

Early detection of T-cell lymphoma with T follicular helper phenotype by *RHOA* mutation analysis

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SUPPLEMENTARY TABLES

Table S1: Sequences of PCR primers and probes used for *RHOA* mutation detection.

	Sequence (5'-3')	Manufacturer
Forward primer	CCTATGACTTCTTGTGCATTGC	Sigma-Aldrich
Reverse primer	TACACCTCTGGGAAGTGGTC	Sigma-Aldrich
Total probe (Cy5)	[Cyanine5]GCTGCCATCCGGAAGAACTG[BHQ3]	Sigma-Aldrich
PNA wild-type clamp	AGCCTGTGGAAAGACA	Panagene (Cambridge Research Biochemicals)
LNA mutant probe (Fam)	[6FAM]AGCC[+T]GT[+G][+T][+A]AAGA[+C]A[BHQ1]	Sigma-Aldrich

Table S2: Mutational information for clonal hematopoiesis of indeterminate potential (CHIP) samples.

Sample	RHOA qPCR	Gene	Nucleotide change	Amino acid change	VAF (%)
CHIP 1	Negative	SRSF2	c.C284A	p.P95H	7.4
CHIP 2	Negative	JAK2	c.G1849T	p.V617F	14.7
		TET2	c.A3797G	p.N1266S	2.9
CHIP 3	Negative	TET2	c.C2020T	p.Q674X	17.5
			c.G3893A	p.C1298Y	15.8
			c.G4249T	p.V1417F	3.1
		TP53	c.C523G	p.R175G	3.5
CHIP 4	Negative	DNMT3A	c.1154delC	p.P385fs	9.1
		SF3B1	c.A2098G	p.K700E	10.4
			c.G1874T	p.R625L	6.5
CHIP 5	Negative	CBL	c.2434+1G>A	Essential splicing site	1.1
		DNMT3A	c.2173+1G>A	Essential splicing site	26.1
		PPM1D	c.C1570T	p.Q524X	4.8
CHIP 6	Negative	CBL	c.G570A	p.W190X	2.3
		SF3B1	c.A2098G	p.K700E	0.9
		SF3B1	c.G1998C	p.K666N	34.2
CHIP 7	Negative	DNMT3A	c.2086delC	p.Q696fs	0.9
		TP53	c.G818A	p.R273H	1.4
		U2AF1	c.A470C	p.Q157P	11.8
CHIP 8	Negative	DNMT3A	c.2567delA	p.E856fs	5.4
			c.1122+1G>-	Essential splicing site	1.3
		TET2	c.C863G	p.P288R	1.9
CHIP 9	Negative	DNMT3A	c.G1904A	p.R635Q	4.0
		TP53	c.G1006T	p.E336X	1.9
CHIP 10	Negative	SF3B1	c.G1998T	p.K666N	10.6
		U2AF1	c.A470G	p.Q157R	10.8
CHIP 11	Negative	DNMT3A	c.G2645A	p.R882H	32.1
CHIP 12	Negative	GNB1	c.A169G	p.K57E	4.2
		SF3B1	c.G1998T	p.K666N	16.2
		TET2	c.944delC	p.S315fs	3.2
		TET2	c.1305delC	p.H435fs	1.2
		TET2	c.A3821G	p.Q1274R	3.3

		TET2	c.T3965A	p.L1322Q	3.4
		TET2	c.C4624T	p.Q1542X	1.3
		TET2	c.C5473T	p.Q1825X	40.9
CHIP 13	Negative	SRSF2	c.C284T	p.P95L	11.7
CHIP 14	Negative	DNMT3A	c.G989A	p.W330X	6.5
		SRSF2	c.C284A	p.P95H	4.0
		TET2	c.C3116A	p.S1039X	2.9
			c.4228delC	p.P1410fs	2.3
			c.4931_4935del	p.P1644fs	1.6
CHIP 15	Negative	SF3B1	c.2098A>G	p.K700E	40.4
(High-risk) CHIP 16	Negative	TET2	c.2031_2032delTG	Cys677fs	36.4

Table S3: Summary of laboratory data of non-diagnostic biopsies in AITL with *RHOA* p.Gly17Val mutation.

Case	Age/ Sex	<i>RHOA</i> mutation	Biopsy site	Core or excision biopsy	Preceding or follow up biopsy	Diagnostic category	Histological & immunohistochemical findings	T-cell clonality	B-cell clonality	Final histological opinion
C2	64/F	Positive	Bone marrow	Trephine biopsy	Follow up	C	There is a nodular and interstitial infiltrate of small lymphoid cells, mainly T-cells, predominantly CD4 ⁺ , no evidence of loss of pan T-cell markers	Weak clonal in polyclonal background	n/a	No definite aberrant T-cell phenotype, lack of molecular evidence, thus not diagnostic
C3-1	66/F	Positive	Bone marrow	Trephine biopsy	Preceding	C	Non-specific changes. A focal cluster of multiple perivascular lymphoid aggregates, dominantly CD4 ⁺ T cells, but very few expressing PD1	Polyclonal	n/a	Suspicious of bone marrow involvement by AITL, but low number of assumed neoplastic cells and no evidence of clonal TCR rearrangement, thus not diagnostic
C3-2	66/F	Positive	Liver	Core biopsy	(Preceding) Same date as diagnostic biopsy	C	Focal inflammatory cells including lymphocytes and histiocytes in parenchyma and portal tracts	Weak polyclonal		No histological evidence of involvement by a T-cell lymphoma
C5	72/ M	Positive	Skin	Punch biopsy	Follow up	C	There is a nodular dermal inflammatory infiltrate, involving all levels of the dermis and also the subcutis. The infiltrate is perivascular and periadnexal, comprising sheets of histiocytes, in areas forming granulomas with occasional giant cells, and inflammatory cells and lymphocytes. Immunohistochemistry shows mixed CD4 ⁺ and CD8 ⁺ T cells with no evidence of loss of pan T-cell markers	Clonal, identical pattern to that of previous biopsy showing AITL	n/a	No histological evidence of involvement by a T-cell lymphoma, although shown by clonality analysis
C7-1	82/ M	Positive	Skin	Punch biopsy	Preceding	C	Mild perivascular chronic inflammatory cell infiltration, plus a predominantly lobular panniculitis associated with focal fat necrosis. The infiltrate is predominantly small T cells, admixed with B cells and histiocytes	n/a	n/a	Panniculitis (please also refer to Table 1)
C7-4										
C7-5										
C9	82/ M	Positive	Bone marrow	Trephine biopsy	Preceding	C	There are numerous lymphoid aggregates, with a paratrabeular location, which are composed of small mixed B and T cells. The majority of T cells are CD4 ⁺ , no loss of pan T-cell markers. EBER-ISH negative	Polyclonal	n/a	Suspicious lymphoma involvement, but not diagnostic as no supporting evidence by clonality and flow cytometry analyses
C11	77/ M	Positive	Right inguinal lymph node	Core biopsy	Preceding	C	Small, fragmented and poorly fixed tissue cores showing polymorphous infiltrates, particularly histiocytes and T cells, and FDC expansion. T cells are predominantly CD4 ⁺ , expressing pan T-cell markers, with a small subset PD1 positive. EBER-ISH negative	Weak clonal	Polyclonal	Suspicious for a T-cell lymphoma, but lacking convincing histological and molecular evidence, advised for re-biopsy
C13	81/ M	Positive	Bone marrow	Trephine biopsy	Follow up	C	There are multiple intraparenchymal & occasionally paratrabeular aggregates of histiocytes, numerous small to slightly enlarged lymphoid cells which are mainly CD4 ⁺ T cells, showing possible loss of CD7, occasionally PD1 positive, but negative for CD10 and BCL6. Occasional EBER ⁺ immunoblasts	Weak polyclonal	n/a	Low lesional cell content and no evidence of clonal TCR gene rearrangement, thus suspicious but not diagnostic of involvement by a T-cell lymphoma
C17	76/ M	Positive	Cervical lymph node	Core biopsy	Preceding	C	Small and poorly preserved tissue fragments. A mixed population of small lymphoid cells, CD30 positive immunoblasts, and plasma cells. There are no recognizable follicular structures on morphology and immunohistochemistry. No evidence of loss of pan T-cell markers, high number of EBER ⁺ immunoblasts	Weak clonal	Clonal	Little tissue for evaluation and dominant B-cell component render difficulty in diagnosis

See Table 1

C19	67/M	Positive	Lymph node	Core biopsy	Preceding	C	A mixture of B and T cells in a vaguely nodular intermixed distribution. T cells are predominantly CD8 ⁺ and show partial loss of CD7. The smaller CD4 ⁺ population shows CD10, PD1 and possibly BCL6 expression. There is FDC expansion and a high number of diffuse EBER ⁺ B cells across a range of B-cell differentiation	Clonal	Clonal	The B cell component looks like immunodeficiency associated LPD and there is a relatively small TFH component. Advised for re-biopsy if lymphadenopathy persists or progresses
C30	79/M	Positive	Cervical lymph node	Excision	Preceding	C	Partial effacement of the lymph node architecture and expansion of the interfollicular area by a polymorphous population of lymphoid cells. B cells express pan B-cell markers (CD20, CD79a, CD19), and MUM1, but negative for CD10 and BCL6. A high proportion of the B cells were EBER ⁺ and showed IG kappa light chain restriction. The lymphoid follicles appeared to be reactive and showed no apparent expansion of TFH cells, with only a few CD10 ⁺ cells spilling out of the germinal centers	Weak multiple clonal re-arrangements	Clonal	Clonal EBV positive polymorphous lymphoproliferation, advised for follow up with a low threshold for re-biopsy. On review with RHOA mutation positivity, pattern-1 AITL is considered
C37-1	81/M	Positive	Lymph node	Core biopsy	Preceding	C	There are multiple, variably preserved germinal centers with slightly expanded FDC-meshworks and a population of CD4 ⁺ /PD1 ⁺ follicular helper T cells in a predominantly peri-germinal centre localization, which express pan T-cell markers and CD10. Germinal center cells are negative for CD10 and BCL6 and these large B cells spill beyond the follicles and show CD30 expression. EBER-ISH highlights scattered positive cells	Weak clonal	Weak clonal	Suspicious of a T-cell lymphoma, particularly AITL, but not diagnostic due to small tissue and lack of strong molecular evidence
C37-3		Positive	Skin	Punch biopsy	Follow up	C	Multiple perivascular infiltrates comprising a mixed population of small to medium atypical lymphoid cells admixed with histiocytes and plasma cells. The lymphoid cells are mixed CD4 ⁺ and CD8 ⁺ T cells. Many T cells express PD1 and show no loss of expression of pan T-cell markers. There are also numerous B cells and scattered EBER ⁺ cells, which are also CD30 ⁺ . The plasma cells display lambda light chain restriction	Weak polyclonal	Weak clonal	A clonal B-cell process with plasmacytoid differentiation, and not entirely excluding a T-cell lymphoma, but not diagnostic
C38-1	80/M	Positive	Lymph node	Core biopsy	Preceding	C	The nodal architecture is effaced by a diffuse mixed infiltrate of small, intermediate and large lymphoid cells. There is mixed CD4 and CD8 ⁺ T cells, with CD4 expression in some large cells. There is loss of CD7 expression in T cells	Weak clonal	Clonal	Suspicious of a T-cell lymphoma, not diagnostic
C38-2	80/M	Positive	Cervical lymph node	Excision	Preceding	C	This is a mostly necrotic sample. In viable areas, a marked proliferation of variably sized blood vessels is noted. The T cells are small to medium sized with preserved pan T-cell marker expression. They are predominantly CD8 ⁺ with a smaller CD4 ⁺ population. There is no significant CD30, PD1, CD10, BCL6 expression in the T cell population. There is expansion of FDC-meshworks. EBER-ISH negative	Weak multiple clonal re-arrangements	Clonal	Not diagnostic, not excluding B-cell lymphoma
C43	64/M	Positive	Cervical lymph node	Core biopsy	Preceding	C	Lymph node architecture appears essentially preserved. There is a striking paracortical hyperplasia by small lymphocytes and histiocytes. T cells are dominantly CD4 ⁺ , show no loss of pan T-cell markers, and appear negative for CD10 and BCL6. PD1 staining is largely restricted to follicular T cells. Some FDC-meshworks appeared slightly ragged. EBER-ISH negative	Polyclonal	n/a	Reactive T-cell hyperplasia

SUPPLEMENTARY FIGURES

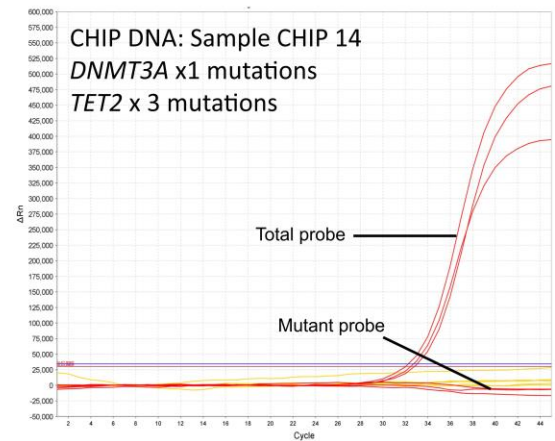
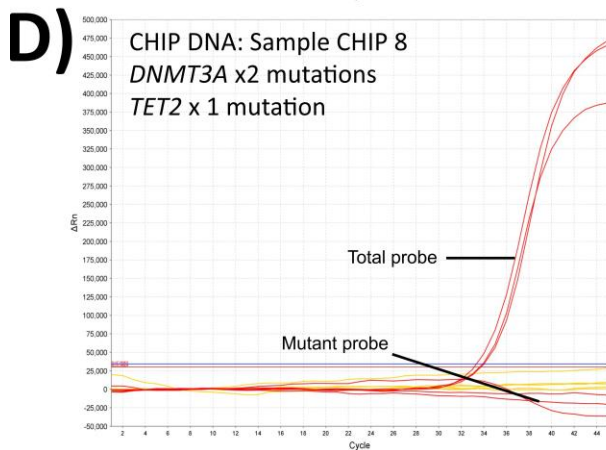
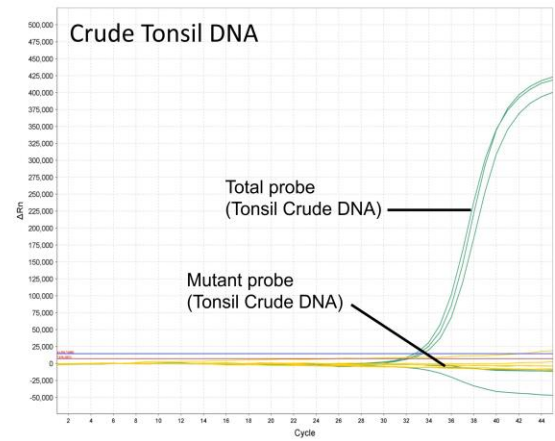
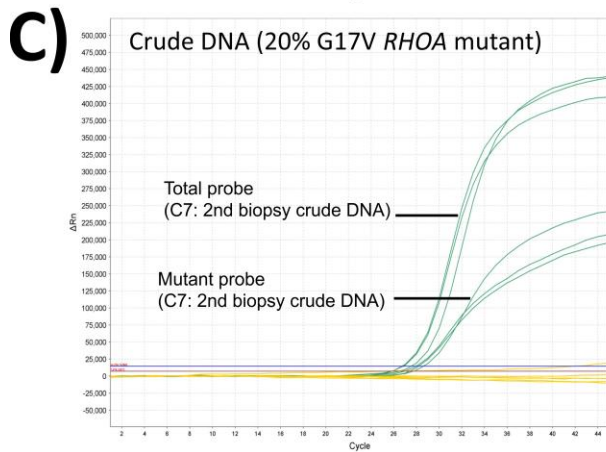
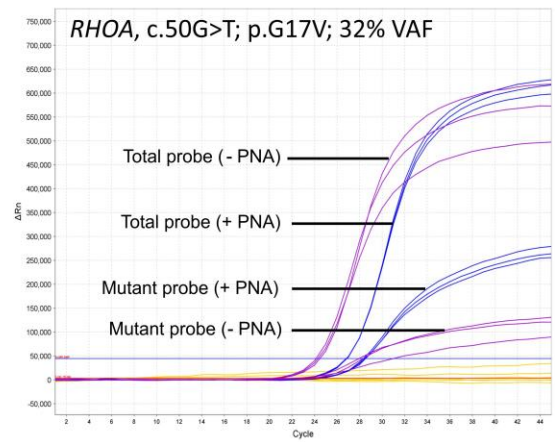
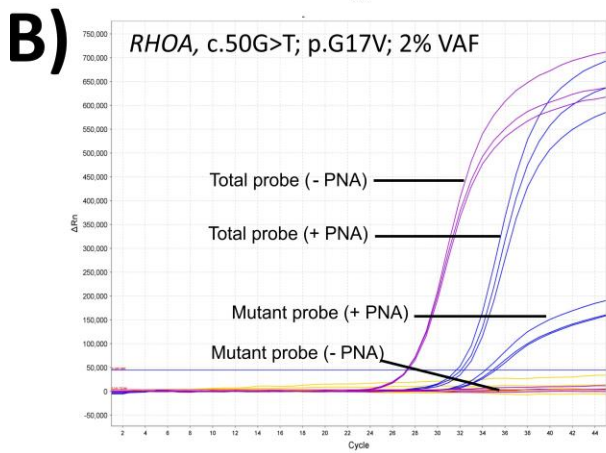
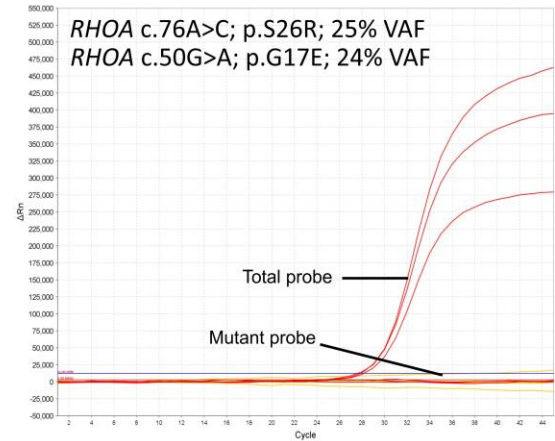
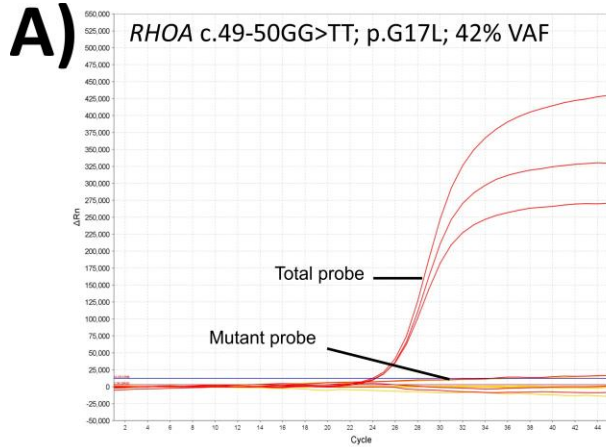
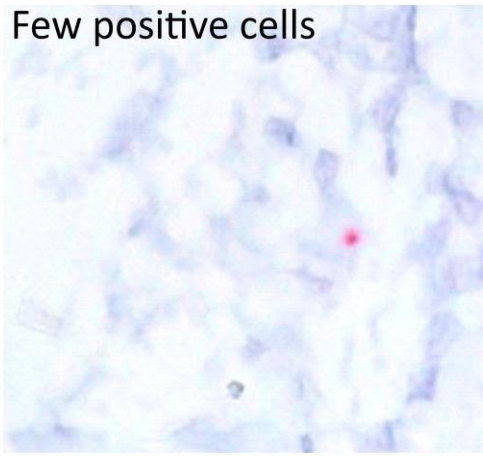


Figure S1. Representative LNA-PNA qPCR plots for detection of *RHOA* p.Gly17Val.

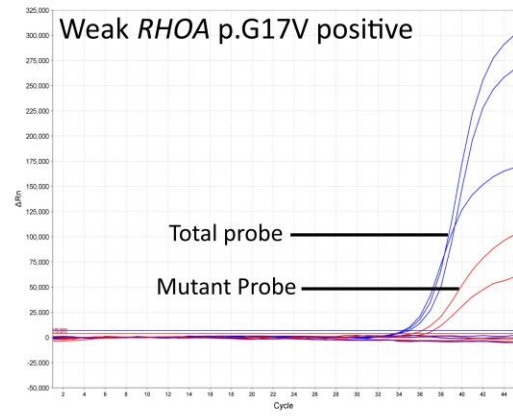
- A) The qPCR is highly specific for *RHOA* (c.50G>T; p.Gly17Val) mutation, not detecting any other *RHOA* changes.
- B) Examples of qPCR for detection of *RHOA* (c.50G>T; p.Gly17Val) mutation in the presence or absence of PNA probe which represses the amplification of the wild-type *RHOA* sequence, thus boosting the detection of the *RHOA* mutation. As demonstrated by the total probe amplification curve shift in presence of PNA clamp, while the mutant probe amplification curve is enhanced.
- C) Examples of qPCR for detection of *RHOA* (c.50G>T; p.Gly17Val) mutation using crude DNA preparations.
- D) Examples of qPCR for detection of *RHOA* (c.50G>T; p.Gly17Val) mutation for clonal hematopoiesis of indeterminate potential (CHIP) cases with both *TET2* and *DNMT3A* mutations.

First Biopsy
(12/08/2015)

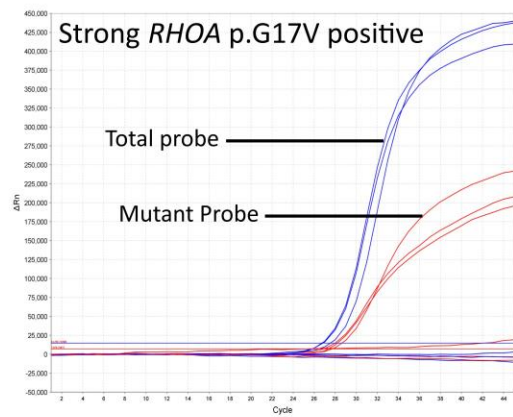
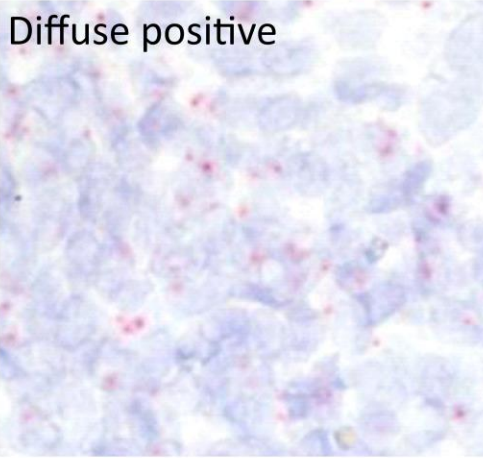
BaseScope-ISH



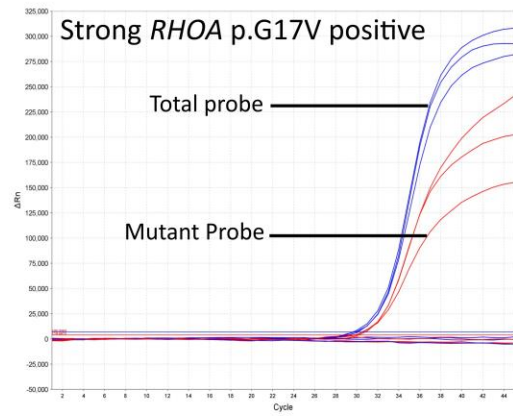
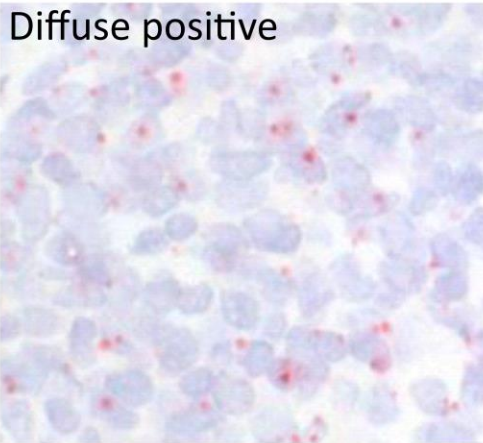
qPCR for *RHOA* p.G17V



Second Biopsy
(11/07/2016)



Third Biopsy
(1/08/2016)



Fifth Biopsy
(30/12/2017)

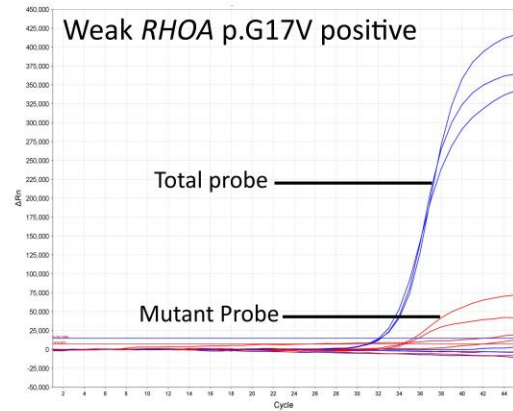
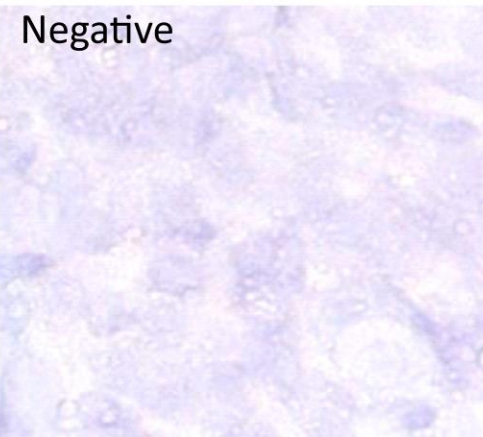


Figure S2: Detection of the AITL cells by BaseScope in situ hybridization (ISH) using a specific probe to the unique VDJ junction sequence of the lymphoma cells (TRB-V5 clone), and qPCR of *RHOA* Gly17Val in case 7.

Left panel: Representative images of BaseScope-ISH;

Right panel: *RHOA* Gly17Val qPCR plots for various biopsies of case 7.