

# Global miRNA profiling reveals key molecules that contribute to different chronic lymphocytic leukemia incidences in Asian and Western populations

Panpan Liu,<sup>1,2\*</sup> Kefeng Wang,<sup>3,4\*</sup> Jianan Li,<sup>1,5\*</sup> Marcia A. Ogasawara,<sup>6</sup> Zhongjun Xia,<sup>1,5</sup> William G. Wierda,<sup>7</sup> Michael J. Keating,<sup>7</sup> Yiqing Li,<sup>3,8#</sup> and Peng Huang<sup>1#</sup>

<sup>1</sup>State Key Laboratory of Oncology in South China, Collaborative Innovation Center for Cancer Medicine, Sun Yat-sen University Cancer Center, Guangzhou, China; <sup>2</sup>Department of Medical Oncology, Sun Yat-sen University Cancer Center, Guangzhou, China; <sup>3</sup>Guangdong Provincial Key Laboratory of Malignant Tumor Epigenetics and Gene Regulation, Sun Yat-sen Memorial Hospital, Sun Yat-sen University, Guangzhou, China; <sup>4</sup>Department of Thoracic Surgery, Sun Yat-sen Memorial Hospital, Sun Yat-sen University, Guangzhou, China; <sup>5</sup>Department of Hematologic Oncology, Sun Yat-sen University Cancer Center, Guangzhou, China; <sup>6</sup>Department of Translational Molecular Pathology, the University of Texas MD Anderson Cancer Center, Houston, TX, USA; <sup>7</sup>Department of Leukemia, the University of Texas MD Anderson Cancer Center, Houston, Texas, USA and <sup>8</sup>Department of Hematology, Sun Yat-Sen Memorial Hospital, Sun Yat-Sen University, Guangzhou, China

\*PPL, KFW and JNL contributed equally as first authors.

#YQL and PH contributed equally as senior authors.

**Correspondence:** P. Liu  
[liupp@sysucc.org.cn](mailto:liupp@sysucc.org.cn)

Y. Li  
[liyiqing@mail.sysu.edu.cn](mailto:liyiqing@mail.sysu.edu.cn)

P. Huang  
[huangpeng@sysucc.org.cn](mailto:huangpeng@sysucc.org.cn)

**Received:** March 21, 2023.

**Accepted:** August 14, 2023.

**Early view:** August 31, 2023.

<https://doi.org/10.3324/haematol.2023.283181>

©2024 Ferrata Storti Foundation

Published under a CC BY-NC license



# Supplementary Materials for “Global microRNA profiling reveals key molecules that contribute to different CLL incidences in Asian and Western populations

## Authors

Panpan Liu,<sup>1,2 \* #</sup> Kefeng Wang,<sup>3,4 \*</sup> Jianan Li,<sup>1,5 \*</sup> Marcia A. Ogasawara,<sup>6</sup> Zhongjun Xia,<sup>1,5</sup> William G. Wierda,<sup>7</sup> Michael J. Keating,<sup>7</sup> Yiqing Li,<sup>3,8 #</sup> Peng Huang<sup>1 #</sup>

*\*PPL, KFW and JNL contributed equally as co-first authors.*

*# PPL, YQL and PH contributed equally as co-senior authors*

## Affiliations

<sup>1</sup> State Key Laboratory of Oncology in South China, Collaborative Innovation Center for Cancer Medicine, Sun Yat-sen University Cancer Center, Guangzhou, China

<sup>2</sup> Department of Medical Oncology, Sun Yat-sen University Cancer Center, Guangzhou, China

<sup>3</sup> Guangdong Provincial Key Laboratory of Malignant Tumor Epigenetics and Gene Regulation, Sun Yat-sen Memorial Hospital, Sun Yat-sen University, Guangzhou 510000, China;

<sup>4</sup> Department of Thoracic Surgery, Sun Yat-sen Memorial Hospital, Sun Yat-sen University, Guangzhou, China

<sup>5</sup> Department of Hematologic Oncology, Sun Yat-sen University Cancer Center, Guangzhou, China

<sup>6</sup> Department of Translational Molecular Pathology, The University of Texas MD Anderson Cancer Center, Houston, Texas 77030, USA

<sup>7</sup> Department of Leukemia, The University of Texas MD Anderson Cancer Center, Houston, Texas 77030, USA

<sup>8</sup> Department of Hematology, Sun Yat-Sen Memorial Hospital, Sun Yat-Sen University, Guangzhou, P. R. China

## **Supplementary Methods**

### **MicroRNA microarray assay**

The Affymetrix GeneChip microRNA 3.0 Array (Affymetrix, Santa Clara, CA, USA) analyses were performed using the standard procedures recommended by the manufacturer. This microRNA array contains probe sets for 5607 human miRNAs. The RNA labeling, microRNA array hybridization, scanning, and quantification were performed according to the Affymetrix GeneChip protocols. The CEL-files of the raw data were obtained using the Affymetrix GeneChip Command Console Software, and the data was pre-processed by Affymetrix Expression Console software 1.2 using the default analysis settings, and Robust Multi-array Average (RMA) method was used for normalization. miRNAs with values above the cutoff of 10 in all samples were chosen for data analysis. The expression values were generated and subsequently normalized using the Affymetrix Gene Chip Command Console (AGCC) Software. In the microRNA profiling, probes with detection rate less than 80% across the subjects were removed to minimize false positive results and improve data quality.

### **Quantitative real-time PCR**

Total RNA was extracted using the TRIzol reagent (Invitrogen, USA). Conversion of mRNA to cDNA was accomplished using the Reverse Transcription System Kit (Promega, Madison, WI, USA) according to the manufacturer's instructions. cDNA was used for quantitative real-time PCR using the Bio-Rad CFX96 real-time PCR detection system (Bio-Rad Laboratories, Richmond, CA, USA), using specific primers in SYBR green reactions to determine representative mRNA levels. The following primers were used: TGR-5, F: 5'-CCCAGGCTATCTTCCCAGC-3', R: 5'-GCCAGGACTGAGAGGAGCA-3'; ASB7, F: 5'-CCGCTTCAGCTCGCCATTAT-3', R: 5'-GAATGTCGATGTTGGCATTGTG-3'; CTDSPL2, F: 5'-CAACTAATGGAGCAGCTTACTCA-3', R: 5'-CGCGTGGGCTGATGA ATAAC-3'; FBX032, F: 5'-TTCAAACCTTGTCGATGTTACCC-3', R: 5'-CCAGGAAA GGATGTGACAGTGT-3'; JOSD1, F: 5'-GGGATACGCTGCAAGAGATTT-3', R: 5'-CCATGACGTTA GTGAGGGCA-3'; MED1, F: 5'-GAAGTGTTGGCTATCTCACA CC-3', R: 5'-TGTCATCCAGT AGGTCAGAAGG-3'.  $\beta$ -actin was used as a control. The All-in-One™ microRNA qPCR Kit (GeneCopoeia, Guangzhou, China) was used for microRNA expression analysis of miR-4485, miR-138, miR-181c, miR-181d, miR181a and miR-363.

## **Target gene prediction**

Target genes of miRNAs were predicted using miRWalk (<http://mirwalk.umm.uni-heidelberg.de>), TargetScan (<https://www.targetscan.org>), miRDB (<https://mirdb.org>), and miRPathDB (<https://mpd.bioinf.uni-sb.de>); A score of  $\geq 0.95$  was used as the cut-off criterion in the prediction analysis to select candidate target genes for further analysis.

## **Establishment of stable cell lines with miR-4485 low or high expression**

Lentiviral miArrest<sup>TM</sup> microRNA inhibitor vector for microRNA-4485 containing the pEZX-AM03 vector, a mCherry reporter gene, a hygromycin selectable marker, and a H1 promoter (Cat. # HmiR-AN2125-AM03, Genecopoeia, China) and miExpress<sup>TM</sup> microRNA Expression Precursor for microRNA-4485 containing the pEZX-MR03 vector, an eGFP reporter gene, a puromycin selectable marker and a CMV promoter (Cat. # HmiR1168-MR03, Genecopoeia, China) were transduced into proper host cells. Polybrene (8 mg/ml, Sigma-Aldrich) was used in lentiviral particle infection of cells. After 24 hours incubation at 37°C, medium was replaced with fresh medium to exclude polybrene. The transduced cells were then selected with puromycin (Invivogen, San Diego, CA, USA) or hygromycin (Invitrogen, Carlsbad, CA, USA) for 2-3 weeks to obtain cells with stable overexpression or low expression of miR-4485. The respective vectors containing scrambled microRNA sequences were used in parallel experiments as controls.

## **Transfection of microRNA oligonucleotides**

MicrON miR-4485-3p mimic (dsRNA oligonucleotides) (Cat. # miR10019019), micrOFF miR-4485 inhibitor (single-stranded oligonucleotides) (Cat. # miR2160621085914), micrON mimic negative control (Cat. # miR1N0000001-1-5) and micrOFF inhibitor negative control (Cat. # miR2N0000002-1-5) were acquired from RiboBio Co., Ltd (Guangzhou, China). Their sequences of oligonucleotides are as follows: micrONTM miR-4485 mimic, 5'-UAACGGCCGCGGUACCCUAA-3', 3'-AUUGCCGGCGCCAUGGGGAUU-5'; micrOFFTM miR-4485 inhibitors, 5'-UUAGGGUACCGCGGCCGUUA-3'. The microRNA mimic and microRNA inhibitor were transfected into cells using a riboFECT CP transfection kit (Cat. C10511-05, RiboBio Co., Ltd). Transfection efficiency was confirmed by fluorescent probe co-transfected.

## **Dual-luciferase reporter assay**

The 3'UTR fragments of TGR5 (also known as GPBAR1) were inserted into the luciferase reporter vector pmiR-RB-REPORT™ (RIBOBIO, Guangzhou, China) between the XhoI and NotI restriction sites. TGR5 3'UTR mutant (Figure 4D) was also constructed and confirmed by sequencing. For the luciferase reporter assay, HEK293T cells were plated at a density of  $1.5 \times 10^4$  cells/well in 96-well plates. After 24 hours, cells were transiently co-transfected with 10  $\mu$ L of miR-4485-3p mimic or Non-target Control and 15  $\mu$ L of wild-type or mutated TGR5 luciferase reporter vector, using Lipofectamine™ 2000 transfection reagent. After 48 hours, luciferase activity was measured using Dual-Glo® Luciferase Assay System (Promega, Fitchburg, USA). The relative luciferase activity was calculated as the ratio of renilla/firefly RLU normalized by the respective control reporter construct. Statistical significance was calculated by unpaired two-tailed t-test, using  $P < 0.05$  as the value for statistical significance.

#### **Establishment of stable cell lines with TGR5-high or -low expression**

TGR5 overexpression plasmid (Cat. #PPL01052-2a) and shRNA-mediated knockdown plasmids (Cat. # PPL01052-3; Sequence #1: CGTCTACTTGGCTCCCAACTT; Sequence #2: CCTCATCATCACCGCGAACCT; #3: GCATTGCCTACCACCCAAGCA; Negative control: GTTCTCCGAACGTGTCACGTT) were purchased from GenePPL Technology Co, Ltd (Nanjing, China). Lentiviral particles containing the TGR5 plasmids were produced in 293T packaging cells. For viral infection, cells were seeded into 6-well plates and grown to 70%–80% confluency, and the culture medium was replaced by fresh RPMI-1640 medium containing the desired lentiviral particles. After 72 hours, the medium was replaced with normal culture medium. Cells with stable overexpression or knockdown of TGR5 were selected with 40  $\mu$ g/mL G418 (TGR5 overexpression) or 2  $\mu$ g/mL puromycin (TGR5-shRNA knockdown) for 3 days.

#### **Cell proliferation assay**

Cell proliferation assay was performed by plating the indicated number of cells ( $1 \times 10^4$  cells/well for 12-well plates;  $1 \times 10^5$  cells/well for 6-well plates) in their respective culture media, and cell numbers were directly counted every 24 hours using an automated cell counter (Cellometer AutoT4, Nexcelom, USA). Each data point represents the mean  $\pm$  SD from three independent experiments.

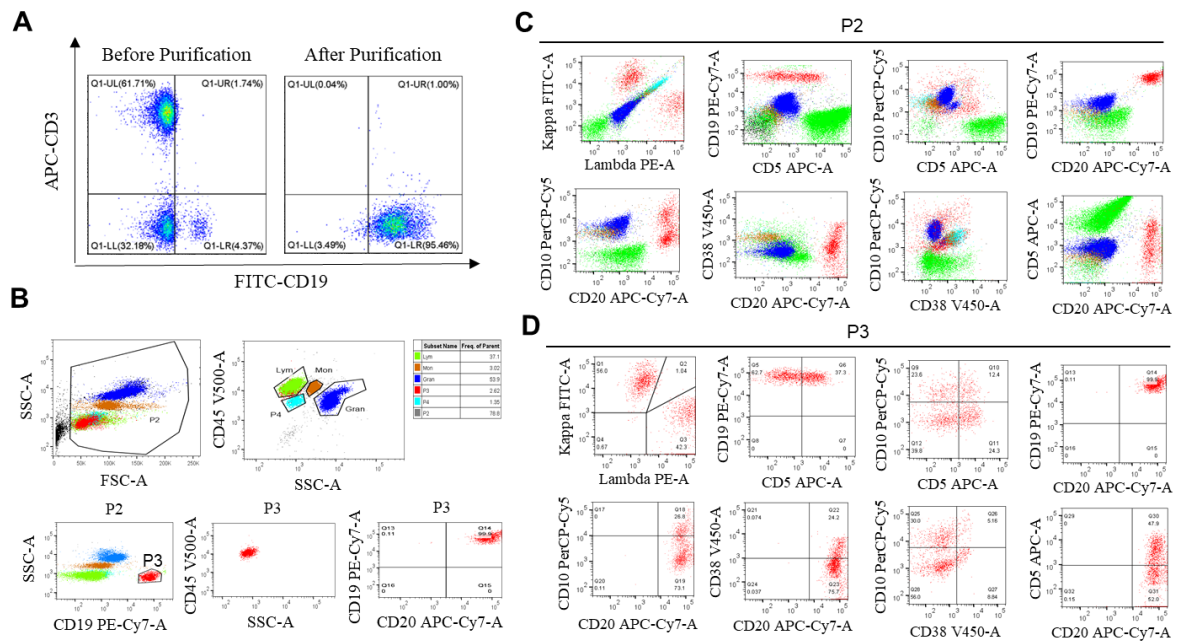
#### **Western blot analysis**

Cell samples were washed with ice-cold PBS and lysed in lysis buffer (containing RIPA and a cocktail of phosphatase inhibitors and protease inhibitors) for 15 minutes on ice. Cell debris was removed by centrifugation (12,000x g) for 15 minutes at 4°C. Protein lysates were analyzed by standard SDS-PAGE, followed by transferring to a nitrocellulose membrane. Proteins of interest were revealed by Western blotting using specific antibodies. Detail information on antibodies used in the Western blot is provided in *Online supplementary Table S5*. The ERK1/2 inhibitor SCH772984 (Cat. # HY-50846) and TGR5 agonist (Cat. # HY-14229) were purchased from MedChemExpress (Shanghai, China).

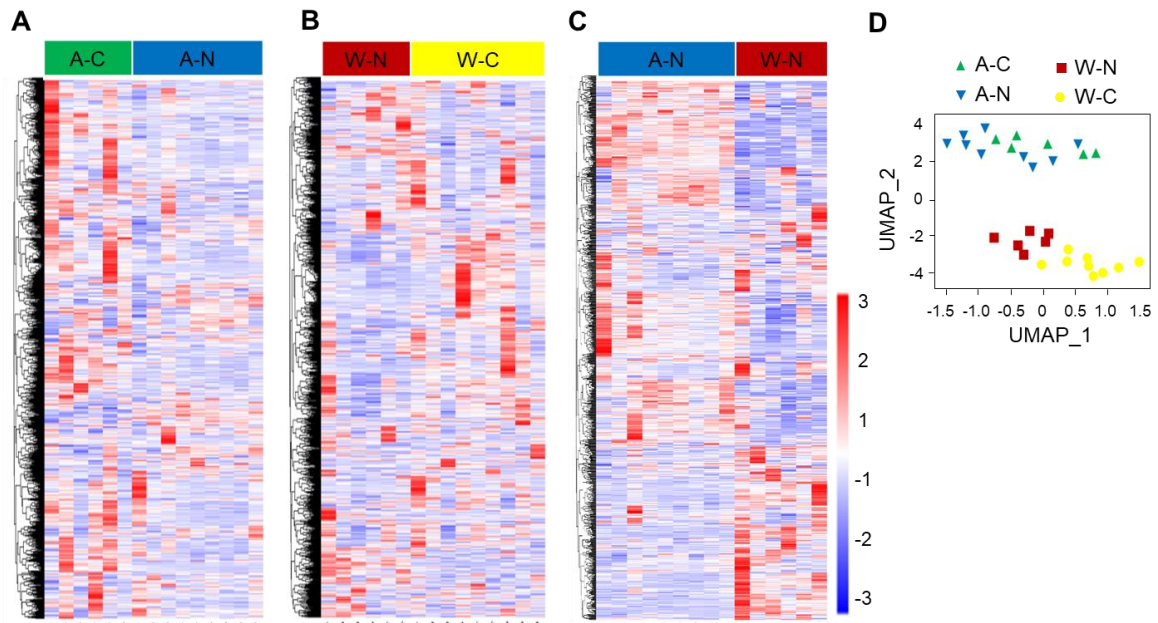
### **Data analysis and statistical tests**

T-SNE (t-distribution stochastic neighbor embedding), a nonlinear dimensionality reduction algorithm with the ability of mapping data in the high-dimensional space to low-dimensional space and retain the local characteristics of the data set, and UMAP (uniform manifold approximation and projection) capable of preserving both local and global structures of the data, were used for microRNA data dimensionality reduction and unsupervised clustering visualization. Multivariate analyses including unsupervised principal component analysis (PCA) and supervised orthogonal projections to latent structures discriminant analysis (OPLS-DA) were employed to assess the data discriminations of various groups. OPLS-DA enables the screen of differential variables by removing the unrelated differences. A 200-permutation test was performed to assess the statistical significance and prevent overfitting of the OPLS-DA model. Two-tailed Student *t*-test was used to determine potential difference between two groups, analysis of variance (ANOVA) with a Tukey's *post hoc* test was applied when more than two groups were compared. For microRNA expression data from small sample number, *p* values were adjusted using Benjamini-Hochberg false discovery rate (FDR) to calculate *q* values. A *p* or *q* value of less than 0.05 was considered statistically significant. Differentially expressed miRNAs were identified using the criteria of fold change (FC) > 2 or  $\log_2 \text{FC} > 1$  and a *q* or *p* value < 0.05. The results of t-SNE and UMAP dimensionality reduction were visualized using Sangerbox 3.0 (<http://www.sangerbox.com/tool>). Multivariate analysis was performed using SIMCA-16 software (Umetrics AB, Umeå, Sweden). Statistical analyses were performed using SPSS 25.0 software (IBM Corp, Armonk, NY, USA) or Graph Pad Prism version 9.0 (GraphPad software Inc., CA, USA).

## Supplementary Figures

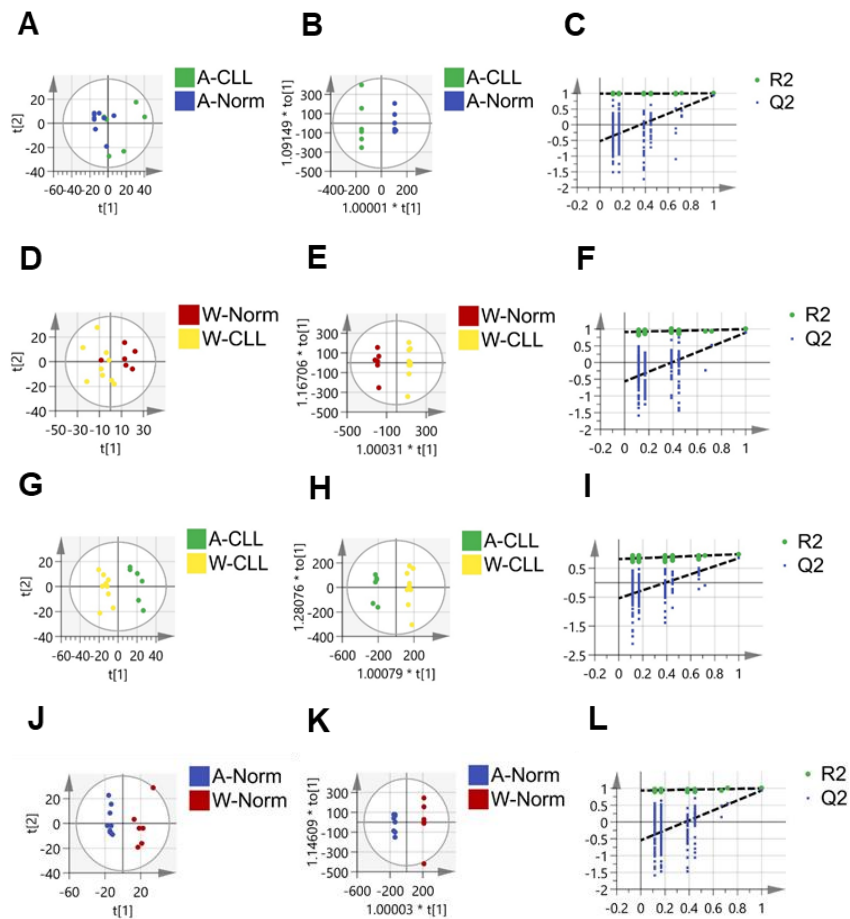


**Supplementary Figure 1. Flow cytometry analysis of B cell purity and monoclonal B cell lymphocytosis.** (A) B cells were analyzed based on their expression of CD19 and absence of T cell marker CD3. The purity of B cells was more than 95% after purification. (B-D) Multi-color flow cytometry analysis of normal peripheral blood to conform the absence for monoclonal B cell lymphocytosis. Whole blood cells were stained with various combinations of fluorochrome-conjugated antibodies as indicated, and specific sub-populations of cells were gated according to their light scattering and staining by specific antibodies. Cells were first gated on size and singularity to exclude dead cells. B-lymphocytes were gated based on CD19 expression. The presence or absence of monoclonal B cell lymphocytosis was evaluated by CD19, CD5, Kappa, and lambda immunophenotyping.

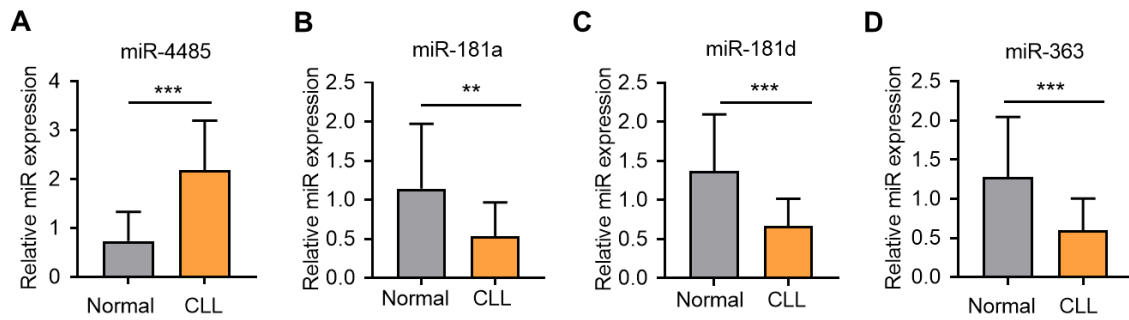


**Supplementary Figure 2. Unsupervised hierarchical clustering and UMAP visualization of microRNA expression.** (A, B, C) Unsupervised hierarchical clustering of microRNA expression by the indicated sample groups (A-C: Asian CLL, A-N: Asian normal, W-C: Western CLL, W-N: Western normal). Each row represents a microRNA, and each column represents an analyzed sample. The color code represents the relative intensity of the microRNA expression signal, with red indicating high expression and blue indicating low expression. (D) Uniform manifold approximation and projection (UMAP) plots depicting sample clustering according to microRNA expression profiles.

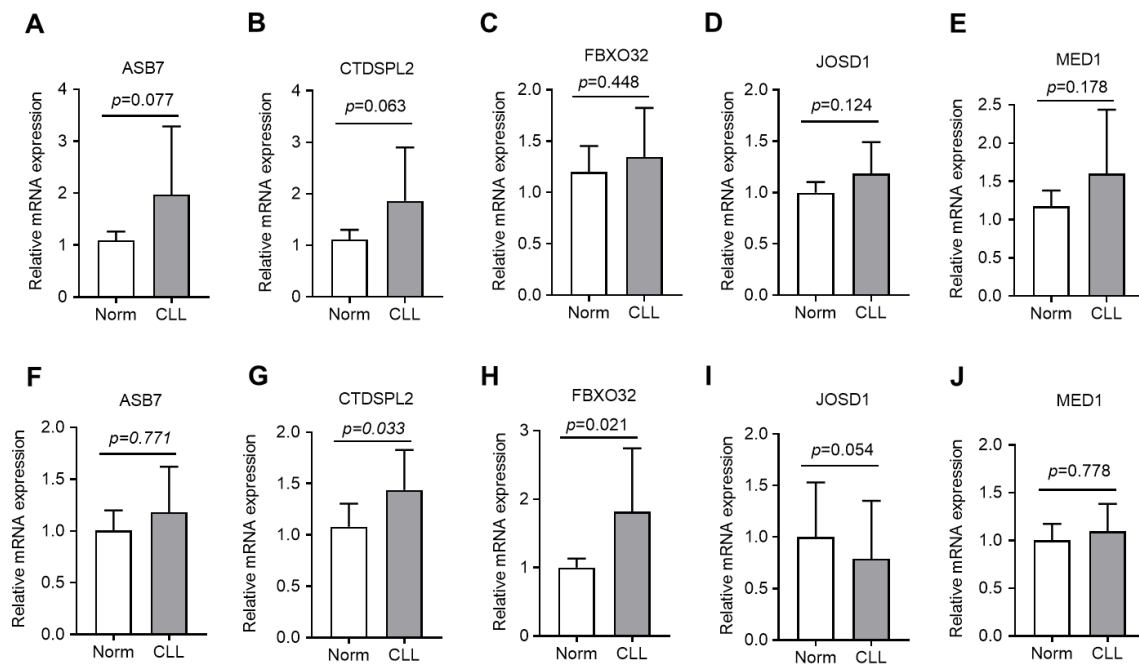




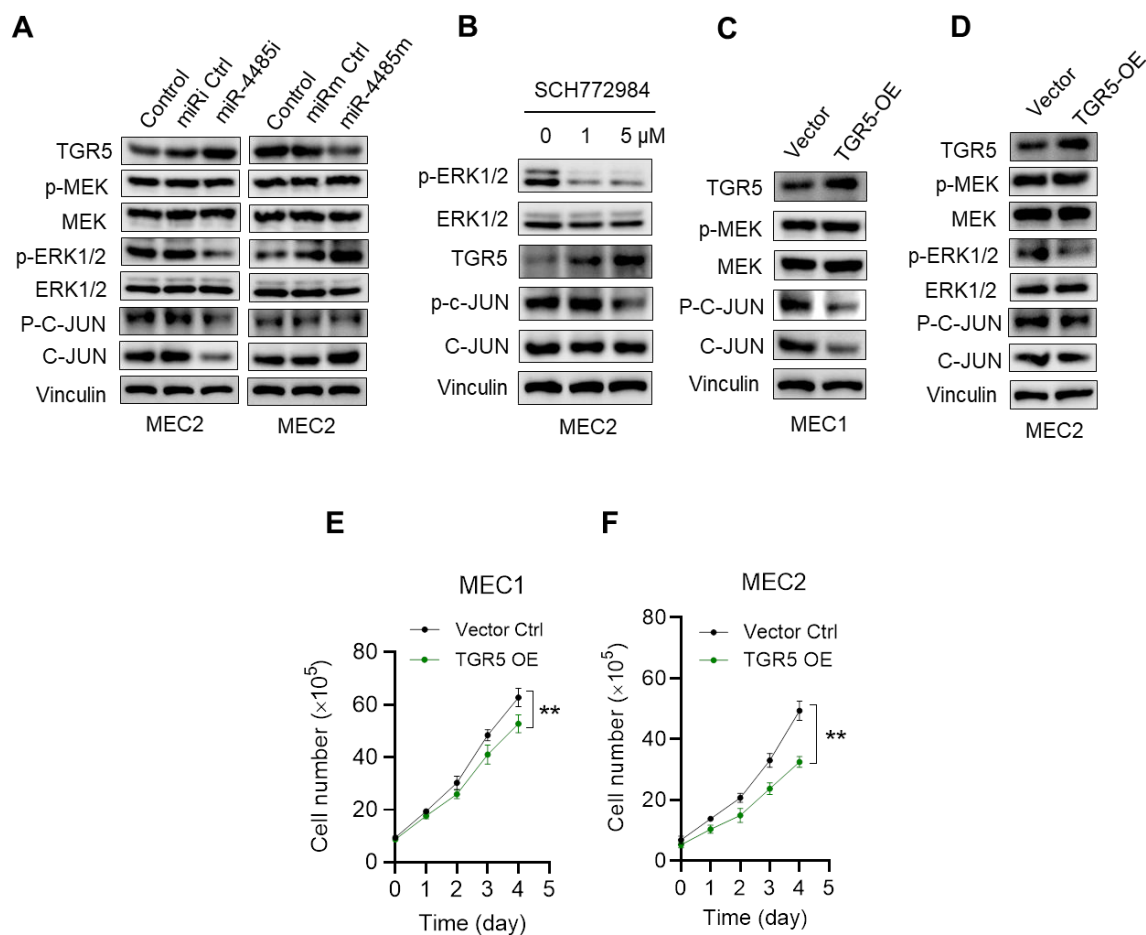
**Supplementary Figure 3. Analysis of microRNA expression in CLL cells and normal B lymphocytes from Asian and Western individuals.** (A) PCA score plots of microRNA expression in CLL cells vs normal B lymphocytes from Asian individuals.  $R^2X=0.551$ . (B) OPLS-DA score plots of microRNA expression in CLL cells vs normal B lymphocytes from Asian group.  $R^2X=0.877$ ,  $R^2Y=1.000$ ,  $Q^2=0.939$ . (C) Permutation test of OPLS-DA model in CLL cells vs normal B lymphocytes from Asian group,  $Q^2= -0.525$ . (D) PCA score plots of microRNA expression in CLL cells vs normal B lymphocytes from Western group.  $R^2X=0.550$ . (E) OPLS-DA score plots of microRNA expression in CLL cells vs normal B lymphocytes from Western group.  $R^2X=0.666$ ,  $R^2Y=0.997$ ,  $Q^2=0.894$ . (F) Permutation test of the OPLS-DA model in CLL cells vs normal B lymphocytes from Western group,  $Q^2= -0.566$ . (G) PCA score plots of microRNA expression in CLL cells from Asian and Western groups.  $R^2X=0.535$ . (H) OPLS-DA score plots of microRNA expression in CLL cells from Asian and Western groups.  $R^2X=0.500$ ,  $R^2Y=0.987$ ,  $Q^2=0.847$ . (I) Permutation test result of the OPLS-DA model in CLL cells from Asian and Western groups,  $Q^2= -0.539$ . (J) PCA score plots of microRNA expression in normal B lymphocytes from Asian and Western groups.  $R^2X=0.572$ . (K) OPLS-DA score plots of microRNA expression in normal B lymphocytes from Asian and Western groups.  $R^2X=0.832$ ,  $R^2Y=0.999$ ,  $Q^2=0.938$ . (L) Permutation test result of the OPLS-DA model in normal B lymphocytes from Asian and Western groups,  $Q^2= -0.545$ . PCA: Principal component analysis; OPLS-DA: Orthogonal projections to latent structures discriminant analysis; PT, permutation test.



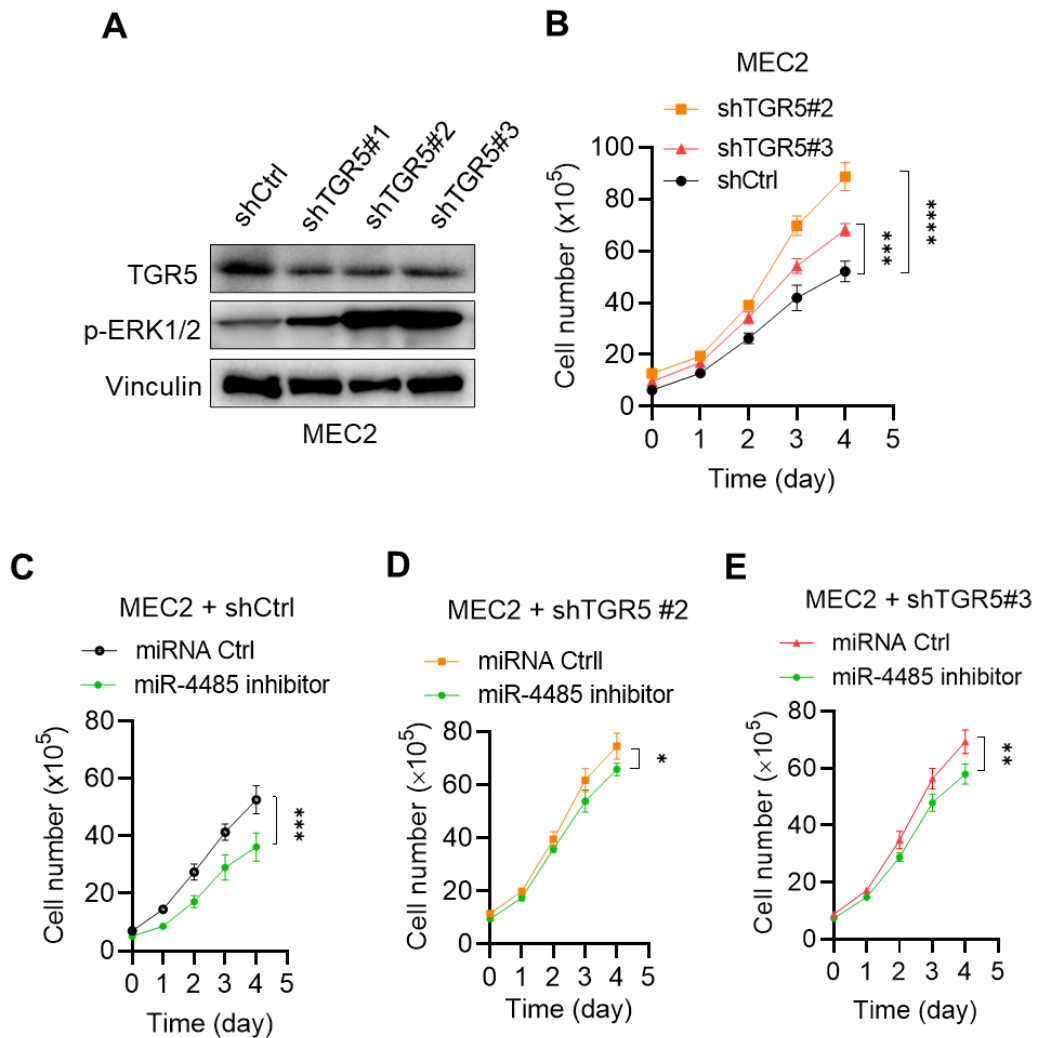
**Supplementary Figure 4.** Comparison of microRNA expression in normal B lymphocytes from healthy individuals and primary CLL cells from patients. Two microRNA expression datasets (GSE108901 and GSE66186) from Gene Expression Omnibus (GEO) database were used for analysis of expression of miR-4485 (A), miR181a (B), miR-181d (C), and miR-363 (D) in normal lymphocytes from healthy donors ( $n=20$ ) and primary CLL cells from patients ( $n=34$ ). To ensure comparability of the two datasets, the series matrix files were downloaded and merged with the inSilicoMerging R software package to adjust for batch effect, using the empirical Bayes methods (DOI: 10.1093/biostatistics/kxj037). Since GSE108901 dataset did not contain miR-181c and miR-138, these two miRNAs were not compared. \*\*,  $P < 0.01$ ; \*\*\*,  $P < 0.001$ .



**Supplementary Figure 5. Comparison of mRNA expression of *ASB7*, *CTDSPL2*, *FBXO32*, *JOSD1* and *MED1* genes in CLL cells and normal lymphocytes.** (A-E) Expression of the indicated genes in CLL cells from Asian patients (n=11) and normal lymphocytes from healthy Asian individuals (n=8). Gene expression was quantified by qRT-PCR. (F-J) Comparison of mRNA expression of the indicated genes in CLL cells from Western patients (n=45) and normal lymphocytes from healthy Western individuals (n=5). The genes expression data were from GEO database (GSE66117). None of the genes met the criteria of more than 2-fold change in expression (FC > 2) and a *P*-value of less than 0.05 to be considered significant.



**Supplementary Figure 6. Role of TGR5 in mediating miR-4485 effect on ERK1/2 and cell proliferation.** (A) MEC2 cells were transfected with miR-4485 mimic or inhibitor oligonucleotides in parallel with their respective oligo controls, and the expression of TGR5, phosphorylated MEK1/2, MEK1/2, ERK1/2, phosphorylated ERK1/2, phosphorylated-c-Jun and c-Jun was measured by Western blot analysis. (B) MEC2 cells were treated with or without the ERK inhibitor SCH772984 (1-5 $\mu$ M, 48 hours), and the indicated molecules were then analyzed by Western blot. (C) MEC1 cells were transduced with TGR5 expression vectors (TGR5-OE) or control vector, and cells were collected for Western blot analysis of indicated proteins. (D) MEC2 cells were transduced with TGR5 expression vectors (TGR5-OE) or the control vector, and the cells were collected for Western blot analysis of indicated proteins. miR-4485i: miR-4485 inhibitor; miRi Ctrl, microRNA inhibitor control; miR-4485m: miR-4485 mimic; miRm Ctrl: microRNA mimic control. (E, F) MEC1 and MEC2 cells were transfected with TGR5 overexpression vector (TGR5-OE) or the vector control, and cell numbers were counted every 24 hours.



**Supplementary Figure 7. Impact of TGR5 on ERK1/2 activation and cell proliferation in MEC2 cells.** (A) Effect of TGR5 knockdown by three shRNA (#1, #2, #3) on ERK1/2 phosphorylation in MEC2 cells, measured by Western blotting. (B) MEC2 cells were transfected with TGR5 knockdown vectors (shTGR5#2, shTGR5#3) or the negative control (shCtrl), and cell proliferation was measured by direct cell counting every 24 hours. (C-E) MEC2 cells transduced with empty vector or shTGR5 to stably knockdown TRG5. The cells with or without TGR5 knockdown were then transfected with miR-4485 inhibitor or the control oligos. Cell numbers were counted every 24 hours. Two-way ANOVA analysis was used to determine the significance of change in cell proliferation. Error bars indicate SD from three independent experiment; \*,  $P < 0.05$ ; \*\*,  $P < 0.01$ ; \*\*\*,  $P < 0.001$ ; \*\*\*\*,  $P < 0.0001$ .

## Supplementary Tables

**Supplementary Table 1. Demographic information of CLL patients whose blood samples were used in this study**

Patient #	Race	Sex/Age	WBC (x10 <sup>9</sup> /L)	Treatment status	IGVH	Del (13q)	Del (17p)	Del (11q)	TP53 mut	Trisomy 12	Trisomy 19
1	W	F/68	240.1	Untreated	M	Yes	No	No	No	No	No
2	W	M/73	157.4	Untreated	M	Yes	No	No	No	No	No
3	W	M/59	49.7	Untreated	M	Yes	No	Yes	No	No	No
4	W	F/79	263.0	Previously Treated	UM	Yes	No	No	Yes	No	No
5	W	M/81	293.0	Untreated	M	Yes	No	No	No	No	No
6	W	F/78	107.5	Untreated	M	Yes	No	No	No	No	No
7	W	M/55	38.9	Untreated	UM	No	Yes	No	No	No	No
8	W	M/60	117.2	Untreated	UM	No	No	No	No	Yes	Yes
9	W	M/74	109.3	Untreated	UM	Yes	No	No	No	No	No
10	A	F/75	262.8	Untreated	M	No	Yes	No	Yes	No	No
11	A	M/36	35.9	Untreated	UM	Yes	Yes	No	Yes	No	No
12	A	M/65	56.2	Untreated	UM	No	No	No	No	No	No
13	A	F/53	31.5	Untreated	UM	No	Yes	No	No	No	No
14	A	M/70	45.0	Untreated	M	No	No	No	No	Yes	No
15	A	M/70	200.2	Untreated	UM	Yes	No	No	No	No	No
16	A	F/57	26.0	Untreated	UM	Yes	No	No	No	No	No
17	A	F/78	12.3	Previously treated	M	No	Yes	No	Yes	No	No
18	A	M/68	32.0	Untreated	M	No	No	No	Yes	No	No
19	A	M/27	52.7	Untreated	M	No	Yes	No	Yes	No	No
20	A	M/46	41.0	Untreated	UM	No	No	No	No	No	No
21	A	M/71	37.1	Untreated	M	No	No	No	Yes	No	No
22	A	M/60	34.18	Untreated	M	No	No	No	Yes	No	No
23	A	M/52	99.84	Untreated	M	No	Yes	No	Yes	No	No
24	A	M/50	56.81	Untreated	UM	No	No	No	No	No	No
25	A	M/54	74.22	Untreated	M	No	Yes	No	No	No	No
26	A	M/61	78.23	Untreated	UM	No	Yes	No	Yes	No	No
27	A	M/64	104.46	Untreated	M	No	Yes	No	No	No	No
28	A	M/70	164.29	Untreated	UM	Yes	No	No	No	No	No
29	A	M/65	18.1	Untreated	M	No	Yes	No	Yes	No	No
30	A	F/63	70.07	Untreated	M	No	No	No	No	Yes	No
31	A	M/63	94.59	Untreated	UM	Yes	No	No	Yes	No	No

**Notes:** WBC, white blood cell counts at diagnosis; F, female; M, male; Treatment status shows if the patient was previously treated or untreated at the time of blood drawing; IGVH, immunoglobulin heavy-chain variable region; W, Western; A, Asian; M, mutated, UM, Unmutated.

**Supplementary Table 2. Differential microRNA expression in CLL cells and normal B lymphocytes from Asian individuals**

Gene ID	Fold Change (CLL/Norm)	Asian samples-															q-value %
		N1	N2	N3	N4	N5	N6	N7	N8	N9	C1	C2	C3	C4	C5	C6	
miR-1295	14.24	3.6	2.6	2.3	3.9	6.2	1.4	4.4	8.4	3.3	212.8	74.3	102.1	3.5	110.3	27.6	0.00
miR-4485	9.49	39.0	14.1	20.2	15.2	21.0	18.9	18.7	22.7	10.0	1524.8	239.4	440.4	67.9	19.9	146.1	0.00
miR-4524-star	9.49	1.3	1.7	1.5	1.5	1.9	1.4	1.4	2.0	1.6	1.5	35.8	24.8	23.4	13.3	26.8	0.00
miR-34a	3.97	9.8	17.9	30.5	13.6	21.4	16.7	18.1	15.9	16.9	100.6	8.4	335.2	52.5	225.8	29.5	0.39
miR-4521	3.85	6.3	17.5	35.1	12.1	17.5	9.7	8.3	19.1	12.2	86.2	110.7	44.7	62.0	9.5	80.1	0.00
miR-486-3p	3.21	51.9	106.9	172.7	92.3	34.5	49.3	64.9	112.2	141.6	414.9	301.5	274.4	134.0	506.0	136.6	0.00
miR-155	3.03	2267.3	2209.7	2711.0	2123.7	2827.9	2695.2	2055.6	2766.2	2636.6	3604.5	13339.1	11085.7	7526.8	5661.0	7490.1	0.00
miR-1973	3.02	74.7	47.2	52.1	52.7	41.4	40.6	49.1	53.2	38.5	362.2	201.8	198.0	94.2	75.7	102.6	0.00
miR-4524	2.83	2.1	1.7	1.7	2.2	2.3	2.6	2.3	2.5	1.6	1.9	11.1	7.8	7.3	4.8	7.1	0.00
miR-1291	2.76	1.8	5.1	9.8	6.3	4.6	4.5	3.8	3.9	7.3	19.5	7.6	18.5	10.0	13.4	14.0	0.00
miR-4304	2.68	2.1	2.6	2.2	1.3	2.5	1.8	1.5	2.3	1.7	10.0	4.5	3.8	2.8	18.2	2.4	0.00
miR-550a	2.67	1.6	6.8	4.0	8.8	5.5	6.1	6.6	8.6	7.0	23.2	22.0	13.5	10.4	16.4	9.3	0.00
miR-550a-star	2.44	3.4	6.1	4.9	8.3	7.2	8.9	6.9	12.5	7.5	14.8	29.3	17.5	11.1	23.5	11.1	0.00
miR-595	2.41	8.3	8.7	2.7	4.4	2.8	5.6	4.9	5.3	2.7	8.4	10.9	10.0	7.9	27.8	9.3	0.00
miR-331-5p	2.41	6.3	8.4	9.1	12.4	11.2	10.1	9.2	8.3	11.0	41.4	20.9	13.9	17.9	37.2	16.8	0.00
miR-21-star	2.24	3.6	7.7	4.5	6.2	7.1	9.2	10.0	7.4	3.7	29.0	15.9	11.2	10.9	9.7	13.0	0.00
miR-4793-3p	2.23	7.5	7.7	45.4	12.0	12.4	8.4	11.2	10.2	18.4	49.2	19.3	24.4	12.4	48.0	32.0	0.64
miR-34a-star	2.22	2.2	1.9	2.8	2.7	2.0	2.0	2.8	2.2	3.1	4.7	1.9	18.7	3.1	17.3	2.4	1.05
miR-574-3p	2.21	71.6	87.4	103.5	89.9	67.0	88.9	110.1	117.5	82.0	155.1	336.0	210.3	158.0	238.7	145.6	0.00
miR-486-5p	2.19	148.9	295.1	313.3	228.7	138.5	144.9	207.7	272.7	276.2	941.2	434.0	425.9	274.5	817.3	278.0	0.00
miR-4484	2.09	31.8	27.9	55.9	31.6	30.1	23.0	25.6	22.0	31.5	103.7	62.7	85.7	56.4	42.5	44.7	0.00
miR-335	2.06	13.3	14.4	10.7	11.0	15.6	19.9	12.7	14.9	12.1	7.3	49.8	38.5	35.5	57.6	17.1	0.64
miR-200a	0.42	7.7	10.3	10.1	12.8	7.3	8.2	7.4	5.5	11.0	3.0	7.6	3.1	5.5	3.3	1.9	0.66

miR-181d	0.24	42.0	67.7	43.2	60.4	54.6	44.0	46.0	44.8	44.8	5.2	12.0	25.3	24.6	2.1	33.4	0.66
miR-363	0.22	290.0	340.2	378.6	483.0	341.1	326.9	286.1	342.9	351.7	149.8	96.7	122.0	248.4	10.1	39.4	0.66
miR-451	0.19	65.2	270.3	40.9	95.4	106.3	115.1	118.9	106.5	69.9	7.8	9.6	46.2	50.6	8.1	28.5	0.00
miR-181b	0.15	301.6	623.1	602.7	971.2	640.4	614.3	559.4	577.1	582.6	99.9	15.4	358.9	216.3	7.9	510.3	4.91
miR-181c	0.13	42.4	76.9	160.2	228.6	148.5	131.1	117.7	113.9	118.9	13.5	2.1	48.2	40.2	2.7	72.7	0.66
miR-181a	0.11	1565.3	2632.8	3260.2	4689.1	2634.1	2494.3	2341.5	2016.6	2343.4	412.0	46.9	1106.1	733.6	18.2	1616.9	1.05
miR-138	0.06	141.1	112.1	97.0	106.8	79.7	76.2	99.5	84.8	102.2	55.8	2.2	7.3	3.4	2.7	5.3	0.00

**Note:** C/N: ratio of miR expression in CLL cells/miR expression in normal B lymphocytes; N1-N9, Normal lymphocytes from 9 healthy Asian individuals; C1-C6, CLL cells from 6 Asian CLL patients.



**Supplementary Table 3. Differential microRNA expression in CLL cells and normal B lymphocytes from Western individuals**

MicroRNA	Fold Change (CLL/Norm)	Western Samples														q-value (%)	
		C1	C2	C3	C4	C5	C6	C7	C8	C9	N1	N2	N3	N4	N5		N6
miR-1295	31.78	246.6	109.8	34.4	160.8	99.0	114.9	34.5	5.1	84.1	1.9	2.0	1.2	1.7	4.0	2.7	0.00
miR-451	28.27	636.0	43.9	57.0	88.9	7.3	57.7	1.8	582.9	109.7	1.5	2.0	2.1	2.5	3.4	1.4	0.00
miR-4793-3p	27.73	6.1	456.1	647.4	215.5	38.6	234.7	807.9	320.7	141.2	9.1	2.6	2.3	12.3	3.9	20.7	0.00
miR-4440	17.53	4.9	326.6	539.0	145.8	38.2	208.5	739.1	315.4	150.9	9.3	2.3	9.5	19.7	5.2	18.5	0.00
miR-210	6.27	137.4	363.5	184.6	186.7	46.1	551.2	63.1	81.2	319.8	22.4	40.4	38.6	18.3	26.4	16.7	0.00
miR-185-star	6.25	3.7	26.9	24.1	36.1	5.2	20.5	81.3	60.7	18.5	3.8	2.3	1.9	6.7	3.1	4.3	0.00
miR-148a	4.82	54.2	56.1	12.8	46.1	70.4	57.1	22.5	7.6	8.0	2.8	2.7	9.7	25.3	2.4	8.3	0.82
miR-3197	4.65	121.7	24.0	28.5	284.0	127.0	197.2	44.8	10.3	164.1	21.1	12.6	13.8	28.5	11.9	10.0	0.82
miR-4524-star	4.50	5.0	2.8	4.6	18.2	8.9	14.6	2.0	4.8	5.7	1.3	1.5	1.2	1.4	1.2	1.3	0.00
miR-3907	4.29	4.7	28.2	32.7	17.2	4.7	27.8	54.9	37.6	9.8	2.5	2.5	5.1	3.7	5.5	8.7	0.00
miR-3148	4.27	1.7	16.9	13.0	7.5	3.5	20.5	50.6	17.9	5.3	3.3	2.2	2.4	2.0	2.3	2.0	0.82
miR-4717-3p	4.17	5.4	30.8	31.2	17.2	6.8	31.6	107.7	44.7	13.0	6.0	6.3	5.6	5.4	3.4	5.6	0.00
miR-486-5p	3.53	1328.7	444.7	1014.6	508.7	263.9	428.4	264.4	2509.7	1165.6	238.5	103.8	279.0	214.2	458.6	67.7	0.82
miR-4496	3.49	20.7	22.6	29.2	38.8	21.6	18.5	94.2	68.0	40.6	6.4	11.2	12.7	3.6	29.8	8.1	0.00
miR-4462	3.47	14.8	54.2	75.1	51.2	17.7	60.0	189.0	64.0	32.9	12.3	7.2	12.2	16.0	21.0	18.6	0.00
miR-4778-5p	3.15	20.9	34.7	19.7	17.2	19.1	28.3	30.4	12.4	28.5	14.7	13.8	6.1	4.6	4.3	5.3	0.00
miR-3135b	3.07	493.8	982.9	2759.9	3461.6	3888.1	1417.2	3238.5	635.8	1878.5	954.8	607.2	591.5	664.9	434.1	258.4	0.82
miR-4486	2.93	45.4	158.4	193.8	112.4	137.9	168.1	306.4	163.9	147.8	119.4	63.8	64.1	35.1	26.8	31.0	0.00
miR-155	2.76	6030.1	7503.2	10732.7	7729.2	10618.7	8300.3	6429.5	4340.3	7667.9	2982.4	2919.8	2722.0	2403.4	2279.7	2949.3	0.00
miR-3178	2.48	349.7	449.1	801.1	552.8	383.1	706.9	977.8	656.4	369.0	196.7	314.5	187.3	169.6	182.4	328.1	0.00
miR-4507	2.47	1001.4	949.8	1859.8	1498.2	943.8	1084.2	2542.6	580.8	1307.3	1005.3	439.9	485.1	279.0	330.8	673.0	0.00
miR-4745-5p	2.47	280.8	370.0	608.3	1274.2	738.7	721.8	514.5	291.7	457.3	264.5	327.4	246.8	116.2	195.4	188.3	0.00
miR-4530	2.43	955.0	1084.4	2360.1	3292.2	1459.6	1640.1	2082.4	824.6	1099.4	1030.3	644.6	742.3	387.7	400.6	695.4	0.00
miR-4322	2.39	16.0	14.7	7.5	41.0	20.7	17.3	10.3	7.0	15.9	4.6	11.5	10.0	4.2	4.4	5.2	1.21

miR-1915	2.38	1389.4	1537.2	2586.1	5785.5	3117.3	2821.5	2351.5	1227.9	2456.2	1501.0	1223.5	1119.0	518.4	850.7	972.1	0.00
miR-4485	2.34	2450.4	4780.3	9210.7	3264.0	3585.6	1601.6	4016.6	6149.4	3047.4	2193.0	2234.4	1851.0	1326.5	922.9	1561.5	0.82
miR-3187-3p	2.31	9.9	4.5	16.0	33.3	29.5	11.7	26.3	14.4	8.5	6.8	10.7	10.0	4.3	4.0	4.6	2.91
miR-2861	2.25	973.1	1347.1	2633.1	4376.0	2384.3	2146.9	2087.0	1025.0	1864.9	1264.2	1119.3	1079.9	466.7	644.1	757.0	0.82
miR-483-5p	2.25	51.0	57.7	42.4	47.8	55.8	57.5	88.2	27.9	50.3	59.9	35.6	16.0	22.2	10.4	17.3	0.82
miR-4497	2.22	2346.2	2502.0	3705.7	4377.4	3092.0	3042.4	3536.9	2595.9	2572.7	2284.2	1395.2	1227.9	856.1	1180.5	1608.4	0.00
miR-1587	2.19	427.7	334.7	701.3	677.2	448.6	374.7	1285.7	156.0	666.0	536.5	190.7	185.2	124.3	168.3	300.1	3.82
miR-4763-3p	2.15	737.4	975.2	1647.6	1990.2	1207.6	1200.2	1833.4	558.1	1199.3	1056.2	614.7	612.9	283.9	440.9	530.8	0.82
miR-3646	2.09	6.5	10.4	24.7	11.9	8.5	11.7	15.2	6.3	13.6	4.4	5.5	6.3	4.7	4.0	7.7	0.00
miR-4505	2.08	562.1	390.7	1057.1	1370.9	705.5	691.4	1436.5	228.8	783.5	530.5	218.4	291.2	271.6	290.4	537.4	3.38
miR-3127-5p	2.07	7.5	6.9	5.7	5.3	11.7	8.9	6.0	3.4	13.3	8.1	5.6	2.1	2.9	2.8	2.1	2.64
miR-1306	2.00	7.2	8.1	5.0	4.3	5.5	4.0	14.5	8.4	5.8	4.3	4.2	3.4	2.6	3.4	2.1	0.82
miR-422a	0.48	48.3	21.7	31.6	24.5	29.8	12.3	33.2	20.4	50.9	57.7	72.9	28.3	107.4	39.2	77.9	4.26
miR-4288	0.48	8.9	4.8	2.0	7.6	3.9	3.7	5.2	5.1	3.4	6.1	10.0	7.5	12.1	11.1	12.2	1.56
miR-1275	0.48	426.7	345.7	232.0	225.2	400.7	243.8	211.3	227.6	184.5	960.0	869.5	653.2	241.7	585.8	400.1	1.56
miR-320e	0.47	109.1	59.3	95.6	119.6	144.4	58.1	117.4	62.7	311.6	304.3	208.4	181.1	234.0	236.9	189.3	2.17
miR-629	0.45	48.4	99.3	18.1	35.2	47.1	42.0	40.6	31.2	80.9	90.6	149.7	83.0	88.8	98.6	93.4	1.09
miR-221	0.43	1139.9	1088.8	648.5	1031.7	1068.8	1035.1	1406.0	989.3	899.8	2146.7	2233.9	2208.8	2591.8	2499.9	2618.9	0.00
miR-181a-star	0.42	1.8	2.8	3.0	8.8	1.6	2.5	2.8	2.8	2.4	2.4	6.3	10.6	11.5	5.5	8.4	3.07
miR-652	0.41	176.6	108.1	103.7	84.0	122.3	113.9	120.0	112.9	108.8	235.7	289.0	241.2	371.7	279.7	265.2	0.00
miR-222	0.39	1257.1	1553.1	1404.3	1882.3	1652.3	1397.4	1928.3	1713.7	1163.9	3495.5	4258.7	4280.3	2853.3	4714.4	3998.0	0.00
miR-4521	0.38	51.9	38.3	59.8	53.3	72.4	20.2	18.5	57.4	18.3	147.1	162.6	64.0	42.6	143.7	111.2	2.17
miR-484	0.38	27.1	16.0	9.1	12.5	12.3	19.5	16.7	32.2	40.0	30.2	53.8	51.6	80.5	52.0	39.3	0.00
miR-3195	0.36	31.4	25.8	48.9	36.8	22.0	20.4	61.5	47.7	16.7	131.7	167.3	66.1	44.3	105.6	65.3	0.00
miR-152	0.36	12.7	23.4	2.9	3.4	8.6	9.0	6.5	3.9	7.7	20.9	10.8	13.5	38.5	22.9	22.1	2.17
miR-4443	0.35	192.8	199.1	183.3	258.3	222.0	226.1	323.9	496.9	391.6	1207.5	571.6	1062.5	778.3	757.1	438.7	0.00
miR-378-star	0.34	5.8	6.0	3.5	8.6	3.4	3.8	7.4	8.7	26.1	11.4	27.8	16.9	27.9	13.9	24.1	1.09
miR-1973	0.30	89.7	56.5	74.7	53.1	45.7	64.3	165.4	73.0	51.1	303.6	200.7	281.4	227.0	189.3	203.0	0.00

miR-146a	0.29	225.8	354.1	288.1	126.9	205.7	493.2	541.7	1394.3	521.0	787.8	1219.3	1493.3	1912.8	990.0	1495.0	0.00
miR-720	0.25	332.6	184.5	69.2	84.7	107.4	95.5	681.9	223.8	38.5	932.9	679.5	367.5	455.6	358.2	785.0	1.09
miR-1260	0.25	6.7	2.7	3.1	2.1	3.5	5.0	7.0	2.6	4.3	5.0	26.7	11.7	7.3	18.6	54.2	0.00
miR-1280	0.19	125.5	76.4	46.9	49.8	45.3	82.8	131.4	150.4	57.2	616.7	392.1	265.0	306.5	325.9	731.0	0.00
miR-1260b	0.16	97.1	80.9	60.3	20.5	55.2	61.3	157.1	163.6	63.2	814.1	361.2	281.2	327.5	416.1	691.6	0.00
miR-363	0.16	29.4	15.3	5.2	26.8	38.4	10.4	18.6	55.5	24.4	73.3	128.8	93.8	191.6	158.1	175.8	0.00
miR-23a-star	0.15	17.2	10.3	21.4	23.3	26.2	18.2	21.9	9.8	22.7	486.6	157.3	147.4	50.7	57.2	108.0	0.00
miR-4286	0.14	26.0	4.8	5.6	12.4	6.5	4.3	7.8	27.0	3.8	94.0	77.0	28.1	80.6	29.1	83.5	0.00
miR-151b	0.13	379.0	22.5	2.8	164.7	24.3	69.1	8.4	453.6	21.5	199.2	256.1	347.7	310.7	559.6	364.8	4.33
miR-3150b-3p	0.12	2.8	3.0	2.4	3.2	3.2	1.6	2.6	2.6	2.5	25.4	24.0	30.5	10.1	22.6	20.0	0.00
miR-193b	0.11	2.9	7.6	7.6	1.4	2.3	6.3	3.2	9.6	14.5	62.6	47.8	67.2	35.1	30.1	31.1	0.00
miR-181c	0.10	3.9	3.0	2.5	2.1	3.5	3.0	20.0	3.0	4.6	17.7	63.5	40.1	28.9	93.5	28.1	0.00
miR-181a	0.08	162.7	246.4	287.8	41.9	75.2	279.5	1096.6	171.2	148.8	2730.5	2250.1	2889.1	2541.9	3323.3	1129.5	0.00
miR-151-3p	0.07	198.5	3.6	1.6	66.3	4.0	16.9	2.3	169.2	7.9	96.0	138.1	230.7	166.8	318.1	224.4	1.56
miR-27a-star	0.07	4.4	6.7	4.2	6.9	15.3	2.5	2.9	4.5	14.5	96.3	115.0	65.6	90.1	34.8	134.6	0.00
miR-181b	0.06	28.5	43.2	77.4	4.9	12.6	46.2	168.9	31.1	24.1	385.7	439.4	818.9	552.2	690.6	357.1	0.00
miR-138	0.06	2.1	2.0	1.4	1.8	2.5	1.7	1.2	1.9	16.1	74.3	107.9	26.5	6.3	19.9	133.3	0.00
miR-181d	0.43	4.04	2.20	3.16	1.42	2.96	3.58	2.77	4.80	4.22	3.67	3.29	21.32	16.72	4.19	7.52	3.01

**Note:** C/N: ratio of miR expression in CLL cells/miR expression in normal B lymphocytes; C1-C9, CLL cells from 9 Western CLL patients; N1-N6, Normal lymphocytes from 6 healthy Western individuals.

**Supplementary Table 4. Changes in genes expression in CLL cells and normal B lymphocytes from GSE66117 dataset.**

Gene name	Fold Change (CLL/Normal)	<i>P</i> Value
<i>TGR5</i>	0.437	0.025
<i>ASB7</i>	1.033	0.771
<i>CTDSPL2</i>	1.275	0.033
<i>FBXO32</i>	1.595	0.021
<i>JOSD1</i>	0.646	0.054
<i>MED1</i>	0.973	0.778

**Note:** Gene expression of more than 2-fold change ( $FC > 2.0$  or  $< 0.5$ ) with a *P* value less than 0.05 was considered statistically significant. Only TGR5 met this criterion; CLL, n=45; normal, n=5.

**Supplementary Table 5. List of antibodies used for Western blot analysis.**

<b>Antibody Name</b>	<b>Species</b>	<b>Catalog#</b>	<b>Manufacturer</b>
Phospho-p44/42 MAPK (Erk1/2) (Thr202/Tyr204) (D13.14.4E) XP® mAb	Rabbit	#4370	Cell Signaling Technology Inc.
p44/42 MAPK (Erk1/2) (137F5) mAb	Rabbit	#4695	Cell Signaling Technology Inc.
G-protein coupled bile acid receptor 1	Rabbit	YT4636	Immunoway
Bcl-2 (D17C4) mAb	Rabbit	#3498	Cell Signaling Technology Inc.
Bcl-xL Antibody	Rabbit	#2762	Cell Signaling Technology Inc.
Stat3 (D3Z2G) mAb	Rabbit	#12640	Cell Signaling Technology Inc.
Phospho-Stat3 (Tyr705) (D3A7) XP® mAb	Rabbit	#9145	Cell Signaling Technology Inc.
MEK1/2 Antibody	Rabbit	#9122	Cell Signaling Technology Inc.
Phospho-MEK1/2 (Ser217/221) (41G9) Rabbit mAb	Rabbit	#9154	Cell Signaling Technology Inc.
c-Jun (60A8) mAb	Rabbit	#9165	Cell Signaling Technology Inc.
Phospho-c-Jun-S63 pAb	Rabbit	AP0048	ABclonal
Vinculin (E1E9V) XP® mAb	Rabbit	#13901	Cell Signaling Technology Inc.
GAPDH (14C10) mAb	Rabbit	#2118	Cell Signaling Technology Inc.