

Anti-apoptotic *BCL2L2* increases megakaryocyte proplatelet formation in cultures of human cord blood

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ABSTRACT

Apoptosis is a recognized limitation to generating large numbers of megakaryocytes in culture. The genes responsible have been rigorously studied *in vivo* in mice, but are poorly characterized in human culture systems. As CD34-positive (*) cells isolated from human umbilical vein cord blood were differentiated into megakaryocytes in culture, two distinct cell populations were identified by flow cytometric forward and side scatter: larger size, lower granularity (LLG), and smaller size, higher granularity (SHG). The LLG cells were CD41a^{High} CD42a^{High} phosphatidylserine^{Low}, had an electron microscopic morphology similar to mature bone marrow megakaryocytes, developed proplatelets, and displayed a signaling response to platelet agonists. The SHG cells were CD41a^{Low} CD42a^{Low} phosphatidylserine^{High}, had a distinctly apoptotic morphology, were unable to develop proplatelets, and showed no signaling response. Screens of differentiating megakaryocytes for expression of 24 apoptosis genes identified *BCL2L2* as a novel candidate megakaryocyte apoptosis regulator. Lentiviral *BCL2L2* overexpression decreased megakaryocyte apoptosis, increased CD41a⁺ LLG cells, and increased proplatelet formation by 58%. An association study in 154 healthy donors identified a significant positive correlation between platelet number and platelet *BCL2L2* mRNA levels. This finding was consistent with the observed increase in platelet-like particles derived from cultured megakaryocytes over-expressing *BCL2L2*. *BCL2L2* also induced small, but significant increases in thrombin-induced platelet-like particle α IIb β 3 activation and P-selectin expression. Thus, *BCL2L2* restrains apoptosis in cultured megakaryocytes, promotes proplatelet formation, and is associated with platelet number. *BCL2L2* is a novel target for improving megakaryocyte and platelet yields in *in vitro* culture systems.

Introduction

Hematopoietic stem cells (HSC) are essential for reconstituting hematopoiesis in the setting of bone marrow (BM) transplantation and are of great value for studies of the basic biology of hematopoiesis. Differentiating HSC into megakaryocytes (MK) in culture has become a standard approach to study megakaryocytopoiesis (MKpoiesis). MK culture systems are also powerful tools for the functional assessment of novel platelet genes, gene variants, protein-coding transcripts, and microRNA associated with platelet reactivity and clinical hemorrhagic or thrombotic disorders. Recent advancements in MKpoiesis research have opened up a new, promising field of *ex vivo* manufacturing of platelets,¹⁻⁴ with the ultimate goal of infusing these products into patients with thrombocytopenia or qualitative

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platelet disorders. Improvements in this technology may also allow 'designer' platelets to be engineered that overcome immunological incompatibility and infection issues.⁵

A major limitation of all MK culture systems is the relatively short time period for which the differentiating MK can be kept viable before they die through apoptosis.⁶ The role of apoptosis during MKpoiesis is somewhat controversial,⁷ with some data supporting a role for the intrinsic pathway of apoptosis in platelet production,⁸⁻¹⁰ while other studies show that MK must restrain apoptosis to survive and progress safely through proplatelet formation and platelet generation.¹¹⁻¹⁵ However, the apoptosis regulators of human cord blood-derived (CB)-MK cultures remain poorly understood. During the course of our studies with cultured CB MK, we observed two distinct populations of cells by forward and side scatter flow cytometry. The aims of the current study were to determine: (i) whether there were viability and apoptosis differences between these two populations; and (ii) molecular mechanism(s) of apoptosis regulation during Mkpoiesis. We also wanted to begin to identify approaches that would reduce MK apoptosis in order to produce greater yields of MK and platelet in cultures. We demonstrate that the anti-apoptosis Bcl2 family member *BCL2L2* (encoding Bcl-w) regulates cultured MK apoptosis, promotes proplatelet formation, and is associated with human platelet number.

Methods

Primary megakaryocyte cultures

Human umbilical cord CB was obtained from the New York Blood Center (New York, NY, USA) under institutional review board (IRB) approval (00108527). CD34⁺ hematopoietic stem and progenitor cells (HSPC) were isolated from human umbilical vein CB and cultured in serum free expansion media supplemented with 25 ng/mL of stem cell factor (SCF) and 20 ng/mL of thrombopoietin (TPO) (Peprotech, Rocky Hill, NJ, USA) for six days. Cells were cultured with 50 ng/mL TPO only from days 6-13.¹⁴ Adult granulocyte-macrophage colony-stimulating factor (GM-CSF) mobilized CD34⁺ HSPC were purchased from the Utah Cell Therapy and Regenerative Medicine Center (Salt Lake City, UT, USA) under the University of Utah IRB approval (00108527).

Megakaryocyte proplatelet formation assay

Day 9 transduced cells were plated at 2×10^4 cells/mL in 60 μ -Dish Grid-500 plates (Ibidi, Fitchburg, WI, USA) using fresh medium supplemented with 50 ng/mL TPO. On day 13, the proplatelet forming (PPF) MK, defined as displaying at least one filamentous pseudopod, were scored with a light microscope blinded as to experimental group. Images were taken at room temperature under 40x objective, numerical aperture 1.35, using FV1000 confocal laser scanning microscope (Olympus, Center Valley, PA, USA). The percentage of PPF MK was calculated as the number of PPF MK compared to the total number of round cultured cells analyzed. An average 200 cells were counted per condition.

Integrin α IIb β 3 activation assessment

Cells were resuspended in Tyrode's buffer (138 mM NaCl, 5.5 mM dextrose, 12 mM NaHCO₃, 0.8 mM CaCl₂, 0.4 mM MgCl₂, 2.9 mM KCl, 0.36 mM Na₂HPO₄, 20 mM Hepes, pH 7.4). Integrin α IIb β 3 activation was quantified with FITC-labeled PAC1 (1:100) (BD Pharmingen) in response to stimulation¹⁵ with no agonist (resting), 100 μ M PAR4-AP [GL Biochem (Shanghai) Ltd., China], 250 nM thrombin (Enzyme Research, South Bend,

IN, USA) or 10 μ g/mL CRP (synthesized at Baylor College of Medicine and cross-linked with glutaraldehyde) for 20 minutes (min) at 37°C, followed by 4% paraformaldehyde fixation at room temperature. Cells were analyzed on a Cytotflex or BD Accuri C6 flow cytometer.

Statistical analysis

All statistical analyses were performed using GraphPad Prism 6 software version 10.1 (La Jolla, CA, USA) and reported as Mean \pm Standard Error of Mean (SEM). Fold changes for *BCL2L2* levels over time in cultures and for lentiviral overexpression were presented as log₂ (fold change) compared to their respective controls (day 6 for changes in *BCL2L2* levels over time and empty vector control for overexpression) and analyzed by a one-sample *t*-test. Log₂ transformation was adopted to have a normally distributed fold-change data, determined by Kolmogorov-Smirnov normality test. The relationship between platelet *BCL2L2* mRNA expression levels and platelet counts in the 154 healthy human donors in the Platelet RNA Expression Study 1 (PRAX-1) was assessed by Pearson's correlation with 95% Confidence Interval (95%CI). For all other analyses, statistical significance was assessed using paired Student *t*-test. *P*<0.05 was considered statistically significant.

See *Online Supplementary Appendix* for further details of the experiments.

Results

Subsets of cultured megakaryocytes show differing viability

CD34⁺ cells were isolated from human umbilical vein CB and cultured to generate MK as described previously.¹⁴ By day 13, CD34⁺ cells were differentiated into MK, as demonstrated by the expression of mature MK markers (*Online Supplementary Figure S1A-D*). Day 13 MK were larger than day 0 HSC (*Online Supplementary Figure S1E*) and demonstrated polyploid MK (*Online Supplementary Figure S1F and G*). Thus, these culture conditions promoted generation of mature, polyploid MK.

Flow cytometric analysis of day 13 cultures revealed distinct cell populations differing by forward scatter (FSC) and side scatter (SSC) characteristics (Figure 1A): larger MK with lower granularity [larger, lower granular (LLG)] and smaller MK with higher granularity [smaller, higher granular (SHG)]. Peripheral blood (PB) mobilized adult CD34⁺ cells cultured under the same conditions also generated similar subpopulations of MK (*Online Supplementary Figure S2*). Approximately 80% of LLG MK expressed both CD41a/CD42a and CD41a/CD42b markers, while only 20-30% SHG MK were double positive for mature MK markers (Figure 1B). Lower surface expression of CD42b (GPIIb α) in day 13 SHG cells compared to LLG cells is consistent with mitochondrial damage¹⁶ as observed in apoptotic adult mobilized HSPC-derived MK.¹⁷

Since the numbers of SHG MK showed less differentiation towards mature MK than the LLG MK, and because SHG MK showed reduced CD42b expression, we considered whether SHG MK may have undergone (or be undergoing) apoptosis. Figure 1C shows few CD41a⁺ LLG MK-bound annexin V [termed phosphatidylserine (PS)^{low}], a marker of apoptosis-induced phosphatidylserine expression, whereas a significantly higher percentage of CD41a⁺ SHG MK bound annexin V (termed PS^{high}). Anti-annexin

V microbeads were used to separate day 13 PS^{Low} LLG MK from PS^{High} SHG MK (purity shown in *Online Supplementary Figure S3*). Transmission electron microscopy revealed that PS^{Low} LLG MK exhibited many typical features of BM MK, including large cells with multi-lobed nuclei, mitochondria, granules and surface protrusions (Figure 2A). In contrast, most of the PS^{High} SHG MK were apoptotic, displaying membrane blebbing, highly condensed or absent nuclei, and few or no granules (Figure 2B). CD41a⁺ LLG MK showed a substantial increase in mitochondrial membrane potential compared to CD41a⁺ SHG MK (Figure 1D), further supporting viability of the former.

Several approaches to characterize the functionality of the LLG and SHG MK were undertaken. The signaling capacity of these different MK populations was assessed and LLG, but not SHG, were able to activate integrin α IIb β 3 in response to agonist stimulation (Figure 3A). In addition, thrombin-induced Ca²⁺ mobilization was observed only in LLG cells but not in SHG cells (Figure 3B). Importantly, by day 13 we observed MK proplatelet formation (PPF). After separating LLG from SHG based on PS exposure, we observed more PPF in PS^{Low} LLG MK than in PS^{High} SHG MK (Figure 3C).

Next, we sought to address whether the LLG population was the origin of the SHG cells. Day 13 PS^{Low} LLG MK

were isolated and labeled with lipophilic cell tracking dye DiO^{18,19} and cultured for an additional 48 hours (h) to follow the 'movement' of the DiO label to SHG MK (Figure 3D). We observed an increase in DiO⁺ LLG MK at day 15 (Figure 3E). More importantly for this experiment, significantly more SHG MK contained tracking dye after an additional two days in culture (Figure 3E). These data indicate that CB-derived MK cultures represent a continuum of cells transitioning from mature viable CD41a^{High} CD42a^{High} PS^{Low} functional LLG MKs into CD41a^{Low} CD42a^{Low} PS^{High} apoptotic SHG MK.

BCL2L2 regulates megakaryocyte apoptosis and larger, lower granular megakaryocyte number

To begin to understand the molecular mechanisms regulating cultured MK apoptosis, we screened 24 genes with established roles in intrinsic and extrinsic apoptosis pathways whose expression changed as MK matured in culture. Because the percentages of SHG MK substantially increased between day 6 and day 13 (*Online Supplementary Figure S4*), we performed gene expression profiling on CD61-purified MK at these time points and observed a 3-fold and 12-fold increase, respectively, in *BCL2L1* and *BCL2L2*, two anti-apoptotic Bcl-2 family members (Figure 4A). *BCL2L1* encodes Bcl-xL and has an established role

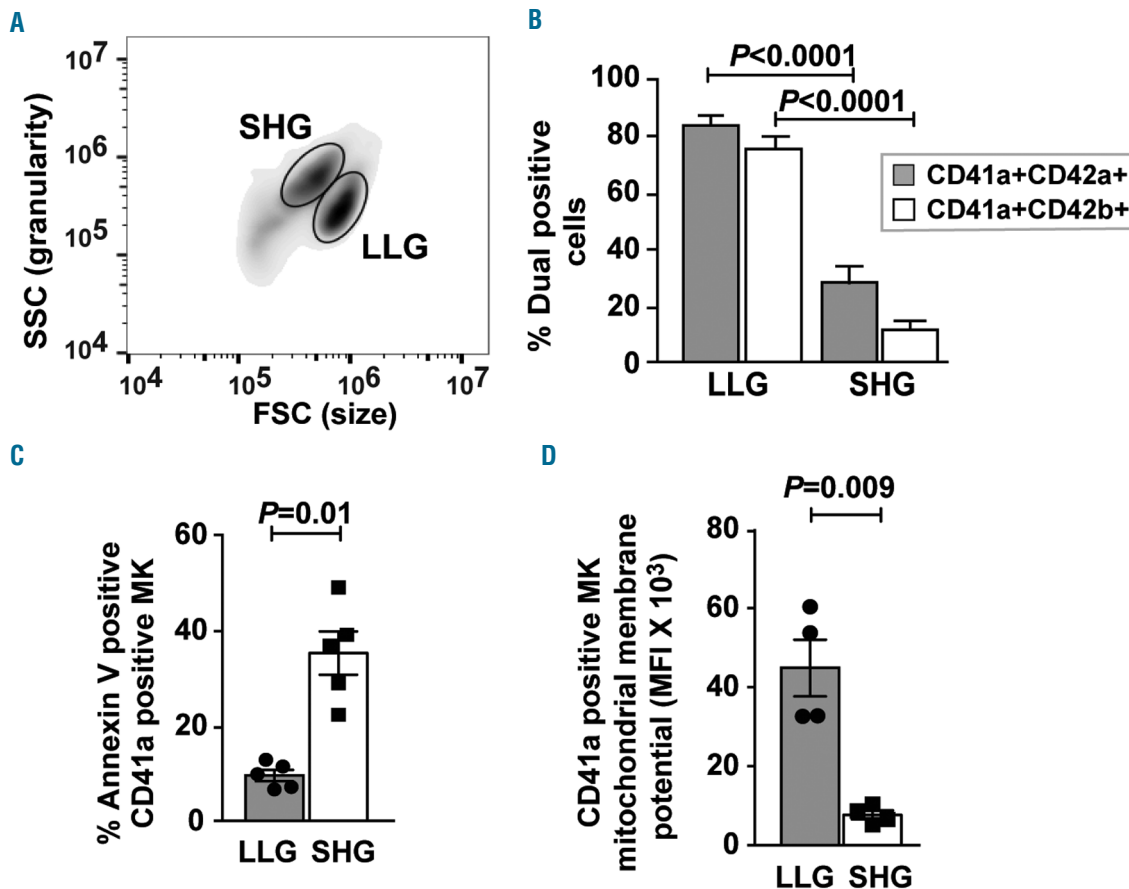


Figure 1. Two distinct populations emerge during cultured megakaryocytopoiesis. (A) Forward scatter (FSC) and side scatter (SSC) flow cytometric analysis of day 13 cultures showing the distinct larger size, lower granularity (LLG) and smaller size, higher granularity (SHG) populations of megakaryocytes (MK). (B) LLG and SHG cells were quantified for dual MK markers CD41aCD42a (n=10) and CD41aCD42b (n=10) by flow cytometry. (C) LLG and SHG MK were analyzed by flow cytometry using PE-labeled annexin V and APC-labeled anti-CD41a (n=5). (D) LLG and SHG MK were stained with APC-labeled anti-CD41a and TMRM to detect mitochondrial membrane potential and analyzed by flow cytometry (n=4). Error bars indicate mean±SEM. All assays performed on day 13 cultures.

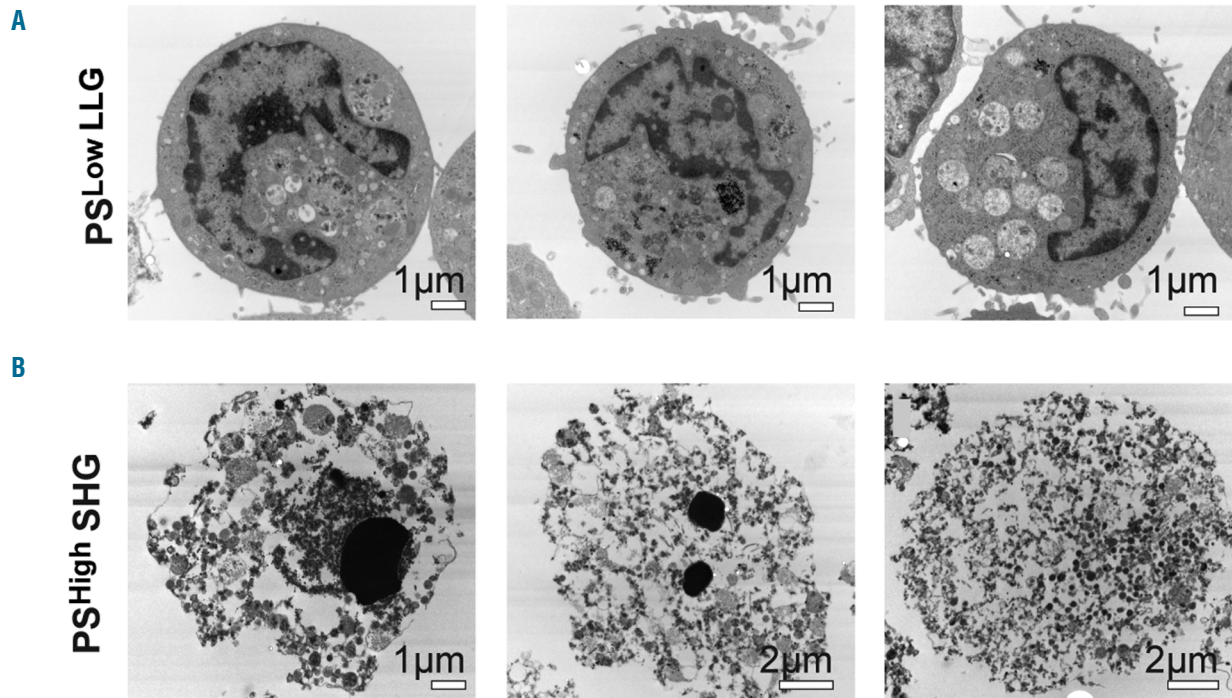


Figure 2. Transmission electron microscopy images of cultured megakaryocytes (MK). (A) Three representative annexin V microbead-isolated PS^{Low} larger size, lower granularity MK. (B) Three representative annexin V microbead-isolated PS^{High} smaller size, higher granularity MK.

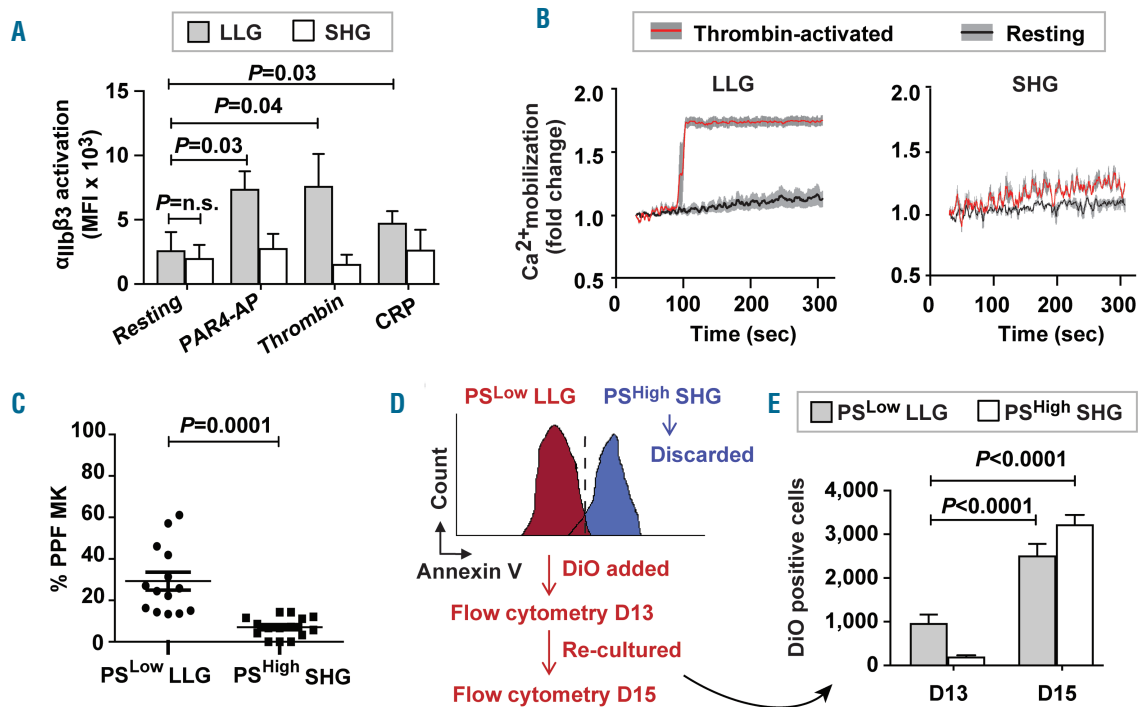


Figure 3. Larger size, lower granularity (LLG) and smaller size, higher granularity (SHG) megakaryocytes (MK) are viable and apoptotic, respectively. Assays performed on day 13 cultures for (A-C). (A) LLG and SHG MK were stimulated with PAR4-AP, thrombin, collagen related peptide (CRP) or no agonist (resting), and PAC1 binding (marker of integrin $\alpha IIb\beta 3$ activation) was quantified by flow cytometry (n=4). n.s.: not significant. (B) LLG and SHG were stimulated with thrombin (red line) or no agonist (resting; black line) and calcium mobilization was measured (gray lines represent Standard Error of Mean for 3 separate experiments). (C) Quantification of proplatelet forming (PPF) MK blinded as to whether samples were PS^{Low} LLG or PS^{High} SHG cells. Data were collected from five independent cords and each data point represents the percentage of PPF MK counted from randomly selected objective field image. An average 200 cells were counted per cord sample. (D) Schematic outlining for DiO labeling experiment. Day 13 PS^{Low} LLG MK were separated from PS^{High} SHG MKs using annexin V microbeads. PS^{Low} LLG MK were labeled with Vybrant DiO cell tracking dye and re-cultured for two days. (E) Bar graph showing numbers of DiO-positive PS^{Low} LLG MKs and PS^{High} SHG MK at day 13 and day 15 (48 hours post DiO addition) (n=6).

in MKpoiesis.^{12,13,20} However, *BCL2L2* (encoding Bcl-w) has not been studied in Mkp0iesis and became the focus of further investigation. The increase in expression of *BCL2L2* observed by RNA sequencing was verified by qPCR on RNA isolated from CD61-purified MK (Figure 4B). This led to the hypothesis that those MK with increasing levels of anti-apoptotic *BCL2L2* would be the LLG, whereas those MK with stable or decreasing levels of anti-apoptotic *BCL2L2* would be the SHG. LLG MK and SHG MK were purified by cell sorting and analyzed for changes in *BCL2L2* expression over time in culture. There was no difference between LLG and SHG cells for *BCL2L2* expression at days 6 and 9, but a significant difference was observed by day 13 due to a fall in *BCL2L2* in SHG cells and concomitant rise in LLG cells (Figure 4C).

We next considered the effects of *BCL2L2* on cultured MK apoptosis and number. Initially, we used the general Bcl-2 family inhibitor, ABT-263, to screen for an effect on CD41a⁺ LLG MK numbers, and observed a significant reduction (Online Supplementary Figure S5). Since ABT-263 inhibits all Bcl-2 family members, we specifically tested the effects of *BCL2L2* on the cultured MK. CB-derived MK cultures were transduced with lentiviral vectors containing *BCL2L2*, and both mRNA and protein increased by day 13 (Figure 5A-C). *BCL2L2* overexpression significantly reduced the percentage of annexin V⁺ CD41a⁺ MK (Figure 5D) and increased the number of CD41a⁺ LLG MK by 19% (1.36×10^5 to 1.61×10^5 ; $P=0.049$).

BCL2L2 regulates megakaryocyte proplatelet formation

Megakaryocyte PPF is believed to be a critical process in thrombopoiesis. By day 13 we observed MK PPF primarily in LLG MK (Figure 3C), and asked whether Bcl-w might affect PPF. Importantly, when we scored MK PPF blinded to lentiviral transduction treatment group, we found that *BCL2L2* overexpression induced a significant 58% increase in PPF MK (Figures 5E and F).

Relationship between BCL2L2 expression and platelet number

Megakaryocyte PPF is tightly linked to PB platelet count,^{10,21,22} raising the possibility that *BCL2L2* may regulate platelet number as well as MKpoiesis. Because we had previously performed genome-wide platelet gene expression profiling in 154 healthy individuals,¹⁴ we were able to query this dataset for such an association. Platelet *BCL2L2* mRNA levels were positively correlated with platelet count in the PRAX1-1 study (Figure 6A).

As is typical of MK culture systems, the day 13 cultures also contained a population of small, hypogranular particles that overlapped the forward and side scatter properties of normal human platelets (Figure 6B). These particles are typically referred to as platelet-like particles (PLP), and we assessed the effect of MK *BCL2L2* overexpression on PLP number and function. Compared to control lentiviral transduction, *BCL2L2* overexpression induced a modest but significant increase in CD41a⁺ PLP from approximate-

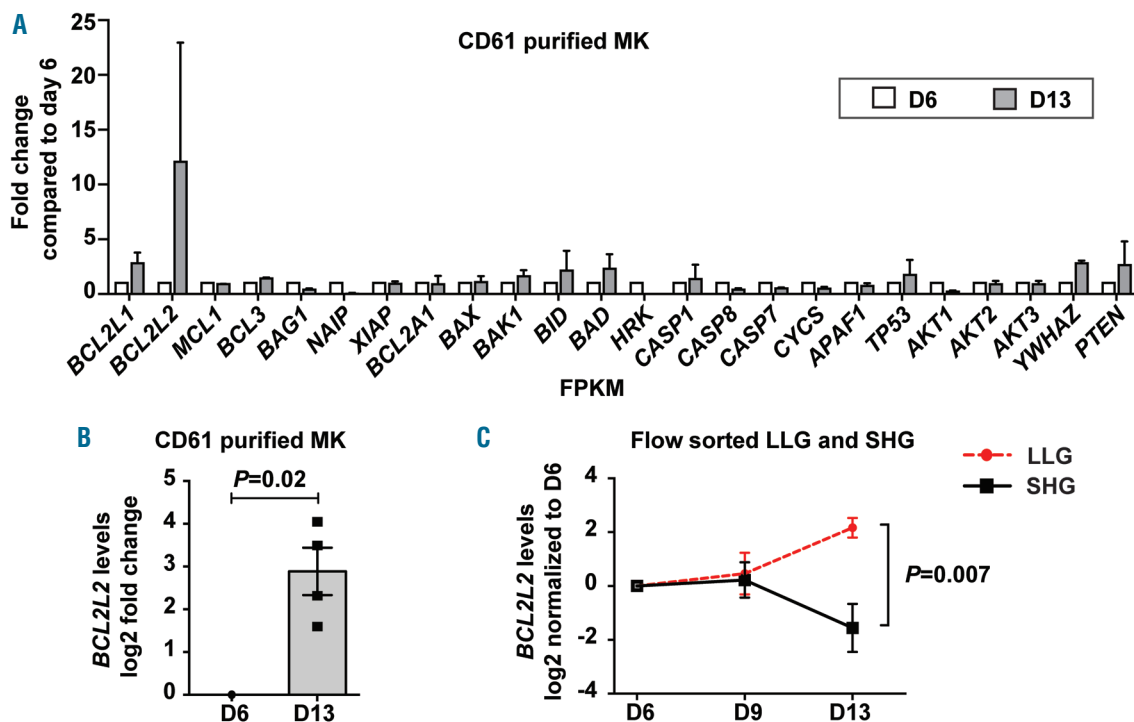


Figure 4. Expression of the anti-apoptotic gene *BCL2L2* increases functional megakaryocytes (MK). CD61⁺ MK RNA was isolated from day 6 and day 13 cultures for analyses in (A) and (B). (A) CD61-positive (*) MK expression of 24 apoptosis-related genes was assessed by RNA sequencing (n=3). Average fold changes in expression for day 13 versus day 6 are shown. (B) CD61⁺ MK *BCL2L2* quantification by quantitative polymerase chain reaction (qPCR) (n=4). (C) Larger size, lower granularity (LLG) and smaller size, higher granularity (SHG) MK were flow cytometrically sorted and *BCL2L2* expression was quantified by qPCR. Fold changes were determined for LLG and SHG MK compared to their respective day 6 expression (n=4 for day 6 and day 13; n=6 for day 9).

ly 10% to approximately 19% (Figure 6C), suggesting *BCL2L2* enables mature platelet production.

Platelet-like particles were stimulated with thrombin to assess their ability to activate α IIb β 3 and release α -granules. PLP from both control and *BCL2L2* lentiviral constructs demonstrated cell activation above background (Figure 5D and E, and flow cytometry plots in *Online Supplementary Figure S6*); the signal was small but consistent in PLP with control lentivirus. Importantly, among these CD41a⁺ and CD42a⁺ PLP, MK *BCL2L2* overexpression enhanced thrombin-induced activation compared to control lentivirus (Figure 6D and E, and flow cytometry plots in *Online Supplementary Figure S6*). PAC1 binding and P-selectin expression in PLP treated with buffer instead of thrombin were not altered by *BCL2L2* overexpression (*data not shown*).

Discussion

There has been exciting progress in the generation of MK and platelets *via* the use of induced pluripotent stem cells, immortalized MK cell lines, and bioreactors,^{1,23,24} but cultured human CB MK remain an important tool for gaining a deeper understanding of MKpoiesis and platelet production. Apoptosis of MK in a culture system remains a major obstacle to progress in the *in vitro* generation of MK and platelets.⁶ A great deal of our understanding about the apoptosis genes regulating MKpoiesis and PB platelet

number is derived from work in murine systems,^{12,20,25-27} while relatively less is known about *in vitro* cultured MK apoptosis. The major finding in the current study is that the pro-survival gene *BCL2L2* restrains apoptosis in cultured human MK, regulates PPF, and is associated with platelet number in healthy humans. We also found that suspensions of CB-derived MK are a useful system for assessing candidate gene function by flow cytometric analysis, and that *BCL2L2* overexpression induced increases in agonist-induced signaling responses in PLP. *BCL2L2* thus becomes another potential target for enhancing MK yields *in vitro*, which may benefit both basic research on MKpoiesis and the long-term goal of producing platelets for transfusion into patients.

Two distinct populations of cells invariably emerged during our CB MK cultures that were easily distinguished by flow cytometry logarithmic scale forward and side scatter measures. Electron microscopic analysis of the LLG MK (Figure 2) demonstrated they were most similar to primary BM MK, and resembled those observed by Cramer *et al.*²⁸ and Chantelot-Bellanne *et al.*²⁹ In contrast, the SHG population represented primarily apoptotic cells resembling the senescent MK observed by Radley *et al.*, where the nucleus fragmented but the cells appeared to change shape and 'round up' rather than fragment.³⁰ Membrane degradation during apoptosis likely contributed to the loss of MK markers (Figure 1B). The most important functional differences between LLG and SHG were the ability to form proplatelets and 'signal' in response to platelet ago-

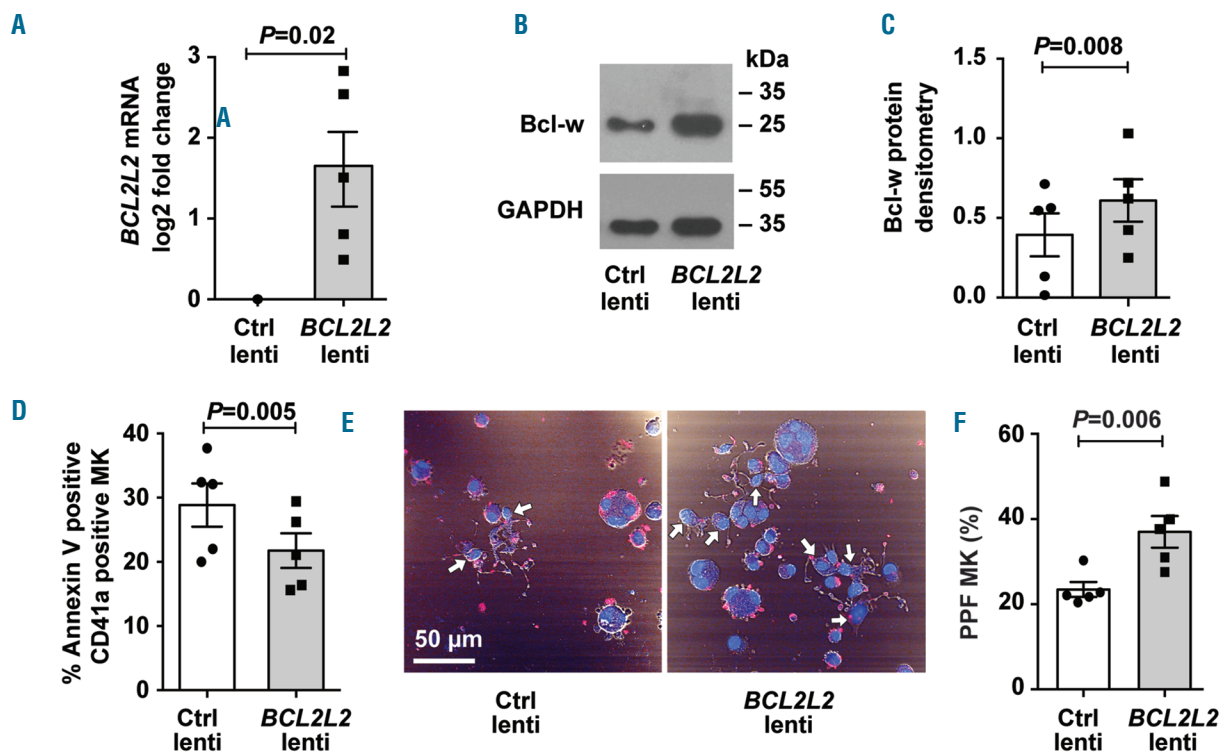


Figure 5. *BCL2L2* overexpression reduces megakaryocyte (MK) apoptosis. (A-F) Transduction of MK cultures with empty lentiviral vector (Ctrl lenti) or lentiviral vector containing *BCL2L2*. (A) Overexpression of *BCL2L2* mRNA was confirmed by quantitative polymerase chain reaction (qPCR). Fold change of *BCL2L2* mRNA levels was determined compared to Ctrl lenti (n=5). (B) Representative western blot image for Bcl-w protein (encoded by *BCL2L2* gene) for *BCL2L2* overexpression compared to Ctrl. (C) Densitometric analysis of western blots for *BCL2L2* overexpression compared to Ctrl (n=5). Glyceraldehyde-3-phosphate dehydrogenase (GAPDH) was used as a normalizer for densitometric analysis. (D) The percentage of annexin V-positive (*) CD41a⁺ MK was determined for *BCL2L2* over-expressed and Ctrl cells by flow cytometry (n=5). (E) Representative image of proplatelet formation in Ctrl and *BCL2L2* over-expressed cells. Image shows α -tubulin staining (red) and nuclear stain, DAPI (blue). White arrows show proplatelet-forming MK. (F) Quantification of proplatelet forming (PPF) MK amongst total cells blinded to treatment group (n=5).

nists (Figure 2A-C). It is worth noting that although PS-positivity marked apoptotic SHG MK, some fraction of LLG MK were also capable of PS exposure in response to thrombin stimulation (*Online Supplementary Figure S7*), perhaps akin to collagen and thrombin activated (COAT) platelets.³¹⁻³³ Thus, simple gating on flow cytometric forward and side scatter measurements, without the need for fluorescently-labeled antibodies, identified functional MK and can improve signal-to-noise read-outs that test the functionality of candidate MK/platelet genes.

Apoptosis was established as a mechanism for platelet lifespan by the Kile laboratory when they showed that the balance between Bcl-xL and Bak regulate murine lifespan *in vivo*.²⁰ The pro-survival Bcl-2 family member Bcl-xL degrades in aging platelets, thus allowing apoptosis to proceed. In addition to Bcl-xL, the Bcl-2 family includes Bcl-2, Mcl-1 and Bcl-w, which are encoded by *BCL2*, *MCL1* and *BCL2L2*, respectively. Altered murine expression of pro-survival Bcl-2 family members has shown variable effects on platelet number. Global chimeric vav-Bcl2 overexpression resulted in normal MK numbers but an approximately 50% reduction in platelet counts,³⁴ whereas MK-specific deletion of *Bcl2* had no effect on platelet number.³⁵ MK-specific deletion of *Bcl2l1* deletion caused MK apoptosis, loss of platelet shedding, and a macrothrombocytopenia.¹² MK-specific deletion of *Mcl1* did not affect MK number or morphology, or platelet count or volume.¹³ Print *et al.* globally inactivated *Bcl2l2* to study its importance in murine physiology.³⁶ Although Mkpoiesis was not a major focus of their study, they reported that three

Bcl2l2^{-/-} mice displayed numbers of MK colony-forming cells and platelets comparable to three wild-type littermates. This latter report appears to differ from our finding in primary human MK and platelets, where *BCL2L2* increased during MK differentiation, overexpression of *BCL2L2* increased the numbers of both the viable CD41a⁺ LLG MK and MK PPF (Figure 4E and F), and *BCL2L2* levels correlated with platelet number (Figure 5A). This apparent discrepancy between human and mouse regarding the effect of *BCL2L2* on MKpoiesis and platelet production could be due to a compensatory upregulation of other Bcl-2 family members in the *Bcl2l2* null mice. Alternatively, small numbers of mice and different conditions may have led to a chance finding in the Print *et al.* study,³⁶ but there also may be species differences in the relative importance of *BCL2L2* because human platelets contain 3.6-fold higher levels of *BCL2L2* transcripts than mouse platelets.³⁷

We showed that the SHG MK were derived from the LLG MK (Figure 2D and E), supporting a process by which viable LLG avoid apoptosis long enough to mature and acquire mature MK markers, but an unknown trigger induces some cells to become dying SHG. The day 9 to day 13 increase in *BCL2L2* is expected to restrain apoptosis in LLG cells, whereas *BCL2L2* reduction should enable apoptosis to proceed in SHG cells (Figure 3C). Perhaps a yet-to-be-defined switch regulates *BCL2L2* levels after day 9 in cultured cells. CB-derived LLG MK began to develop proplatelet extension around day 12 and peaked at days 13-15, similar to what had been reported by Balduini *et al.*³⁸ Because SHG appear days before PPF MK (*Online*

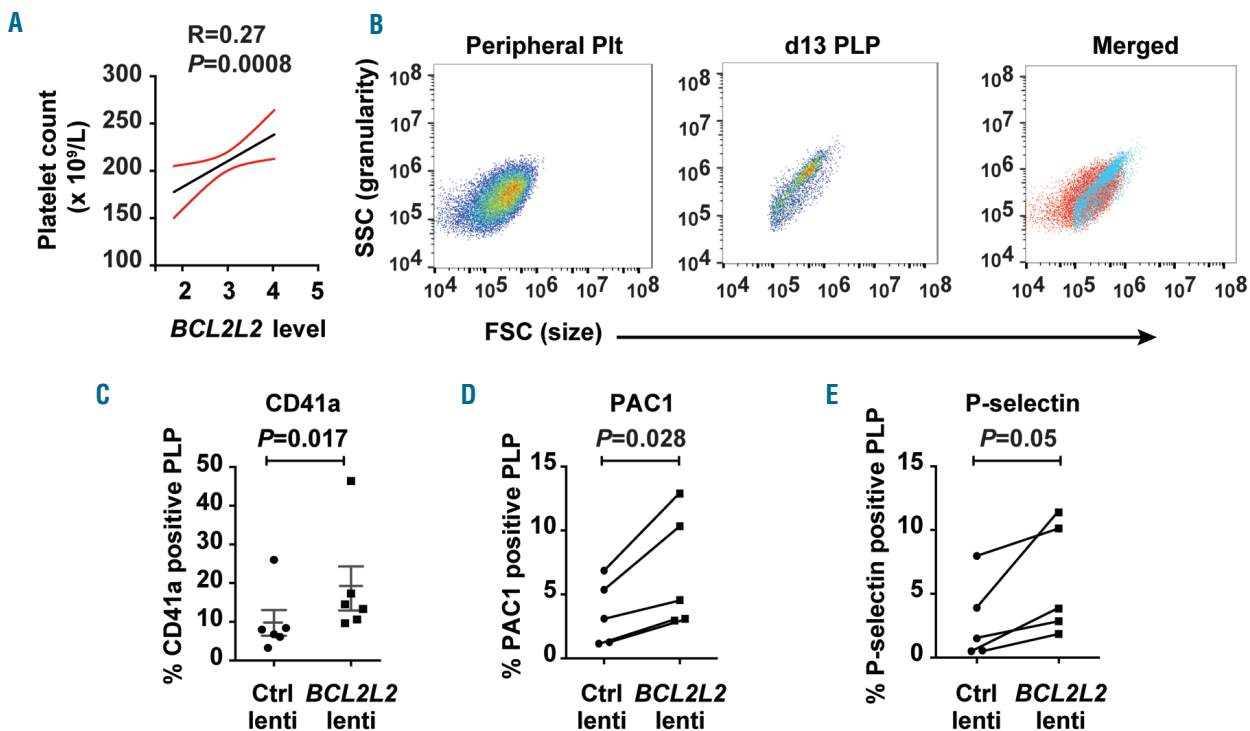


Figure 6. Relationship of *BCL2L2* to blood platelet number and cultured platelet-like particles (PLP). (A) *BCL2L2* is positively associated with peripheral blood platelet counts in 154 healthy individuals in the PRAX1 study. (B) Forward scatter and side scatter flow cytometric analysis of human peripheral blood platelets (Peripheral Plts), day 13 PLP and the merged image. (C-E) Flow cytometric assessment of cultured PLP after MK transduction with empty lentiviral vector (Ctrl lenti) or lentiviral vector containing *BCL2L2*. PLP were defined by forward scatter, side scatter and presence of platelet-specific surface marker (see Methods). (C) The percentages of day 13 PLP that were CD41a-positive (*) (n=6). (D) The percentages of day 13 PLP that bound PAC1 after stimulation with 250 nM thrombin (n=5). (E) The percentages of day 13 PLP that expressed P-selectin after stimulation with 250 nM thrombin (n=5).

Supplementary Figure S4), they do not seem to be 'exhausted' MK post release of proplatelets.

Sim *et al.* used adult mobilized HSPC to culture MK that also gave rise to populations of functional and apoptotic MK.¹⁷ Our work differs from this primarily by studying the molecular mechanism of apoptosis. We are unaware of direct comparisons among any source of MK (neonatal, adult, murine, embryonic stem cells, etc.) in terms of the degree of apoptosis in culture, but it is reasonable to consider that some molecular mechanisms will be shared. In this regard, our data show no statistically significant difference between *BCL2L2* levels in neonatal and adult MK ($P>0.47$), and adult MK also display more PPF with *BCL2L2* overexpression (Online Supplementary Figure S8). As with MK, there is no statistically significant difference between *BCL2L2* levels in neonatal and adult platelets.³⁹

The finding of a significant positive correlation between human platelet *BCL2L2* levels and PB platelet counts (Figure 5A) supports a mechanism whereby Bcl-w increases MK PPF which causes increased blood platelets. Numerous apoptosis genes have been studied for effects on Mkpoiesis.^{8-13,19,40,41} We observed that pro-apoptotic genes *BAX* and *BAK1* showed little to no change in expression from day 6 to day 13 in MK cultures (Online Supplementary Figure S9A and B). However, similar to *BCL2L2*, anti-apoptotic gene *BCL2L1* showed increased expression in LLG over time, and was higher at day 13 in LLG compared to SHG (Online Supplementary Figure S9C). Taken together, these studies suggest *BCL2L1* and *BCL2L2* restrain apoptosis and that this may be the key mechanism for prolonging MK survival in the late stages of differentiation in culture. We cannot exclude a role for other apoptosis genes in Mkpoiesis and platelet production. It is also reasonable to consider that, like Bcl-xL and Bak, Bcl-w may also impact the life span of the human PB platelet.²⁰ However, additional studies are needed to address this possibility.

The *BCL2L2*-induced increase in MK PPF led to experiments assessing production of PLP. *BCL2L2* overexpression roughly doubled the number of CD41a⁺ PLP and increased the ability of these PLP to show a functional response when activated with thrombin (Figure 5C-E). However, we did not observe any significant correlation between *in vivo* *BCL2L2* expression with other platelet

agonists such as ADP, PAR1-AP and PAR4-AP (www.Plateletomics.com, as reported by Simon *et al.*⁴⁰). Nevertheless, we would emphasize that the percentages of functional PLP were small, although consistent and statistically significant.

Our findings have several research and clinical implications. Firstly, we have shown that simple gating on flow cytometric forward and side scatter measurements allows the identification of the subset of viable and functional MK. This approach represents a substantial improvement in the signal-to-noise read-outs to test the functionality of candidate MK/platelet genes identified in GWAS. Secondly, promotion of apoptosis by inhibiting of Bcl-2 family members is under investigation for the treatment of a variety of malignancies;⁴¹⁻⁴³ a limiting toxicity has been thrombocytopenia.^{42,43} Bcl-2 family members, *MCL1* and *BCL2L1* are the most frequently amplified in 26 tumor types,^{44,45} whereas *BCL2L2* amplification is rare.⁴⁶ Perhaps selective targeting of Bcl-2 family members that spares Bcl-w could minimize thrombocytopenia. Lastly, progress toward the manufacture of *in vitro* platelets may benefit from the pro-survival benefits on MK generation and PPF of Bcl-w. *BCL2L2* overexpression may have synergistic or additive effects when used in combination with an antagonist of the aryl hydrocarbon receptor (StemRegenin 1), *BCL2L1* overexpression, abscisic acid, and other factors that have been shown to increase numbers of proplatelet-forming MK in culture.^{1,47,48}

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