

The insecticides permethrin and chlorpyrifos show limited genotoxicity and no leukemogenic potential in human and murine hematopoietic stem progenitor cells

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SUPPLEMENTARY FIGURES

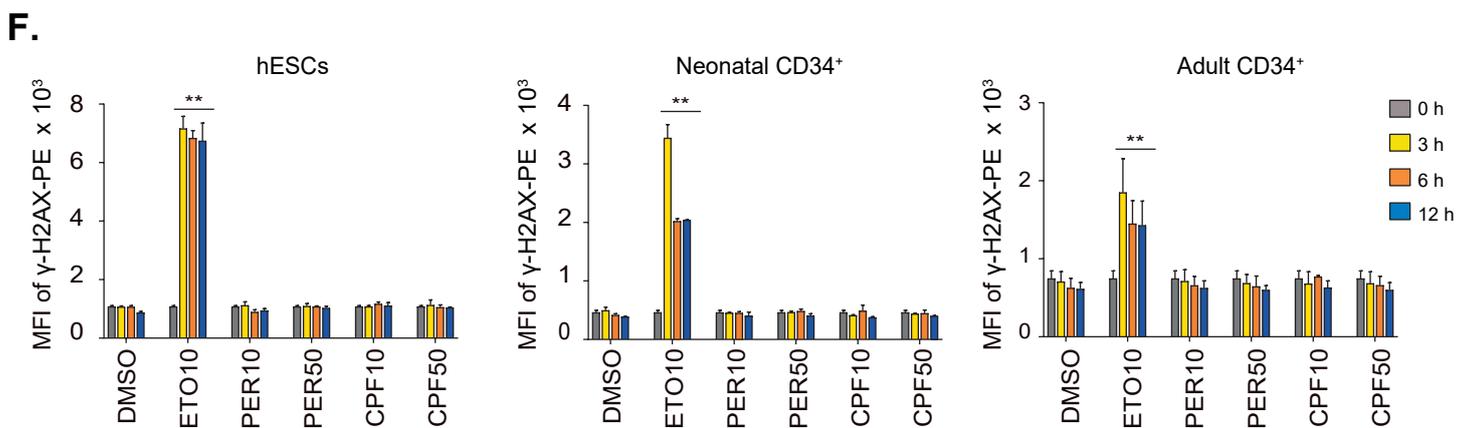
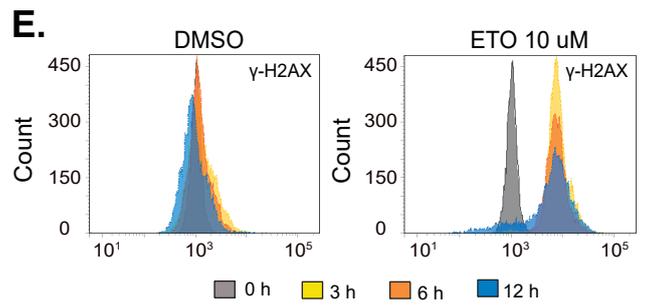
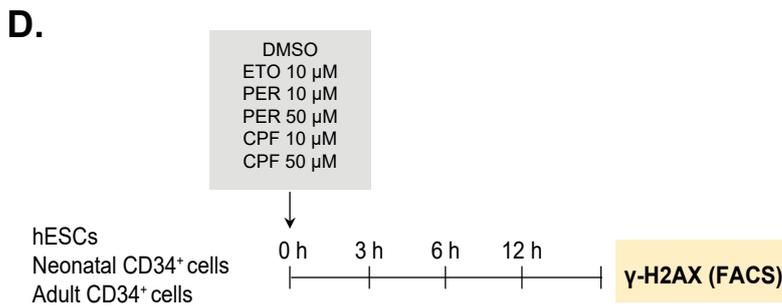
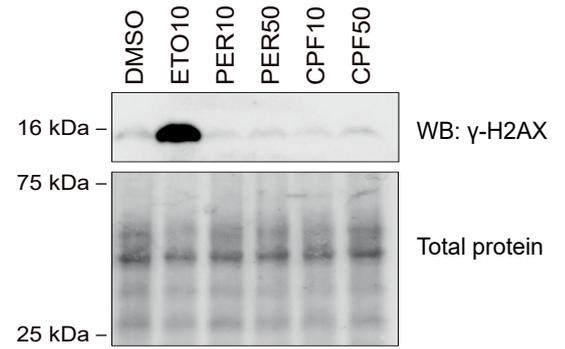
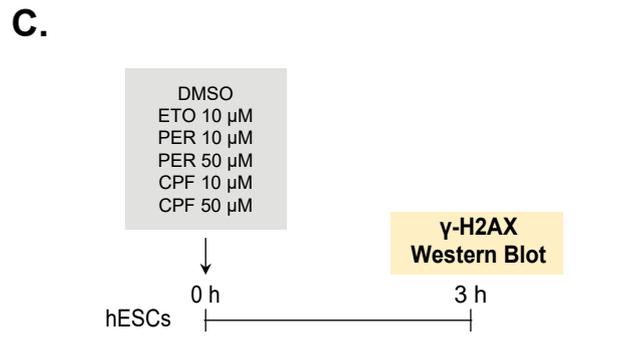
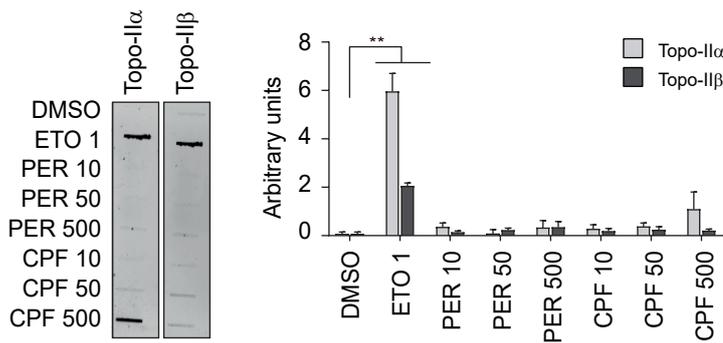
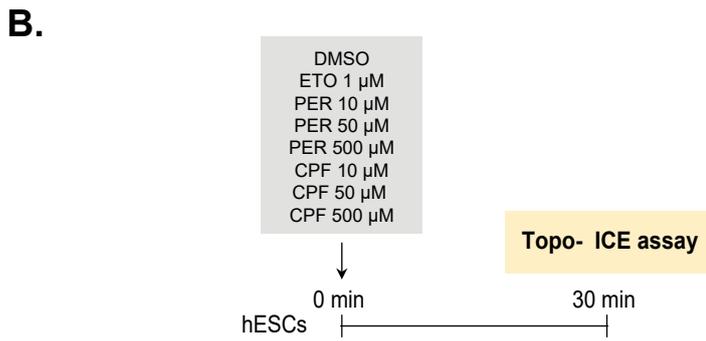
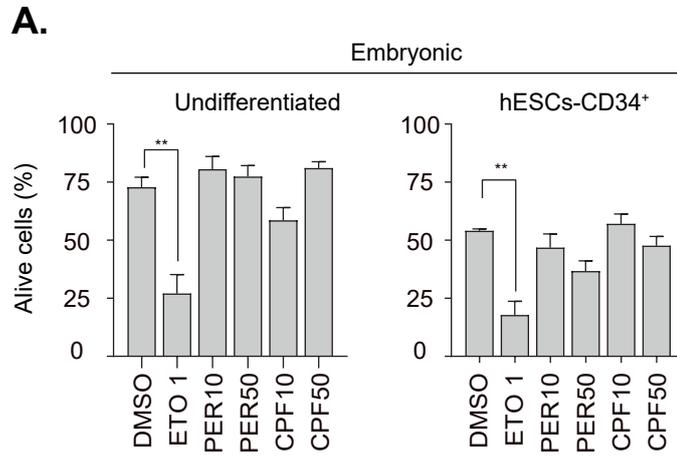
Figure S1. Permethrin and chlorpyrifos are not cytotoxic, are not *bona fide* topo-II poisons and do not affect DNA-DSB formation-repair in embryonic, neonatal or adult human CD34⁺ HSPCs. A) Viability of hESCs after 24h exposure to etoposide (ETO), permethrin (PER) and chlorpyrifos (CPF). **B)** Upper panel: Experimental design to evaluate the Topo-II poisoning ability of increasing concentrations of PER and CPF in a human topoisomerase *in vivo* complex of enzyme assay (ICE assay). DMSO- and ETO 1 μ M-treated hESCs were used as a negative and positive control, respectively. Lower panel: Representative image (left) and quantification (right) of the topo-II isoforms, topo-II α and topo-II β , covalently bound to genomic DNA following 30 min exposure. **C)** Upper panel: Experimental design to analyze the ability of increasing concentrations of PER and CPF to generate DSBs in hESCs, as determined by levels of phosphorylated H2AX at ser139 (γ -H2AX) by western blot. Lower panel: Western blotting of γ -H2AX after the indicated treatments. **D)** Experimental design to analyze the ability of increasing concentrations of PER and CPF to generate and repair DNA DSBs (determined by γ -H2AX by FACS) in hESCs, neonatal and adult CD34⁺ cells over 12 h. **E)** Representative FACS analysis of γ -H2AX levels on DMSO- and ETO-treated cells.

F) Median fluorescence intensity (MFI) of γ -H2AX determined at 0 h, 3 h, 6 h, and 12 h after a single-pulse of the indicated treatments. * $p < 0.05$, ** $p < 0.01$ compared with the 0 h time point.

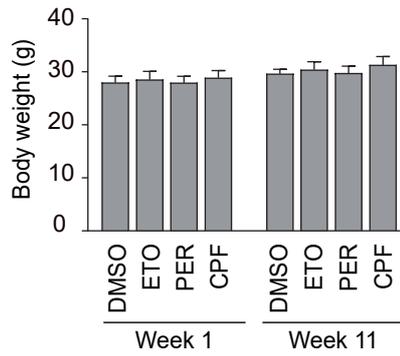
Figure S2. Continuous exposure to permethrin and chlorpyrifos fails to induce *MLL* breaks or leukemia in NSG-reconstituting human CD34⁺ HSPCs. **A)** Mice body weight at week 1 and week 11 of the experiment. Each NSG mouse weights an average of 30 g throughout the 12-week experiment. **B)** Actual water consumption per cage at early (3–7) and late (8–11) weeks of the experiment. If each NSG mouse drinks 5 ml of water/day, 140 ml of water consumption are expected per cage per week (the horizontal dotted line). **C)** Cartoon depicting the assumptions used to calculate the theoretical and actual concentration of etoposide, permethrin and chlorpyrifos at which mice were exposed in drinking water. For the target concentration of 10 mg/day/kg (= 0.3 mg/day/mouse) for permethrin and chlorpyrifos, one would expect a consumption of 8.4 mg/cage/week ($0.3 \text{ mg/day} \times 4 \text{ mice/cage} \times 7 \text{ days} = 8.4 \text{ mg/cage/week}$). For the target concentration of 5 mg/day/kg (= 0.15 mg/day/mouse) for etoposide, a consumption of 4.2 mg/cage/week ($0.15 \text{ mg/day} \times 4 \text{ mice/cage} \times 7 \text{ days} = 4.2 \text{ mg/cage/week}$) is expected. Based on the mice weight and water consumption shown in A and B, the actual intake of permethrin, chlorpyrifos and etoposide was 8.5, 9 and 4.2 mg/kg/day, respectively. **D)** GC-MS quantification of the PER metabolite, 3-phenoxybenzoic acid (3-PBA), and CPF metabolite, 3,5,6-trichloro-2-pyridinol (TCPy), in serum and urine. 3-BPA and TCPy were exclusively detected in mice exposed to permethrin, chlorpyrifos, respectively. **E)** Gating strategy to analyze human hematopoietic reconstitution in NSG mice by FACS. Human cells were identified as CD45⁺HLA-ABC⁺. Within this subpopulation, the immature (CD34⁺CD45⁺), myeloid (CD33⁺CD45⁺) and B-cell lymphoid (CD19⁺CD45⁺) graft was analyzed.

Figure S3. Exposure to etoposide, permethrin, and chlorpyrifos during pregnancy. **A)** Number of pups per litter after pregnancy exposure to the indicated compounds. **B)** Percentage of male and female pups in each treatment group. **C)** Scheme showing the purification steps to isolate Lin⁻c-Kit⁺ (LK cells)

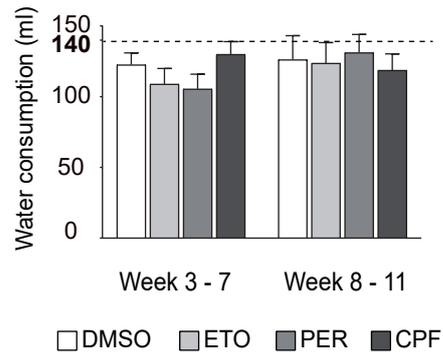
from BM to perform iFISH analysis. **D)** FACS gating strategy to analyze mature hematopoietic cells (myeloid, B and T) from peripheral blood and Lin-Sca-1⁺cKit⁺ (LSK) and hematopoietic stem and progenitor cells (HSPCs) subpopulations from bone marrow. From the HSPCs, we analyzed the hematopoietic stem cells (HSCs), multipotent hematopoietic progenitors (MPPs), and restricted hematopoietic progenitor cells 1 and 2 (HPC-1 and HPC-2) subsets. **E)** White blood cell (WBC), red blood cell (RBC) and platelet (PLT) counts in each treatment group in mothers and pups at weaning and in the adult offspring.



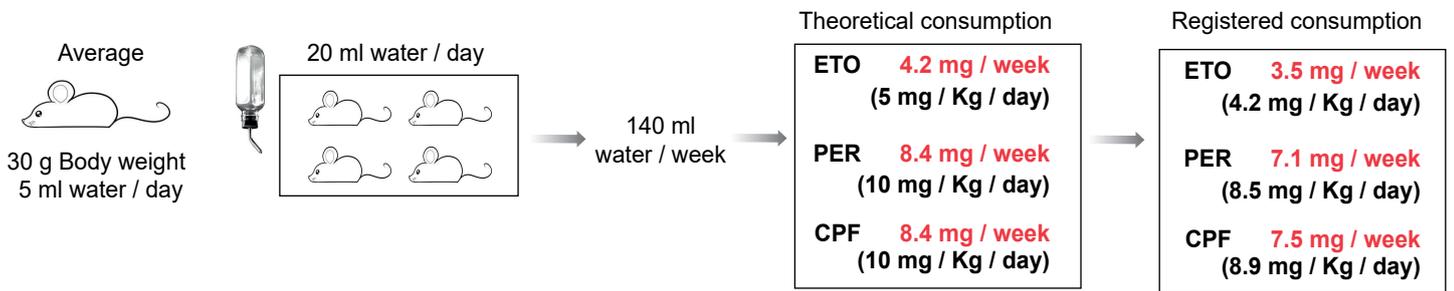
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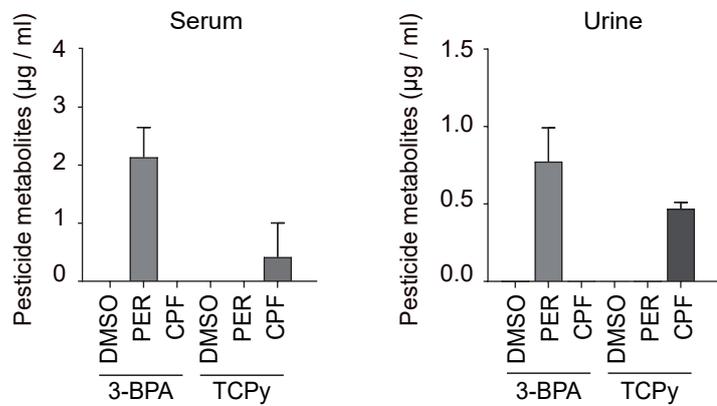
B.



C.



D.



E.

