## **SUPPLEMENTARY APPENDIX**

#### Profiling the mutational landscape of coagulation factor V deficiency

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# SUPPLEMENTARY MATERIAL Profiling the mutational landscape of coagulation factor V deficiency

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## Supplementary Table 1: List of previously reported mutations in the *F5* gene with relevant references.

cDNA level *	Native protein	Mature protein **	Reference
c.692T>G	p.Met231Arg	p.Met203Arg	1
c.1321C>T	p.Arg441Cys	p.Arg413Cys	2
c.2218C>T	p.Arg740X	p.Arg712X	3
c.2862delT	p.Ser955AlafsX4	p.Ser927AlafsX4	4
c.3088C>T	p.Arg1030X	p.Arg1002X	3
c.3296delA	p.Asp1099AlafsX72	p.Asp1071AlafsX72	5
c.3481C>T	p.Arg1161X	p.Arg1133X	6
c.3924_3927delTCAG	p.Ser1308ArgfsX24	p.Ser1280ArgfsX24	7
c.4906G>A	p.Glu1636Lys	p.Glu1608Lys	8
c.5189A>G	p.Tyr1730Cys	p.Tyr1702Cys	9
c.6304C>T	p.Arg2102Cys	p.Arg2074Cys	10
c.6305G>A	p.Arg2102His	p.Arg2074His	11
c.6419G>A	p.Gly2140Asp	p.Gly2112Asp	12
c.6443T>C	p.Met2148Thr	p.Met2120Thr	13
The HR2 Haplotype: p.Met385Thr, p.His1299Arg, p.Met1736Val, p.Asp2194Gly			14

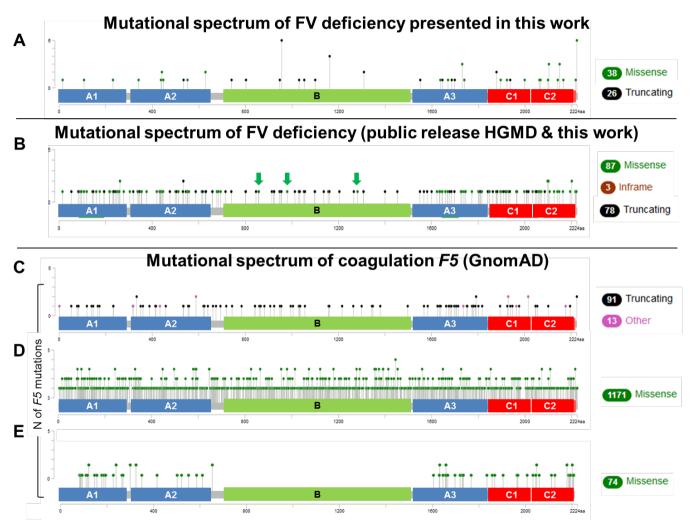
The table reports the previously reported mutations identified in our case series of 50 FV-deficient patients All mutations, are named following the official recommendations of the Human Genome Variation Society (HGVS, http://www.hgvs.org/mutnomen/recs-DNA.html). For mutation names at the protein level, the three-letter code annotation was adopted. In the case of the HR2 allele, the 4 major missense variants contributing to the haplotype are listed.

Nd, not done; Homo, homozygous; Hetero, heterozygous; Comp hetero, compound heterozygous.

<sup>\*</sup> Numbering starting from ATG, according to NM 000130.4.

<sup>\*\*</sup> Numbering omitting the signal peptide (28 amino-acid long).

#### **SUPPLEMENTARY FIGURE 1**



**Supplementary Figure 1: Mutational spectrum of the** *F***5 gene.** The five lolliplots report *F***5** mutations projected on the schematic A1-A2-B-A3-C1-C2 domain representation of the human FV protein. In all schemes, mutations are reported as circles (green for missense mutations; black for nonsense, nonstop, point frameshift deletion/insertion, and splice site mutations; brown for inframe deletions and insertions; violet for all other types

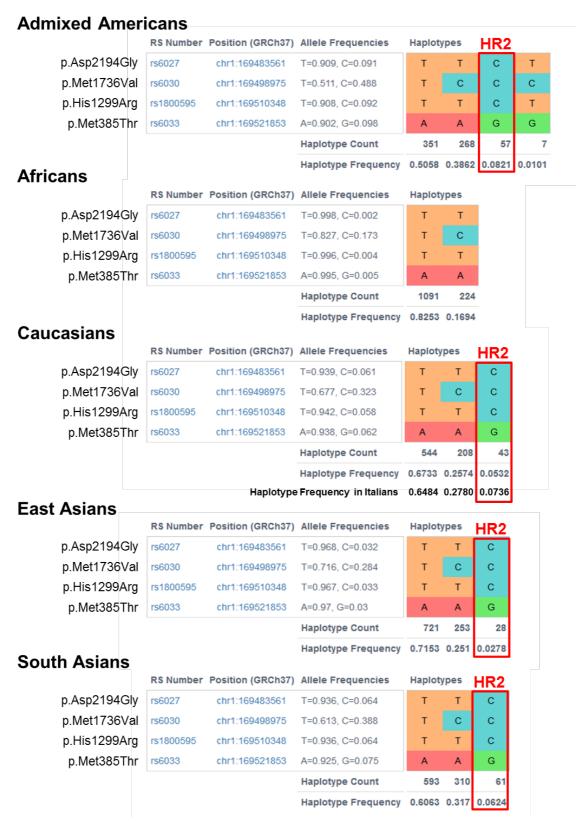
of mutations, such as long duplications). In all cases, except in panel B, the score on the left corresponds to the number of patients/individuals carrying the relevant mutation.

In particular:

- A) Mutational spectrum emerging from the present work;
- **B)** Mutational spectrum of FV deficiency merging the data from the present work together with those reported in the Human Gene Mutation Database (HGMD, public version, http://www.hgmd.cf.ac.uk/ac/index.php; accessed on April 05, 2019). The only three missense mutations identified in the B domain, are pointed by a green arrow;
- **C-E)** Mutational spectrum of the *F5* gene emerging from the data present in the GnomAD repository (data on >125,000 exomes and >15,000 genomes from European, African, Asian, and Admixed American populations; https://gnomad.broadinstitute.org/; accessed on April 05, 2019; all populations were included in this representation). In panel C, all mutations except missense variants are reported. In panel D, only missense variants are present, independently from their pathogenicity. In panel E, all missense variants predicted ad damaging by three out of three predictions programs (MutationAssessor/SIFT/Polyphen-2) are reported. From the comparison between panels D and E, it emerges the high tolerance to missense substitutions of the unstructured B domain.

Lolliplots were produced using the NM\_000130 RefSeq and the MutationMapper software, freely available through the Cbioportal (https://www.cbioportal.org/mutation\_mapper). The the distribution of the different types of mutations in the *F5* genes in panel B is not significantly different from that reported in panels C+E (Pearson chi-square test =0.14).

#### **SUPPLEMENTARY FIGURE 2**



**Supplementary Figure 2: Frequency distribution of the HR2 haplotype across different populations.** The genotypes of the four main polymorphisms constituting the HR2 haplotype were used to calculate the HR2 frequency distribution across different populations. Data on Admixed Americans, Africans, Caucasians, east Asians, and South Asians were retrieved from the 1000 Genomes Project reference panels (data from Phase 3, Version 5;

http://www.internationalgenome.org/). For the Italian population, we retrieved genotypes of the HR2 polymorphisms from our in-house cohort of 3541 individuals, for whom whole-exome sequencing (WES) data are available. WES was performed at the Broad Institute (Boston, MA, USA); exome capture methods, sequencing, variant annotation, and data processing of the samples have been described previously.<sup>15</sup>

Haplotypes were produced through the LDhap application web tool of the LDlink suite (https://ldlink.nci.nih.gov/?tab=ldhap).

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