

Prognostic value of positron emission tomography/computed tomography in transplant-eligible newly diagnosed multiple myeloma patients from CASSIOPEIA: the CASSIOPET study

¹⁸F-fluorodeoxyglucose (¹⁸F-FDG) positron emission tomography (PET)/computed tomography (CT) is a reliable imaging technique for evaluating and monitoring multiple myeloma (MM) patients with a prognostic value for progression-free survival (PFS) and overall survival.¹⁻⁵ PET/CT-positive features at diagnosis have indeed been found to correlate with poorer outcomes.^{3,6} Here, we report the first results of CASSIOPET, a companion study of CASSIOPEIA (*clinicaltrials.gov*. Identifier: NCT02541383),^{7,8} which evaluated the prognostic value of baseline PET/CT on PFS. Study design and eligibility criteria for the CASSIOPEIA trial have been previously published.⁷ Briefly, transplant-eligible patients with newly diagnosed MM (NDMM) were randomized 1:1 to receive four 28-day, pre-autologous stem cell transplant (ASCT) induction cycles and two 28-day post-ASCT consolidation treatment cycles of daratumumab plus bortezomib/thalidomide/dexamethasone (D-VTd) or bortezomib/thalidomide/dexamethasone (VTd) in CASSIOPEIA Part 1. The primary endpoint, stringent complete response, was evaluated 100 days after ASCT. In CASSIOPEIA Part 2, patients achieving a partial response (PR) or better 100 days post-ASCT underwent a second 1:1 randomization to observation or maintenance therapy with intravenous daratumumab 16 mg/kg every 8 weeks for up to 2 years.

Among patients randomized in CASSIOPEIA, those eligible for inclusion in CASSIOPET had received a PET/CT scan ≤ 6 weeks before randomization in CASSIOPEIA. Patients were excluded if they were unable to access or undergo PET/CT investigation, had uncontrolled diabetes, or had received steroids ≤ 12 hours before the PET/CT scan. All patients provided written informed consent.

The primary endpoint of the CASSIOPET study is PFS from the second CASSIOPEIA randomization. This PFS analysis for CASSIOPEIA was recently reported.⁸ CASSIOPET analyses reported here evaluate the prognostic value of PET/CT at baseline on PFS from the first CASSIOPEIA randomization, PFS differences between baseline PET-negative versus PET-positive patients in each treatment arm, and the effect of daratumumab on PET/CT negativity at post-consolidation.

PET/CT scans were performed at baseline and post-consolidation (day 100 ± 7 days] post-ASCT). All patients had fasted for ≥ 6 hours before the PET/CT scan. No dexa-

methasone was to be administered ≤ 12 hours before the PET/CT scan. Blood glucose levels were measured before ¹⁸F-FDG injection with a preferred glycemia level ≤ 150 mg/dL. No insulin was administered ≤ 2 hours before ¹⁸F-FDG injection, and no oral contrast was given. Whole-body imaging was performed 55 to 75 minutes after the ¹⁸F-FDG injection. First scout and low-dose CT data (head to feet) were obtained, followed by PET data acquisition, image reconstruction, and analysis.

Acquired imaging data were uploaded to a central electronic repository system (KEOSYS, Saint-Herblain, France) and analyzed using the IMAGYS platform. Five-point Deauville scores (range, 1-5)⁹ were applied to bone marrow (BM), bone focal lesions (FL), extramedullary disease (EMD), and paramedullary disease (PMD). Localization of the most intense ¹⁸F-FDG uptake was identified, and the maximal standardized uptake value (SUV_{max}) was calculated. Bone SUV_{max} was defined as the hottest value between BM, FL, and PMD. PET images were interpreted (blinded to patient treatment) by an independent team of nuclear-medicine physicians with extensive MM experience. PET/CT scan assessments did not include separate assessments of CT scans; thus, patients may have been PET-negative but could still display lytic lesions in the CT scan.

PET-complete response (PET-CR) was defined as an uptake of less than or equal to the mediastinal blood pool in all localizations. PET-unconfirmed CR (PET-uCR) was defined as an uptake between the mediastinal blood pool and liver. PET-PR was defined as a decrease in the number and/or activity of BM, FL, EMD, or PMD but persistence of lesions with uptake above liver activity or BM uptake above liver activity. Patients with PET-stable disease (PET-SD) had no significant modification of FL, EMD, or PMD compared with baseline. Patients with PET-progressive disease (PET-PD) had a new lesion (FL, EMD, or PMD) compared with baseline. PET/CT-positive patients were defined as patients with PET-PR and PET-SD. PET/CT-negative patients were defined as patients with PET-CR and PET-uCR. Clinical response was assessed according to International Myeloma Working Group criteria.¹⁰

The prognostic effect of including explanatory PET/CT variables on PFS was assessed using Cox regression models. Seven baseline PET/CT characteristics were

chosen based on expert knowledge: PET positivity, presence of FL, BM infiltration, PMD, EMD, FL SUV_{max} , and bone SUV_{max} .

The prognostic effect of each of the seven baseline PET characteristics was estimated using a univariable Cox model in addition to the prognostic effect of known prognostic factors: serum lactate dehydrogenase (LDH) levels, serum β_2 microglobulin concentration, cytogenetic risk, and International Staging System (ISS) disease staging.

The prognostic effect of each of the seven baseline PET characteristics was then estimated adjusting for the treatment group and revised ISS (r-ISS). These covariates were chosen based on expert knowledge without statistical covariate selection procedures due to the relatively small number of PFS events. Adjustment for treatment group accounts for the randomized design of CASSIOPEIA, of which CASSIOPET is an ancillary study. r-ISS is the current stratification score for myeloma patients and combines ISS (which includes serum β_2 microglobulin and serum albumin), cytogenetic risk, and serum LDH level into a single variable.

A final multivariable Cox model was constructed including the seven baseline PET characteristics and adjusting for treatment and r-ISS. At the time of analysis, 20 PFS events were observed in the D-VTd group and 34 in the VTd group. Thus, the third multivariable Cox model results are exploratory and should be interpreted cautiously due to the low ratio of events per variable.

Proportional hazards and log-linearity of effects were assessed. No statistically significant violations of the pro-

portional hazard assumption were detected at the customary 5% P value threshold using the Schoenfeld residuals. No violation of the log-linearity assumption was detected using P splines. The presence of multicollinearity was assessed using the variance inflation factor; no value exceeded 2 for all PET/CT characteristics.

The log-rank estimator with Kaplan–Meier representation was used to describe PFS. Baseline and post-consolidation PET/CT negativity rates were compared between treatment groups using the chi-square test, odds ratios, and 2-sided 95% confidence intervals (CI). The role of interactions between baseline PET positivity and treatment in the PFS distribution could not be assessed, as zero PFS events were observed in the D-VTd PET-negative group, leading to a hazard ratio (HR) of 0 with a non-estimable variance using classical statistical tests.

The primary results of CASSIOPEIA Part 1 have been reported (median follow-up, 18.8 months).⁷ The current analysis of CASSIOPET was performed using patient data with a median follow-up time of 29.2 months. Of 1,085 patients enrolled in CASSIOPEIA, 268 (D-VTd, $n=137$; VTd, $n=131$) had assessable baseline PET; 184 (D-VTd, $n=101$; VTd, $n=83$) patients were also PET-evaluable post-consolidation (Figure 1). Baseline characteristics of patients with assessable baseline PET were similar to those in the overall CASSIOPEIA trial (*Online Supplementary Table S1*). At baseline, 54 patients (20%) were PET-negative and 214 (80%) were PET-positive.

PFS was better for baseline PET-negative versus PET-positive patients (hazard ratio [HR]: 0.42, 95% CI: 0.18–0.97,

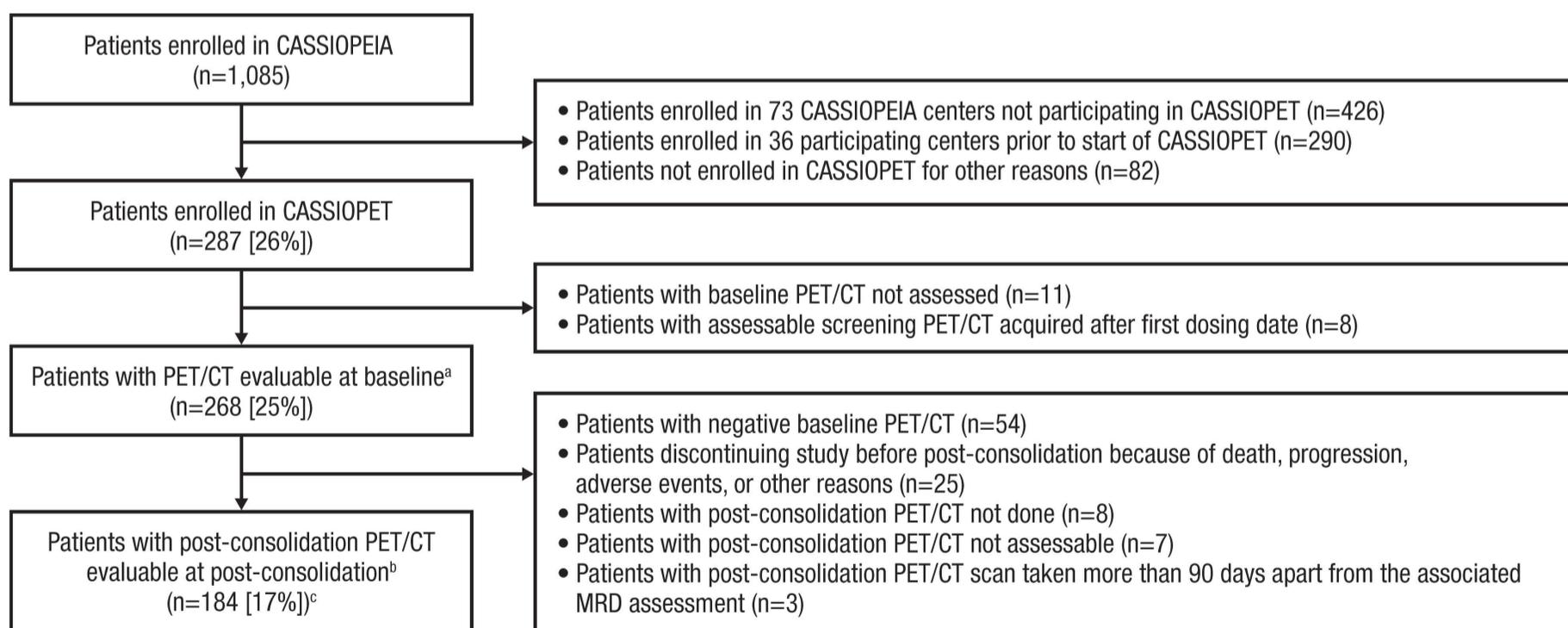
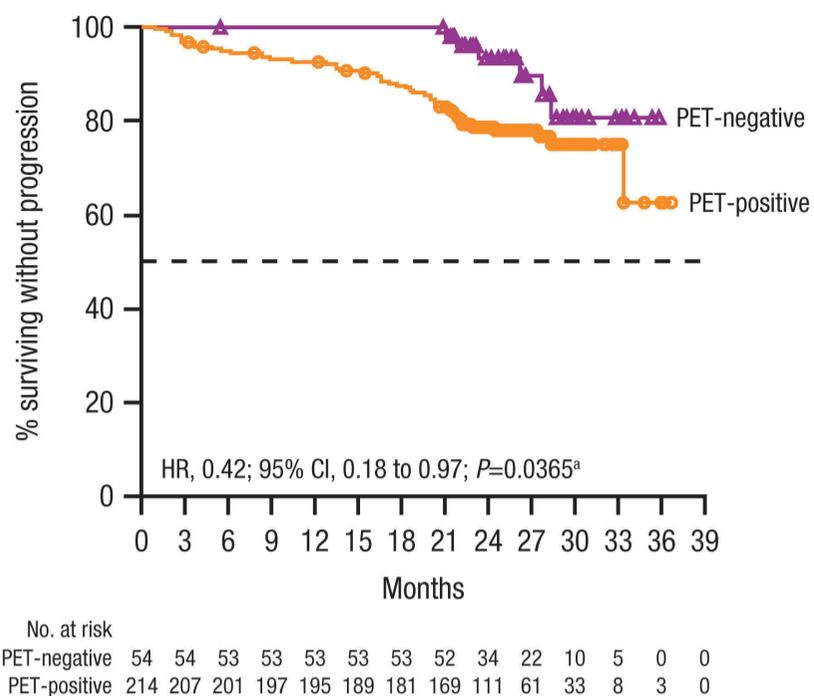


Figure 1. STROBE flow chart for CASSIOPET. ASCT: autologous stem cell transplant; MRD: minimal residual disease; PET: positron emission tomography; PET/CT: positron emission tomography/computed tomography. ^aBaseline PET-evaluable patients were defined as patients with assessable baseline PET acquired before the first dosing date. ^bPost-consolidation PET-evaluable patients included patients with assessable day 100 post-ASCT PET data and positive baseline PET but excluded patients with a date of PET/CT post-consolidation $>\pm 90$ days from the date of the day 100 MRD assessment. ^c13 patients had unevaluable baseline PET/CT but evaluable post-consolidation PET/CT and were included in the post-consolidation analysis.

A



B

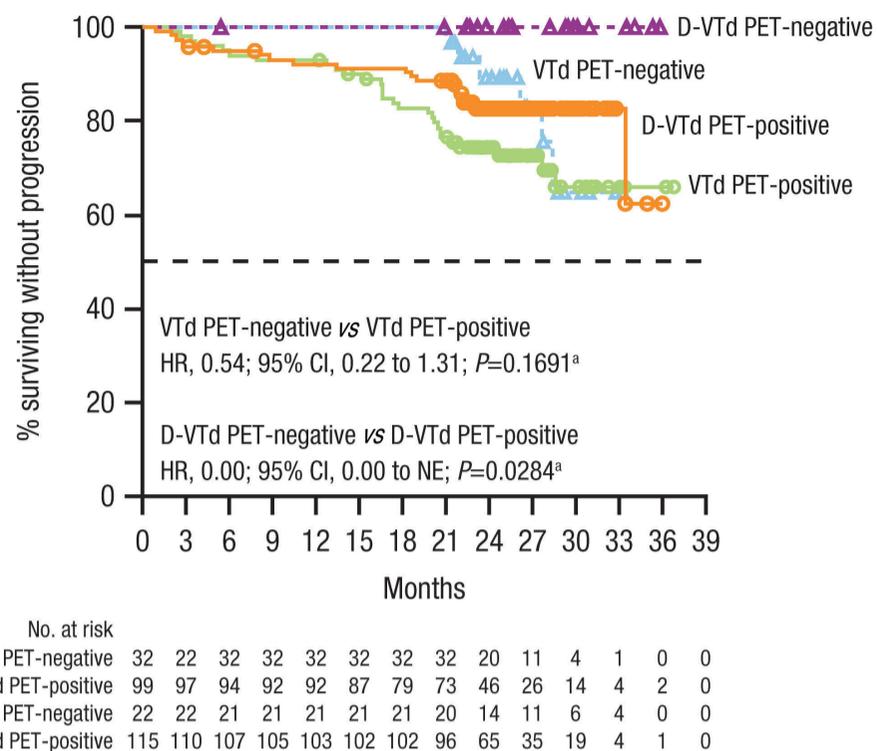


Figure 2. Progression-free survival outcomes by baseline positron emission tomography/computed tomography status in CASSIOPET.

(A) Progression-free survival (PFS) for baseline positron emission tomography (PET)-negative patients *versus* PET-positive patients and (B) PFS for baseline PET-negative patients *versus* PET-positive patients by treatment group. Baseline PET assessments were performed prior to the first dose of study drug, and PFS was based on time from first randomization. CI: confidence interval; D-VTd: daratumumab plus bortezomib/thalidomide/dexamethasone; HR: hazard ratio; NE: not estimable; VTd: bortezomib/thalidomide/dexamethasone. ^aBased on a log-rank test.

$P=0.0365$; Figure 2A). The 12- and 18-month PFS rates were higher in patients who were PET-negative (12- and 18-month rates, 100%) *versus* PET-positive (12-month rate, 93%; 18-month rate, 87%) at baseline. When stratified by treatment group, PFS was better among patients who were PET-negative *versus* PET-positive in the D-VTd arm. However, PFS was not significantly different in the VTd arm (Figure 2B). By univariable analysis, baseline PET characteristics associated with PFS were PMD ($P<0.001$), EMD ($P=0.034$), FL ($P=0.047$), FL SUV_{max} ($P=0.043$), and bone SUV_{max} ($P=0.021$). All these characteristics, except for FL, remained prognostic factors when adjusting for treatment arm and r-ISS (Table 1). A multivariable analysis including all PET/CT characteristics and adjusting for treatment arm and r-ISS showed that PMD (HR: 3.16, 95% CI: 1.60-6.28) and EMD (HR: 2.32, 95% CI: 1.04-5.19) remained independently associated with a higher risk of relapse or death (Table 1).

Of the 184 patients with post-consolidation PET measurements, 118 (64%) were assessed as PET-CR and 47 (26%) as PET-uCR (Online Supplementary Table S2). Seventeen (9%) patients were assessed as PET-PR and two (1%) as PET-SD. Overall, 165 (90%) patients were PET-negative and 19 (10%) were PET-positive. The rates of PET negativity were high and similar between the D-VTd (90%) and VTd (89%) groups.

Results of the CASSIOPET study presented here confirm that baseline PET/CT findings have a prognostic value for PFS. PFS was indeed better for baseline PET-negative

versus PET-positive NDMM patients, including patients treated with daratumumab. The presence of PMD, EMD, FL, and the FL SUV_{max} and bone SUV_{max} were associated with shorter PFS. When adjusting for treatment arm and classical NDMM (r-ISS) prognostic score, PMD and EMD had independent prognostic value. PET-CR post-consolidation rates were high and similar in both D-VTd and VTd groups.

PET/CT is negative in approximately 10-20% of symptomatic MM patients. This study shows that PET/CT negativity, even if considered as false-negative for disease detection, could be considered for its prognostic value. Rasche *et al.* demonstrated that ^{18}F -FDG PET/CT may be considered ineffective for approximately 11% of patients due to low expression of the hexokinase 2 enzyme.¹¹ However, another study of 90 NDMM patients receiving novel agents during induction therapy showed that low hexokinase 2 expression associated with PET/CT negativity correlated with relatively better prognosis *versus* PET/CT-positive patients.⁴ Baseline PET/CT-negative patients may thus represent a less aggressive subgroup of MM patients, associated with better outcomes in the setting of quadruplet therapy and ASCT.

This prospective study demonstrates PMD as an independent prognostic factor in MM. Previous prospective studies have shown the prognostic value of EMD, SUV_{max} , and FL number.^{2,5,6,12,13} However, these studies neither described nor assessed PMD as a potential prognostic biomarker. In the prospective IMAJEM study that

demonstrated the prognostic value of EMD, EMD was detected at a similar percentage (7.5%) as in CASSIOPET (5-11%), but PMD was considered as FL.² The independent prognostic value of PMD shown here is consistent with data from Rasche *et al.*, indicating the presence of large focal lesions as a strong independent poor prognosis factor in NDMM.^{14,15}

Spatial heterogeneity can limit the sensitivity of risk classification based on cytogenetics and gene expression pro-

filing because these tests are based on cells obtained from a single BM biopsy. Rasche *et al.* have shown that high-risk genomic alterations can be present in focal lesions, yet absent in other locations.¹⁴ Combined with the results of other studies,^{2,5} several PET/CT characteristics could be defined as possible high-risk biomarkers and used to define high-risk patients at the initial diagnosis of symptomatic MM.

The IMAJEM study² used background liver uptake to define

Table 1. Univariable and multivariable analyses of the prognostic value of baseline positron emission tomography (PET) characteristics on progression-free survival based on all patients with PET measurements at baseline (54/268 progression-free survival events).

Baseline characteristics	PFS events (n/N)	Univariable analysis		Analysis adjusted for treatment group and r-ISS		Multivariable analysis adjusted for treatment group, r-ISS, and all baseline PET/CT characteristics	
		HR (95% CI)	P value	HR (95% CI)	P value	HR (95% CI)	P value
PET status							
Positive	48/214	1.00	0.037	1.00	0.039	1.00	0.372
Negative	6/54	0.42 (0.18-0.97)		0.41 (0.17-0.95)		0.55 (0.15-2.04)	
Presence of FL							
No	12/88	1.00	0.047	1.00	0.051	1.00	0.753
Yes	42/180	1.90 (1.00-3.60)		1.90 (1.00-3.62)		0.84 (0.29-2.44)	
Presence of diffuse BM infiltration ^a			0.220		0.234		0.922
No	24/139	1.00		1.00		1.00	
Yes	30/129	1.40 (0.82-2.39)		1.39 (0.81-2.39)		1.03 (0.54-1.96)	
Presence of PMD			<0.001		<0.0001		0.001
No	36/221	1.00		1.00		1.00	
Yes	18/47	2.81 (1.59-4.98)		3.82 (2.11-6.92)		3.16 (1.60-6.28)	
Presence of EMD			0.034		0.012		0.041
No	46/247	1.00		1.00		1.00	
Yes	8/21	2.21 (1.04-4.69)		2.68 (1.24-5.77)		2.32 (1.04-5.19)	
FL hottest SUV _{max} ^{b,c}		1.03 (1.00-1.06)	0.043	1.06 (1.02-1.10)	0.002	0.96 (0.85-1.08)	0.479
Bone SUV _{max} ^c		1.04 (1.01-1.07)	0.021	1.06 (1.03-1.10)	<0.001	1.07 (0.96-1.19)	0.223
LDH							
<Upper limit	24/155	1.00	0.017	— ^d	—	—	—
≥Upper limit	28/103	1.92 (1.11-3.31)		— ^d		—	
Cytogenetic risk							
Standard	41/219	1.00	0.158	— ^d	—	—	—
High	13/49	1.56 (0.84-2.92)		— ^d		—	
Serum β ₂ microglobulin			0.009		—		—
<3.5 mg/L	27/167	1.00		— ^d		—	
3.5-5.4 mg/L	13/62	1.43 (0.74-2.78)		— ^d		—	
>5.4 mg/L	14/39	2.68 (1.40-5.12)		— ^d		—	
ISS stage			0.010		—		—
I	18/118	1.00		—		—	
II	22/111	1.34 (0.72-2.50)		— ^d		—	
III	14/39	2.80 (1.39-5.65)		— ^d		—	

BM: bone marrow; CI: confidence interval; EMD: extramedullary disease; FL: focal lesion; HR: hazard ratio; ISS: IMWG International Staging System; IMWG: International Myeloma Working Group; LDH: lactate dehydrogenase; PET: positron emission tomography; PFS: progression-free survival; PMD: paramedullary disease; r-ISS: IMWG revised International Staging System; SUV_{max}: maximum standardized uptake value. ^aDiffuse BM infiltration is considered to be present if visual analysis (Deauville scale) of BM uptake indicates the residual uptake to be > liver activity (4) or >> liver activity (5); otherwise, the diffuse BM infiltration is considered to be absent. ^bImputed to 1 for patients with no presence of FL. ^cHighest result among FL hottest SUV_{max}, BM uptake SUV_{max}, PMD hottest SUV_{max}. Imputed FL hottest SUV_{max} to 1 for patients with no presence of FL. ^dCovariates not included in the adjusted analysis.

PET/CT negativity, similar to the CASSIOPET study, and was recommended in the recent standardization by Zagni *et al.*⁵ Regardless of the differing efficacies and regimens, both studies support the prognostic value of PET/CT.

In conclusion, baseline PET/CT findings appear to have a prognostic value for PFS. Longer follow-up in CASSIOPEIA Part 2 will provide additional insight.

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Contributions

All authors contributed to the study design, study execution, data analysis, and manuscript writing. All authors provided a full review of the manuscript and are fully responsible for all content and editorial decisions, were involved in all stages of manuscript development, and have approved the final version.

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Data-sharing statement

The data sharing policy of Janssen Pharmaceutical Companies of Johnson & Johnson is available at <https://www.janssen.com/clinical-trials/transparency>. As noted on this site, requests for access to the study data can be submitted through Yale Open Data Access (YODA) Project site at <http://yoda.yale.edu>.

References

- Jamet B, Bailly C, Carlier T, et al. Interest of Pet imaging in multiple myeloma. *Front Med (Lausanne)*. 2019;6:69.
- Moreau P, Attal M, Caillot D, et al. Prospective evaluation of magnetic resonance imaging and [¹⁸F]fluorodeoxyglucose positron emission tomography-computed tomography at diagnosis and before maintenance therapy in symptomatic patients with multiple myeloma included in the IFM/DFCI 2009 trial: results of the IMAJEM study. *J Clin Oncol*. 2017;35(25):2911-2918.
- Cavo M, Terpos E, Nanni C, et al. Role of ¹⁸F-FDG PET/CT in the diagnosis and management of multiple myeloma and other plasma cell disorders: a consensus statement by the International Myeloma Working Group. *Lancet Oncol*. 2017;18(4):e206-e217.
- Abe Y, Ikeda S, Kitadate A, et al. Low hexokinase-2 expression-associated false-negative ¹⁸F-FDG PET/CT as a potential prognostic predictor in patients with multiple myeloma. *Eur J Nucl Med Mol Imaging*. 2019;46(6):1345-1350.
- Zamagni E, Nanni C, Dozza L, et al. Standardization of ¹⁸F-FDG-PET/CT according to Deauville criteria for metabolic complete response definition in newly diagnosed multiple myeloma. *J Clin Oncol*. 2021;39(2):116-125.
- Zamagni E, Patriarca F, Nanni C, et al. Prognostic relevance of ¹⁸-F FDG PET/CT in newly diagnosed multiple myeloma patients treated with up-front autologous transplantation. *Blood*. 2011;118(23):5989-5995.
- Moreau P, Attal M, Hulin C, et al. Bortezomib, thalidomide, and dexamethasone with or without daratumumab before and after autologous stem-cell transplantation for newly diagnosed multiple myeloma (CASSIOPEIA): a randomised, open-label, phase 3 study. *Lancet*. 2019;394(10192):29-38.
- Moreau P, Hulin C, Perrot A, et al. Maintenance with daratumumab or observation following treatment with bortezomib, thalidomide, and dexamethasone with or without daratumumab and autologous stem-cell transplant in patients with newly diagnosed multiple myeloma (CASSIOPEIA): an open-label, randomised, phase 3 trial. *Lancet Oncol*. 2021;22(10):1378-1390.
- Nanni C, Zamagni E, Versari A, et al. Image interpretation criteria for FDG PET/CT in multiple myeloma: a new proposal from an Italian expert panel. *IMPeTUs (Italian Myeloma criteria for PET USE)*. *Eur J Nucl Med Mol Imaging*. 2016;43(3):414-421.
- Kumar S, Paiva B, Anderson KC, et al. International Myeloma Working Group consensus criteria for response and minimal residual disease assessment in multiple myeloma. *Lancet Oncol*. 2016;17(8):e328-e346.
- Rasche L, Angtuaco E, McDonald JE, et al. Low expression of hexokinase-2 is associated with false-negative FDG-positron emission tomography in multiple myeloma. *Blood*. 2017;130(1):30-34.
- Bartel TB, Haessler J, Brown TLY, et al. F¹⁸-fluorodeoxyglucose positron emission tomography in the context of other imaging techniques and prognostic factors in multiple myeloma. *Blood*. 2009;114(10):2068-2076.
- Michaud-Robert AV, Zamagni E, Carlier T, et al. Glucose metabolism quantified by SUV_{max} on baseline FDG-PET/CT predicts survival in newly diagnosed multiple myeloma patients: combined harmonized analysis of two prospective phase III trials. *Cancers (Basel)*. 2020;12(9):2532.
- Rasche L, Chavan SS, Stephens OW, et al. Spatial genomic heterogeneity in multiple myeloma revealed by multi-region sequencing. *Nat Commun*. 2017;8(1):268.
- Rasche L, Angtuaco EJ, Alpe TL, et al. The presence of large focal lesions is a strong independent prognostic factor in multiple myeloma. *Blood*. 2018;132(1):59-66.