

Biallelic mutations in the SARS2 gene presenting as congenital sideroblastic anemia

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doi:10.3324/haematol.2021.279138

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Supplemental tables

Table S1

	Variables	Proband	Normal reference range
Blood biochemistry test	Urea (mmol/L)	34.4	1.6-6.5
	Creatinin (μmol/L)	171	26-42
	Uric acid (μmol/L)	790	120-320
	Sodium (mmol/L)	140	136-146
	Potassium (mmol/L)	5	3.1-4.7
	Calcium (mmol/L)	2.53	2.20-2.70
	Phosphate (mmol/L)	1.31	1.3-1.85
	Haptoglobin (g/L)	<0.08	0.47-1.25
	Total bilirubin (μmol/L)	10	0-17
	Lactate dehydrogenase (IU/L)	267	125-243
Blood Count	Hemoglobin (g/dL)	9	11.5-13.5
	Hematocrit (%)	25.7	34-40
	Mean Corpuscular Volume (fL)	80	75-81
	White Blood Cells (G/L)	9.4	6.0-17.0
	Neutrophils (G/L)	5.8	1.5-8.5
	Platelets (G/L)	170	175-500
	Reticulocytes (G/L)	63	
	Schizocytes	<1%	
Iron status	Iron (μmol/L)	20	7.0-30
	Transferrin (g/L)	1.88	1.9-3.02
	Total Iron Binding Capacity (μmol/L)	47	45-80
	Transferrin saturation (%)	43	16-35
	Ferritin (μg/L)	1621	15-80

Supplemental Table 1: Laboratory data of the proband at diagnosis

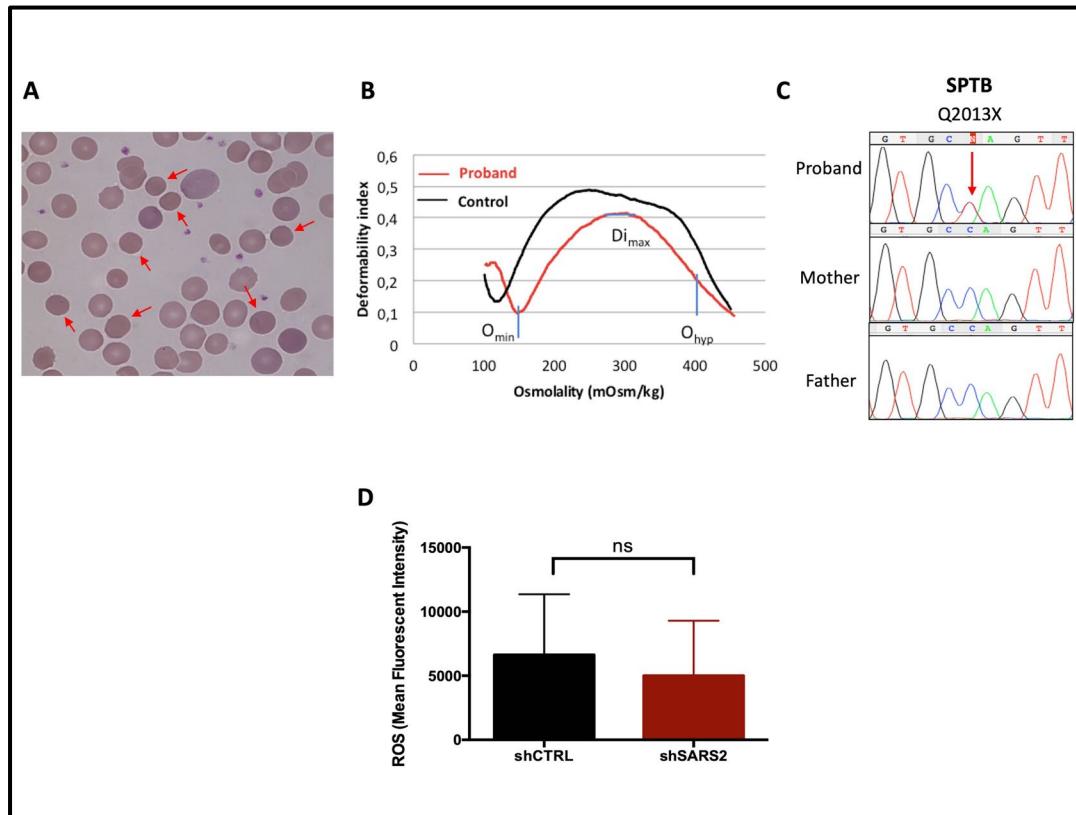
Table S2

Authors, year	Gender	Family	Diagnosis	Age at diagnosis	Anemia	Hemoglobin at diagnosis (g/dL)	Other cytopenia	Age at death	SARS2 mutation
This article	M	1	HUPRA syndrome	3 years old	Yes	9	No	5 years old	compound heterozygous c.1031G>A (p.R344Q) c.1205G>A (p.R402H)
Belostotsky et al, 2011	M	2	HUPRA syndrome	4 months	Yes	9	Thrombocytopenia, leukopenia	14 months	homozygous c.1169A>G (p.D390G)
Belostotsky et al, 2011	F	2	HUPRA syndrome	7 months	Yes	4,8	Leukopenia	10 months	homozygous c.1169A>G (p.D390G)
Belostotsky et al, 2011	F	3	HUPRA syndrome	4 months	No	N/A	No	13 months	homozygous c.1169A>G (p.D390G)
Rivera et al, 2013	F	4	HUPRA syndrome	15 months	Yes	8,4	No	26 months	homozygous c.1205G>A (p.R402H)
Rivera et al, 2013	M	4	HUPRA syndrome	2 months	Yes	7,6	No	21 months	homozygous c.1205G>A (p.R402H)
Linnankivi et al, 2016	M	5	Progressive spastic paresis	10 years old	No	N/A	No	Alive	homozygous c.1347>A (splicing mutation)
Zhou et al, 2021	F	6	HUPRA syndrome	4 years old	Yes	8,7	No	5 years old	compound heterozygous c.667G>A (p.V223M) c.1205G>A (p.R402H)

Supplemental Table 2: Hematological and molecular findings in patients carrying pathogenic SARS2 variants.⁷⁻⁹ N/A: not available

Supplemental figures

Figure S1



Supplemental Figure 1

(A) Peripheral blood smear of the patient showing the presence of spherocytes. (B) Ektacytometry curves of the proband and a control. O_{min} represents the osmolarity at which 50% of the cells hemolyze. D_{max} is the maximum elongation that the red blood cells can achieve under shear stress. O_{hyp} is the osmolarity at which the D_{max} is 50% of its maximum value. In the proband, the right shift of the O_{min} indicates a reduced surface to volume ratio; the D_{max} decrease reflects decreased red cell deformability due to a loss of surface area; and the left shift of the O_{hyp} corresponds to the dehydration of the cells (C) Sanger-sequencing validation of *SPTB* c.6037C>T (p.Q2013X) mutation detected by NGS. Red arrow indicates the position of the nucleotide's substitution. (D) At day 6 of the second phase of culture, reactive oxygen species (ROS) production was measured by flow cytometry in shCTRL and shSARS2-transduced cells using the CellROX deep red reagent. Results are represented as Mean Fluorescence Intensity of the stained cells. Error bars represent standard deviation (SD) from mean of three independent experiments. *P*-values are determined by two-tailed *t*-test. ns: not significant.