appears that such abnormalities are mostly observed during the first two years of TKI therapy. While there is a significant risk of a second myeloid malignancy in patients with -7 CCA/Ph , less than half of these patients will develop MDS/AML. The aggregate data provide evidence in support of the commonly held view that preemptive therapeutic strategies are not justified in all patients with detectable -7 CCA/Ph- . Nevertheless, once a diagnosis of AML is confirmed in these patients, intensive treatment strategies, including allogeneic BM transplantation, are ineffective in most patients. One may speculate on the role of TKI in the mechanism of MDS development and the presence of -7/del(7q) CCA/Phabnormalities. The mutagenic effect of TKI on hematopoietic stem cells is not yet fully understood. However, it has been reported that a gastrointestinal stromal tumor patient developed MDS with monosomy 7 during imatinib treatment, suggesting that imatinib plays a direct role in causing MDS.12

The routine monitoring of CML patients is currently molecular assessment of the response. However, cytogenetic analysis is still relevant and should be performed with a BM smear certainly in cases of cytopenia during TKI therapy. Signs of dysplasia with -7 CCA/Ph- cells should be considered as a red signal and a switch to alternative treatment be discussed.

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Another piece of the puzzle added to understand t(4;11) leukemia better

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The story about t(4;11) leukemia, involving the
 MLL/KMT2A gene from chromosome 11q23.3 and

the *AF4/AFF1* gene from chromosome 4q21, is still

a mystery. The study by Agraz-Doblas *et al.*, published in *MLL/KMT2A* gene from chromosome 11q23.3 and a mystery. The study by Agraz-Doblas *et al.*, published in this issue of *Haematologica*, adds some new and important information regarding the mysterious pathomechanism.¹ Agraz-Doblas *et al*. showed, for the first time, that the therapeutic outlook of patients with expression of both reciprocal MLL fusions, MLL-AF4 and AF4-MLL, is promising, but only 50% of the investigated patients seem to have this favorable condition; patients expressing only the *MLL-AF4* allele have an event-free survival of 10% and an overall survival of 30%. Moreover, only leukemic cells expressing both fusion alleles display the typical *HOXA* signature.

The fact that t(4;11) patients can be divided into two subgroups on the basis of *HOXA* transcription was first recognized by Trentin et al. in 2009,² and later confirmed by Stam *et al.* and Kang *et al.* in 2010 and 2012, respectively.3,4 The missing *HOXA* transcription was correlated with overexpression of either IRX1 or IRX22,4 and a 3-fold higher relapse rate^{3,4} Experimental overexpression of IRX1 revealed an interesting mechanism because it resulted in EGR1-3 expression.⁵ EGR1 and EGR2 both control the $p21^{\text{CIP1}}$ gene and, thus, shut down the cell cycle and may even induce cellular quiescence, a known mechanism of resistance to treatment. CDK6 counteracts the actions of EGR proteins.⁶ The second mechanism involves the IRX proteins, which are able to turn on HOXB4, a known stem cell marker of hematopoietic cells that activates factors such as TAL1, GATA factors, TGFB1, etc. Thus, expression of MLL-AF4 alone - with upregulated IRX proteins but without HOXA expression - may provoke treatment resistance or a stem cell-like mechanism which is

not possible when AF4-MLL is present. This could be a rational explanation for the observed clinical behavior of both groups of patients.

Another explanation could lie in the recent findings from Yokoyama's laboratory.7,8 Okuda *et al.* elegantly showed that one of the functions of the pSer domain of the AF4 protein⁹ - which is fused to the N-terminus of MLL in the MLL-AF4 fusion protein - is recruitment of the SL1 complex. The SL1 complex is usually bound to RNA Pol I, which is present in the nucleolus and required for the transcription of rRNA genes. SL1 is artificially recruited to MLL-AF4, but not to native MLL or AF4 complexes.10 This powers up MLL-AF4 leading to a strong increase in gene transcription. The simple presence of the MLL-AF4 fusion protein causes a condition of severe stress, because it compromises protein biosynthesis, and cells may therefore easily display a phenotype of growth arrest or senescence.¹¹ This is probably one of the reasons why it is so complicated to generate a true MLL-AF4 mouse model, and why so many laboratories have failed so far. With the exception of a recent study in which a hybrid between human MLL and mouse Af4 was used,¹² no-one had been able to develop a satisfactory model with only the human MLL-AF4 fusion. Since the sequences of the pSer domains of human and mouse AF4 differ slightly, it is very plausible that the human/mouse chimeric MLL-Af4 is unable to attract the SL1 component and thus does not impair protein biosynthesis. This needs to be tested in future experiments.

So, what is the precise role of AF4-MLL? AF4-MLL has been shown to strongly enhance gene transcription by overwriting the transcriptional elongation control.^{10,13} This massive increase in gene transcription (3- to 4-fold more mRNA) may help its molecular counterpart, MLL-AF4, to set the programming of its target genes, even under conditions of nucleolar stress (see Figure 1). In addition, we have shown by ATAC sequencing in two independent cell lines that AF4-MLL strongly activates chromatin in a very short time frame (*unpublished data from our laboratory*). The expression of AF4-MLL for 48 h was sufficient to open up the chromatin of all chromosomes apart from the centromeric regions and to massively increase gene transcription. Thus, the presence of AF4- MLL would allow the expression of any gene of interest, and increases the plasticity of the tumor cells. According to the data shown by Agraz-Doblas *et al.* this results in

Figure 1. Proposed fuctions of both t(4;11) fusion proteins. MLL-AF4 binds to Menin1/LEDGF and SL1 to target gene promotors and strongly activate gene transcription, while AF4-MLL overwrites transcriptional elongation control and strongly activates chromatin within a very short time window. Patients who express both fusions diplay HOXA gene signatures and have a better outcome, while patients who either do not express AF4-MLL or actively repress it, usually activate the homeobox proteins IRX1/2. Expression of both proteins has been correlated with a worse outcome (IRX1/2 strongly activate EGR1-3 and HOXB4).

cells that are more vulnerable to chemotherapy because the presence of AF4-MLL was associated with a much better treatment outcome.¹

Lastly, expression of AF4-MLL alone was shown to be necessary and sufficient to cause acute leukemia.14 Our group used a low expression retroviral vector backbone (PIDE vector) to express MLL-AF4, AF4-MLL or both in LSK cells purified from C57BL/6 mice. Empty vector or MLL-AF4 alone did not result in the development of leukemia, while AF4-MLL or the expression of both fusion genes resulted in full-blown pro-B-acute lymphoblastic leukemia and mixed-lineage leukemia. The latency was 9 months and the penetrance was only 35%. However, this could be attributed to a low infection rate with about 1/1,000 cells for the AF4-MLL and 1/10,000 cells for the MLL-AF4 construct because these constructs were oversized for *in vitro* packaging (11.3 kb for MLL-AF4 and 13.3 kb for AF4-MLL). Therefore, an estimated 200 cells in 200,000 non-tranduced cells were transplanted into primary mice, which nevertheless caused a disease outbreak (MLL-AF4 with only 20 cells did not work). It is noteworthy that all "leukemic cells" subsequently tested positive for the transcription and integrity of the appropriate transgenes, while the investigated white blood cells of mice who did not develop leukemia remained negative in reverse transcriptase and genomic polymerase chain reaction experiments. This indicates that the leukemia-negative mice either never received or lost the cells carrying the corresponding transgene (negative selection of MLL-AF4 alone).

We, therefore, assume that, in humans, AF4-MLL and MLL-AF4 are both necessary, but AF4-MLL could presumably be shut off after "preparing the ground" for MLL-AF4, and that this process of shutting down AF4- MLL makes the leukemic disease even more aggressive (positive selection). This explains in part the molecular situation diagnosed in human patients with leukemia, regardless of whether they are infants or adults. It would be of interest to compare primary diagnostic material with relapsed material from the same patient, and determine whether AF4-MLL expression is lost in the relapse, in order to have another argument in favor of the above mentioned hypothesis.

The study by Agraz-Doblas *et al.* adds another, important piece to the puzzle of the molecular mechanism of $t(4;11)$ leukemia.¹ It is to be hoped that the precise mechanism of this disease can be understood soon, because the full picture is needed in order to develop new drugs that can really help patients with t(4;11) leukemia.

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