)	0 (0)	16 (42)	<.0001
RBC transfusion need at 12 and/or 24 weeks; n (%)	43 (41)	0 (0)	21 (42)	22 (58)	.0003
Splenomegaly reduction ≤30% at 12 and 24 weeks; n (%)	34 (32)	0 (0)	3 (6)	31 (82)	<.0001
Splenomegaly reduction <30% at 12 weeks; n (%)	46 (44)	0 (0)	13 (26)	33 (87)	<.0001
Splenomegaly reduction <30% at 24 weeks; n (%)	40 (38)	0 (0)	7 (14)	33 (87)	<.0001

Notes: †HMR mutations include pathogenic variants in any of the following genes: ASXL1, EZH2, IDH1, IDH2, SRSF2 or U2AF1; >2 HMR mutations indicates the presence of 2 or more mutations (2 or more mutations in the same gene are counted as 1). ‡RAS pathway mutations include pathogenic variants in any of the following genes: NRAS, KRAS, and CBL. A VAF of 2% was used as threshold value for somatic variants.

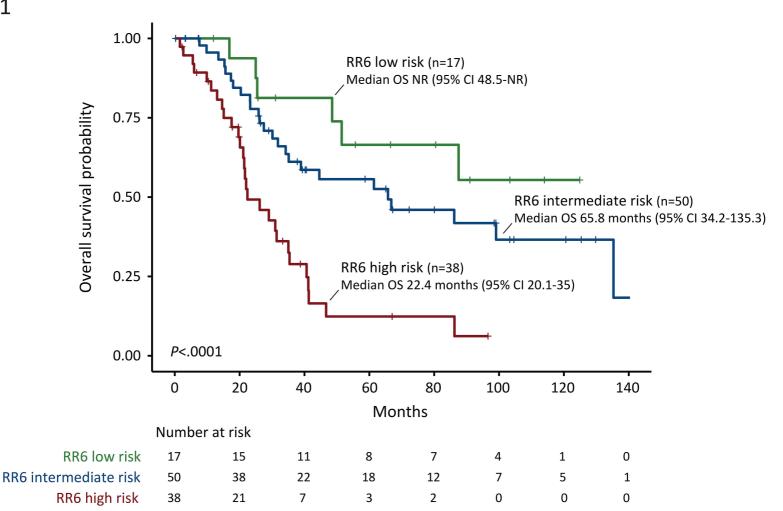
Abbreviations: AB: allele burden; BM: bone marrow; DIPSS: Dynamic International Prognostic Score System; ET: essential thrombocythemia; HMR: high molecular risk mutation; LCM: left costal margin; MF: myelofibrosis; MPN: myeloproliferative neoplasm; PB: peripheral blood; PMF=primary myelofibrosis; Pre-PMF: prefibrotic-PMF; PV: polycythemia vera; RASp: RAS pathway; RBC: red blood cell; RR6: Response to Ruxolitinib After 6 Months; Rux: ruxolitinib; VAF: variant allele frequency; WHO: world health organization.

Figure legend

Figure 1. Kaplan-Meier estimates of overall survival in ruxolitinib-treated patients according to the RR6 model. *Abbreviations*: CI: confidence interval; NR: not reached; OS: overall survival; RR6: Response to Ruxolitinib After 6 Month.

Figure 2. Prognostic performance of conventional clinical prognostic and molecularly integrated models. A. Comparison of the prognostic performance of RR6 model, DIPSS, and RR6 integration with high molecular risk signatures. B. Brier score for prediction of death measured over time for RR6 model, DIPSS^{bl} and DIPSS^{bl} and DIPSS^{w24}. C. Time-dependent AUC for prediction of death for RR6 model, DIPSS^{bl} and DIPSS^{w24}. D. Brier score for prediction of death measured over time for RR6 model and its integration with HMR^{mt} and/ore RASp^{mt}. E. Time-dependent AUC for prediction of death for RR6 model and its integration with HMR^{mt} and/ore RASp^{mt}. F. Comparison of the prognostic performance of RR6 model and its integration with high molecular risk signatures in the validation cohort. *Notes*: Asterisk and bold indicate the best values. †HMR mutations include pathogenic variants in any of the following genes: *ASXL1*, *EZH2*, *IDH1*, *IDH2*, *SRSF2* or *U2AF1*. ‡RAS pathway mutations include pathogenic variants in any of the following genes: *NRAS*, *KRAS* or *CBL*. *Abbreviations*: AUC: area under the curve; DIPSS^{bl}: Dynamic International Prognostic Scoring System at baseline; DIPSS^{w24}: Dynamic International Prognostic Scoring System at week 24; HMR^{mt}: high molecular risk mutation; RASp^{mt}: RAS pathway mutation; RR6: Response to Ruxolitinib After 6 Month.

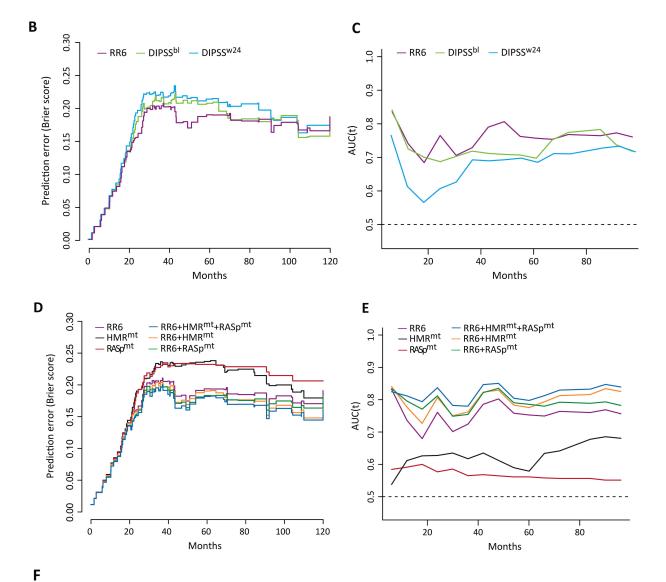
Figure 1



Α

Comparison of the prognostic performance of RR6 model, DIPSS, and RR6 integration with high molecular risk signatures	

		Events at 12 months		Events at 24 months		Events at 36 months		Events at 48 months	
	C-index	Brier score	AUC						
RR6	66.0	0.033	73.4	0.078	75.9	0.117	72.2	0.135	80.0
DIPSS	65.7	0.033	71.9	0.081	68.0	0.123	71.2	0.145	70.2
DIPSS	63.4	0.034	61.1	0.085	60.5	0.131	68.9	0.154	68.9
HMR ^{mt†}	58.2	0.035	61.0	0.084	62.6	0.129	61.6	0.154	61.0
RASp ^{mt‡}	54.1	0.034	58.9	0.082	57.5	0.127	56.3	0.153	56.3
RR6+HMR ^{mt}	68.7	0.033	77.5	0.077	80.4	0.115	76.0	0.131	82.7
RR6+RASp ^{mt}	68.6	0.032	79.3	0.075	80.9	0.112	75.2	0.129	83.3
RR6+HMR ^{mt} +RASp ^{mt}	70.5*	0.032*	80.9*	0.074*	83.5*	0.10*	77.8*	0.126*	84.8*



Comparison of the prognostic performance of RR6 model and its integration with high molecular risk signatures in the validation cohort (76 patients of the original RUXO REL-MF cohort)

		Events at 12 months		Events at 24 months		Events at 36 months		Events at 48 months	
	C-index	Brier score	AUC						
RR6	70.4	0.16*	82.2	0.19	78.4	0.20	72.5	0.19	72.5
RR6+HMR ^{mt}	74.7	0.17	92.9*	0.17*	82.0*	0.17*	75.9	0.17*	75.9
RR6+RASp ^{mt}	73.0	0.20	80.7	0.20	78.6	0.20	75.7	0.20	75.7
RR6+HMR ^{mt} +RASp ^{mt}	75.7*	0.17	91.3	0.17*	81.3	0.17*	79.3*	0.17*	79.3*

Supplemental Table 1. Results of univariate and multivariate Cox proportional hazards model of OS for RR6 model and genetic variables

	Univariat	te	Multivari	ate
	HR (95% CI)	<i>P</i> value	HR (95% CI)	P value
RR6 model				
Low risk	Referenc	е	Referen	ce
Intermediate risk	1.7 (0.7-4.3=	.2	1.4 (0.5-3.5)	.5
High risk	4.9 (2.0-12.2)	.0005	4.4 (1.7-11.1)	.0020
MPN drivers				
JAK2 mutated; n (%)	0.8 (0.4-1.4)	.4		
JAK2 ^{V617F} AB; median (range); evaluable=81	-	.1		
CALR mutated; n (%)	1.3 (0.6-2.6)	.5		
MPL mutated; n (%)	0.3 (0-2.4)	.3		
Myeloid neoplasm-associated genes				
ASXL1 mutated; n (%); evaluable=103	1.7 (1-2.9)	.0447		
CBL mutated; n (%); evaluable=100	2.9 (0.7-12.1)	.1		
CSF3R mutated; n (%); evaluable=86	-	-		
CUX1 mutated; n (%); evaluable=79	-	-		
DNMT3A mutated; n (%); evaluable=99	0.9 (0.1-6.2)	.9		
EZH2 mutated; n (%); evaluable=103	1.5 (0.6-3.4)	.4		
IDH1/2 mutated; n (%); evaluable=103	2.2 (0.5-9.3)	.3		
KIT mutated; n (%); evaluable=98	-	-		
KRAS mutated; n (%); evaluable=97	1.5 (0.6-3.7)	.4		
NF-E2 mutated; n (%); evaluable=90	0.8 (0.3-2)	.6		
NRAS mutated; n (%); evaluable=97	1.8 (0.9-3.9)	.1		
PTPN1 mutated; n (%); evaluable=86	0.9 (0.2-3.6)	.8		
RUNX1 mutated; n (%); evaluable=98	2 (0.8-5.1)	.1		
SETBP1 mutated; n (%); evaluable=86	5 (0.7-38)	.1		
SF3B1 mutated; n (%); evaluable=99	0.4 (0.1-1.6)	0.2		
SH2B3/LNK mutated; n (%); evaluable=99	2 (0.9-4.3)	.1		
SRSF2 mutated; n (%); evaluable=103	2.9 (1.2-7.4)	.0237		
TET2 mutated; n (%); evaluable=100	1.1 (0.6-2)	.7		
TP53 mutated; n (%); evaluable=98	-	-		
U2AF1 mutated; n (%); evaluable=99	1.7 (0.2-12.5)	.6		
ZRSR2 mutated; n (%); evaluable=86	1.8 (1.8-3.8)	.8		
HMR mutations [†] ; n (%); evaluable=103	2.2 (1.3-3.7)	.0048	2.5 (1.4-4.6)	.0023
≥2 HMR mutations; n (%); evaluable=103	1.7 (0.8-3.7)	.2	,	
>1 RASp mutation [‡] ; n (%); evaluable=99	3.4 (1.4-8.2)	.0064	6.1 (2.2-17)	.0005
Cytogenetics	, ,		,	
Conventional two-tiered cytogenetic; evaluable=92				
Favorable karyotype; n (%)	Referenc	e	Referen	ce
Unfavorable karyotype; n (%)	2.1 (1.1-4.2)	.0263	3.2 (1.5-6.7)	.0019
Revised three-tiered cytogenetic; evaluable=92	, ,		,	
Favorable karyotype; n (%)	Reference	e		
Unfavorable karyotype; n (%)	1.5 (0.8-2.8)	.2		
Very high risk karyotype; n (%)	2.5 (0.7-8.2)	.1		

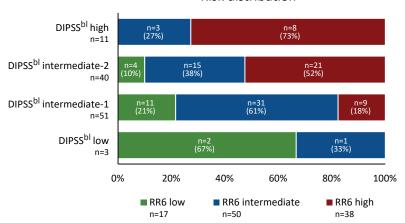
Notes: †HMR mutations include pathogenic variants in any of the following genes: ASXL1, EZH2, IDH1, IDH2, SRSF2 or U2AF1; ≥2 HMR mutations indicates the presence of 2 or more mutations (2 or more mutations in the same gene are counted as 1). ‡RAS pathway mutations include pathogenic variants in any of the following genes: NRAS, KRAS, and CBL.

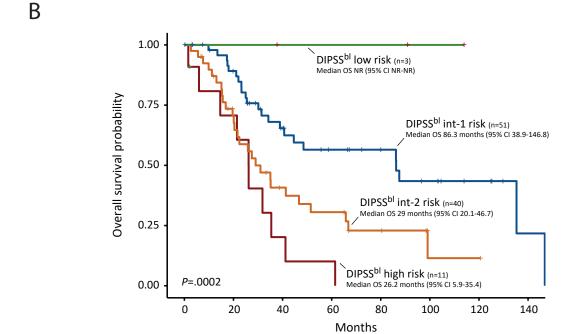
Abbreviations: AB: allele burden; HMR: high molecular risk mutation; MPN: myeloproliferative neoplasm; RASp: RAS pathway; RR6: Response to Ruxolitinib After 6 Months.

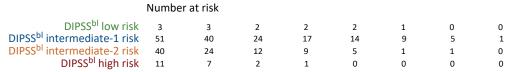
Supplemental Figure 1





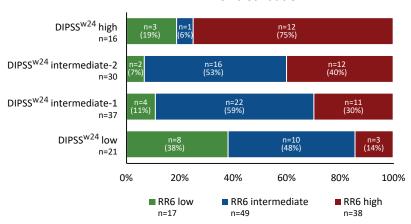






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Week 24 DIPSS vs RR6 Risk distribution



Supplemental Figure 1. Patient distribution and overall survival according to the RR6 and DIPSS models. **A.** Cross table illustrating risk distribution of patients across the RR6 model and DIPSS^{bl}. **B.** Kaplan-Meier estimates of overall survival in ruxolitinib-treated patients according to DIPSS^{bl}. **C.** Cross table illustrating risk distribution of patients across the RR6 model and DIPSS^{w24}. *Abbreviations*: CI: confidence interval; DIPSS^{bl}: Dynamic International Prognostic Scoring System at baseline; DIPSS^{w24}: Dynamic International Prognostic Scoring System at week 24; NR: not reached; OS: overall survival; RR6: Response to Ruxolitinib After 6 Month.

Supplemental Figure 2

Comparison of the prognostic performance of RR6 model and its integration with high molecular risk signatures in transplant-age patients (<70 years)										
		Events at 12	months	Events at 24	months	Events at 36	months	Events at 48	months	
	C-index	Brier score	AUC							
RR6	67.3	0.021	77.1	0.052	79.2	0.084	70.8	0.106	78.7	
HMR ^{mt†}	60.6	0.022	59.2	0.056	64.8	0.090	64.2	0115	66.6	
RASp ^{mt‡}	50.4	0.022	45.7	0.058	48.3	0.097	51.3	0.127	51.6	
RR6+HMR ^{mt}	70.8	0.021	80.0	0.052	85.2	0.082*	75.1	0.102*	82.2*	
RR6+RASp ^{mt}	67.0	0.021	78.1	0.052	78.8	0.084	69.6	0.106	77.7	
RR6+HMR ^{mt} +RASp ^{mt}	71.0*	0.021	80.6*	0.052	85.6*	0.082*	75.2*	0.102*	81.9	

Supplemental Figure 2. Comparison of the prognostic performance of RR6 model and its integration with high molecular risk signatures in a cohort of 116 transplant-age patients (≤70 years). *Notes:* Asterisk and bold indicate the best values. †HMR mutations include pathogenic variants in any of the following genes: *ASXL1*, *EZH2*, *IDH1*, *IDH2*, *SRSF2* or *U2AF1*. ‡RAS pathway mutations include pathogenic variants in any of the following genes: *NRAS*, *KRAS* or *CBL*. *Abbreviations:* AUC: area under the curve; HMR^{mt}: high molecular risk mutation; RASp^{mt}: RAS pathway mutation; RR6: Response to Ruxolitinib After 6 Month.