

Radical surgery and venetoclax plus azacitidine in an octogenarian with acute myeloid leukemia

A 79-year-old female patient was admitted in December 2020 to our hospital due to swelling and redness of her left forearm (Figure 1A). Besides arterial hypertension and a history of sleep apnoea her medical history was unremarkable. Laboratory evaluation revealed a decreased white blood cell count (WBC) of $0.8 \times 10^9/L$ (range, $3.5-9.8 \times 10^9/L$), a hemoglobin value of 11.6 g/dL (range, 13.5–17.5 g/dL) and platelet count of $112 \times 10^9/L$ (range, $140-360 \times 10^9/L$). A differential blood cell count showed 16% myeloid blast cells. Moreover, laboratory evaluation revealed acute renal failure with a creatinine of 128 $\mu\text{mol/L}$ (range, 45–84 $\mu\text{mol/L}$) and massively elevated C-reactive protein (CRP) of 229 mg/L (range, <5 mg/L). Renal insufficiency improved rapidly and creatinine values normalized within 5 days due to the application of intravenous fluids. Bone marrow evaluation showed myeloid blast cells of 58%. Cytogenetic analysis revealed a complex karyotype ($90,XXXX,-17,-21[12]/47,XX,+8[8]/46,XX[7]$) and molecular analysis an *IDH2* R172K mutation. All other tested mutations (*CEBPA*, *IDH1*, *NPM1*, *FLT3-ITD*, *FLT3-TKD*) were unmutated. Thus, the diagnosis of acute myeloid leukemia (AML) with myelodysplasia-related abnormalities was made¹ and the patient was classified as high-risk.² The swelling and reddening on the left forearm were diagnosed as phlegmone and treatment with broad-spectrum antibiotics (clindamycin 600 mg orally, three times daily; meropenem 1 g intravenously, three times daily) as well as mold-active antifungal prophylaxis with posaconazole was started immediately. A computed tomography (CT) scan showed extensive phlegmonous-inflammatory changes of the skin and tissue. Thus, surgical intervention with rapid debridement was urgently indicated. The infection was split surgically, showing a partially avital muscle extensor carpi ulnaris without signs of necrotizing fasciitis. Besides necrosectomy, partial resection of the muscle and fascia were performed, which was covered with synthetic wound dress-

ing (Figure 1B). Histopathological analysis revealed an avital tissue of the muscle and tendon without signs of malignancy, bacteria or necrotizing fasciitis. After surgical intervention, all fingers were moveable and sensitivity was intact. Overall, two surgical wound revisions had to be performed within 5 days with additional necrosectomy, resulting in a severe lesion, which had to be covered with an autologous mesh graft from her left thigh. We started AML treatment with azacitidine (AZA) 75 mg/m² subcutaneously, days 1–5 and venetoclax (VEN) 100 mg orally (dose ramp-up), days 1–18 on day 7 after diagnosis of AML and improvement of the wound conditions. VEN/AZA were dose-reduced due to severe infection as well as concurrent antifungal prophylaxis with posaconazole. No signs of tumor lysis syndrome occurred. Besides, therapy with broad-spectrum antibiotics was continued. In the following 2 weeks, three additional surgical interventions were required for the installation and changing of a vacuum pump. Surprisingly, the wound conditions improved drastically despite AML treatment (Figure 2A). Thus, antibiotics were reduced to monotherapy with meropenem 1 g intravenously three times daily on day 13 after admittance. Overall, only two packed red blood cells were transfused during the first cycle of VEN/AZA.

First AML response assessment was performed on day 13 after the start of VEN/AZA treatment. Bone marrow cytology revealed a decline of myeloid blast cells to 11%. Thus, VEN was stopped on day 18 to allow further wound healing and ingrowing of the mesh graft. Repeated bone marrow evaluation on day 20 showed a further decline to 9% myeloid blast cells. The patient was still bicytopenic with WBC of $0.9 \times 10^9/L$ (range, $3.5-9.8 \times 10^9/L$), a hemoglobin value of 7.3 g/dL (range, 13.5–17.5 g/dL), whereas the platelet count rose spontaneously to $194 \times 10^9/L$ (range, $140-360 \times 10^9/L$). The vacuum pump was removed 23 days after installation.



Figure 1. Picture of left arm of the patient with acute myeloid leukemia. (A) Left arm at diagnosis showing an infection with edematous swelling and redness. (B) Left arm after first surgery showing the synthetic wound dressing.



Figure 2. Picture of the left arm of the patient with acute myeloid leukemia. (A) Autologous mesh graft and (B) complete wound healing roughly 5 months after diagnosis of acute myeloid leukemia.

Hematologic recovery with WBC $2.2 \times 10^9/L$, a hemoglobin value of 8.9 g/dL and platelet count of $238 \times 10^9/L$ occurred 12 days after VEN was stopped.

Thus, the patient could be discharged 43 days after admission to our hospital. Antibiotics were reduced to oral prophylaxis with ciprofloxacin 500 mg twice daily.

After discharge, the patient was followed-up routinely in our outpatient department for continuation of VEN/AZA treatment in reduced dosage (AZA 75 mg/m² subcutaneously, days 1-5; VEN 100 mg orally once daily, days 1-14, repeated every 28 days) due to concurrent antifungal prophylaxis with posaconazole. After the second cycle of VEN/AZA treatment bone marrow evaluation revealed complete remission (CR). Besides, the *IDH2* mutation was no longer detectable by digital droplet polymerase chain reaction (sensitivity of 1:10,000 for mutated to wild-type *IDH2*).³ The therapy was continued for two more cycles. Thereafter, VEN/AZA treatment was suspended for 6 weeks due to patient's wish. In the following cycles the dosage of VEN/AZA was continued as before to prevent hematologic toxicity (AZA 75 mg/m² subcutaneously, days 1-5; VEN 100 mg orally once daily, days 1-14; repeated every 28 days). Treatment with VEN/AZA was well tolerated and no grade 3 or higher toxicity occurred.

The wound improved further and finally healed after roughly 5 months (Figure 2B). A recent follow-up phone call in September 2022 revealed that the patient is feeling well roughly 20 months after start of treatment. VEN/AZA treatment is continued as stated above in a close-to-home outpatient setting and AML is still in CR.

The combined therapy of VEN and hypomethylating agents (HMA) has led to high CR and improved overall survival rates in newly diagnosed AML not eligible for intensive chemotherapy as compared to monotherapy with HMA,⁴ leading to the recent Food and Drug Administration and European Medicine Agency approval of VEN in combination with HMA or low-dose cytarabine for older adults with newly diagnosed AML.⁵ Particularly, impressive survival benefit was shown in patients with *IDH2*- or *NPM1*-mutated AML.⁶ Indeed, continued treatment with VEN/AZA resulted in a deep and durable molecular remission in our patient, who harbored an *IDH2* mutation at diagnosis.

Thus, VEN/AZA seems currently the best option for older patients with newly diagnosed AML and one of the aforementioned mutations.

One of the main obstacles in AML therapy and one of the main reasons for early death are infections due to immunosuppression of the underlying disease as well as myelotoxic effects of the therapy.⁷ We here present the first case of successful VEN/AZA treatment despite a severe infection, requiring repetitive surgical interventions.

VEN-based regimens are associated with significant myelosuppression, requiring dose adjustment.⁸ The currently recommended dose of VEN in case of concurrent treatment with strong CYP3A4 inhibitors (posaconazole) is 50 mg/daily.⁸ In our case, we decided to reduce both, the duration of VEN/AZA as well as the dose of VEN due to significant comorbidities and concurrent antifungal prophylaxis with posaconazole. Antifungal and antibiotic prophylaxis was given only in case of neutrophil count below $0.5 \times 10^9/L$. The patient tolerated the therapy very well without any further toxicities or need to use granulocyte colony stimulating factor. Currently, we continue with lower-intensity VEN/AZA treatment indefinitely.

To date, the optimal treatment duration of VEN-based lower-intensity regimens is unknown.⁸ However, patients aren't particularly enamored with the concept of treatment with parenteral VEN/AZA extending indefinitely. Thus, finding some way of using oral VEN/AZA would seem to be an urgent clinical approach.⁹ Some patients with VEN-sensitive genomics, such as *NPM1* and *IDH2* mutations, who are in deep remission might also be candidates for treatment discontinuation and active surveillance.¹⁰

In conclusion, VEN/AZA seems to be safe and feasible, even in patients with severe infections, although larger data are needed for further evaluation.

Authors

Florian Ramdohr,¹ Robert Hennings,² Astrid Monecke³ and Sabine Kayser^{1,4,5}

¹Medical Clinic and Policlinic I, Hematology and Cellular Therapy, University Hospital Leipzig, Leipzig; ²Department of Plastic, Aesthetic and Special Hand Surgery, Clinic and Polyclinic for Orthopaedics, Traumatology and Plastic Surgery, University Hospital Leipzig, Leipzig; ³Department of Pathology, University Hospital Leipzig, Leipzig; ⁴NCT Trial Center, National Center of Tumor Diseases, German Cancer Research Center (DKFZ), Heidelberg and ⁵Institute of Transfusion Medicine and Immunology, Medical Faculty Mannheim, Heidelberg University, German Red Cross Blood Service Baden-Württemberg-Hessen, Mannheim, Germany

Correspondence:

S. KAYSER - s.kayser@dkfz-heidelberg.de

<https://doi.org/10.3324/haematol.2022.282282>

Received: October 24, 2022.

Accepted: December 6, 2022.

Prepublished: December 22, 2022.

References

1. Swerdlow SH, Campo E, Harris NL. WHO classification of tumours of haematopoietic and lymphoid tissues. Revised 4th Edition. 2017.
2. Döhner H, Estey E, Grimwade D, et al. Diagnosis and management of AML in adults: 2017 ELN recommendations from an international expert panel. *Blood*. 2017;129(4):424-447.
3. Bill M, Jentzsch M, Bischof L, et al. Impact of IDH1 and IDH2 mutation detection at diagnosis and in remission in patients with AML receiving allogeneic transplantation. *Blood Adv*. 2023;7(3):436-444.
4. DiNardo CD, Jonas BA, Pullarkat V, et al. Azacitidine and venetoclax in previously untreated acute myeloid leukemia. *N Engl J Med*. 2020;383(7):617-629.
5. DiNardo CD, Tiong IS, Quaglieri A, et al. Molecular patterns of response and treatment failure after frontline venetoclax combinations in older patients with AML. *Blood*. 2020;135(11):791-803.
6. Kayser S, Levis MJ. Updates on targeted therapies for acute myeloid leukaemia. *Br J Haematol*. 2022;196(2):316-328.
7. Logan C, Koura D, Taplitz R. Updates in infection risk and management in acute leukemia. *Hematology Am Soc Hematol Educ Program*. 2020;(1):135-139.
8. Maiti A, Konopleva MY. How we incorporate venetoclax in treatment regimens for acute myeloid leukemia. *Cancer J*. 2022;28(1):2-13.
9. Levis M. By any other name... *Blood*. 2022;140(15):1657-1658.
10. Chua CC, Hammond D, Kent A, et al. Treatment free remission (TFR) after ceasing venetoclax-based therapy in responding patients with acute myeloid leukemia. *Blood Adv*. 2022;6(13):3879-3883.

©2023 Ferrata Storti Foundation

Published under a CC BY-NC license 

Disclosures

No conflicts of interest to disclose.

Contributions

FR and SK were responsible for the concept of this paper, contributed to the literature search data collection, treated the patient, analyzed and interpreted data, and wrote the manuscript; RH treated the patient and critically revised the manuscript; AM performed research and critically revised the manuscript. All authors approved the submission.

Acknowledgments

The authors acknowledge support from the University of Leipzig within the program of Open Access Publishing.

Data-sharing statement

Questions regarding data sharing should be addressed to the corresponding author.