# NKG2D-based chimeric antigen receptor therapy induced remission in a relapsed/refractory acute myeloid leukemia patient

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# Supplementary information for:

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# **Supplementary Figures**



**Fig. S1: Patient's CYAD-01 phenotype and** *in vitro* **functionality.** (A) A sample of the infusion product was analyzed for cell subset composition and memory phenotype. CD3<sup>+</sup> viable singlet cells showed a high CD8 preponderance with both CD4 and CD8 subsets possessing a majority of CD62L<sup>10</sup> phenotype. As shown in the CD8 subset, both CD62L<sup>hi</sup> and CD62L<sup>10</sup> populations were CD95<sup>+</sup> and CD28<sup>10</sup>. Together this suggests the infusion product to be composed primarily of CD8<sup>+</sup> T cells that possess an effector memory phenotype. 88% of the CD4<sup>+</sup> T cells and 94% of the CD8<sup>+</sup> T cells were NKG2D+. (B) Patient's CYAD-01 product was incubated in the presence or absence of NKG2D blocking antibody (CD314 Ab) (light and dark grey bars, respectively) with PANC-1 and K562 cancer cells (at a 1:1 ratio). After 24h of incubation, supernatants were harvested and analyzed for IFN-γ secretion. Dotted line represents the limit of detection (LOD). Each bar represents the mean and SD of one experiment conducted at least in duplicate, \*\*\*\*: p < 0.001. (C) Patient's CYAD-01 cells were cultured at a 1:1 ratio with PANC-1 cells in the presence or absence of CD314 blocking Ab. After a 20h

incubation the CYAD-01 cells were washed away and the remaining PANC-1 cells stained with AlamarBlue to quantify the remaining proliferating PANC-1 cells. Each bar represents one experiment conducted in triplicate with the corresponding SD, \*: p < 0.05.



**Fig. S2: Immunohistochemistry analysis of NKG2D ligand expression in bone marrow sample taken before first CYAD-01 injection.** One BM biopsy was subject to a range of IHC staining protocols specific for the following NKG2DL: MICA/MICB, ULBP1, ULBP2/5/6, ULBP3. In addition, a sample of each was prepared applying standard hematoxylin and eosin (H&E) staining (data not shown). All samples were examined and graded by light microscopy. Representative stainings of ULBP1 (A), ULBP2/5/6 (B) and ULBP3 (C). It has to be noted that ULBP2/5/6 are predominantly stained in the nucleus and that ULBP1, ULBP3 and MICA/B (Fig. 2B) display cytoplasmic staining as well.

#### **Supplementary Methods** 1

#### 2 Study THINK design

3 The THINK (THerapeutic Immunotherapy with NKR-2) trial is an open-label Phase I study which primarily aims to assess the safety and clinical activity of the CYAD-01 treatment 4 5 administered three times at 2 weeks intervals between each administration without prior lymphodepleting chemotherapy in patients with refractory or relapsing malignancies, including 6 7 patients with metastatic or locally advanced colorectal cancer, urothelial carcinoma, triple-8 negative breast cancer, pancreatic cancer, recurrent epithelial ovarian and fallopian tube 9 carcinoma, AML/MDS or MM. The study is split into two segments; a dose escalation segment evaluating three dose-levels  $(3x10^8, 1x10^9 \text{ and } 3x10^9 \text{ cells/injection})$  to determine the 10 11 recommended dose of CYAD-01 cells and an expansion phase to investigate the clinical activity 12 across multiple tumor indications while extending the safety study.

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## **Manufacture of cell products**

14 CYAD-01 (previously known as NKR-2) refers to the viable cell population obtained after 15 retroviral transduction of autologous T-cells with the NKG2D-based CAR. CYAD-01 will be 16 supplied cryopreserved in bags containing a T-cell dose in accordance with the dose-level 17 which is to be administered.

#### **Characterization of the patient CYAD-01** 18

CYAD-01 identity (% NKG2D on CD4<sup>+</sup>, CD4<sup>+</sup>CD8<sup>+</sup> and CD8<sup>+</sup> T cells), purity (% viable CD3<sup>+</sup>) 19 20 and viability (with 7AAD dye) are assessed by flow cytometry. Cell yield is assessed by cell 21 counting (exclusing Trypan blue). In vitro product functionality/potency is evaluated by 22 assessment of IFN-y secretion via ELISA and metabolic activity of tumor cells via Alamar Blue 23 assay upon co-culture of CYAD-01 with NKG2D ligand-expressing tumor cells (Panc-1 cells

1	and /or K562). NKR-2 microbiological safety was confirmed by absence of microbiological
2	growth, assessed by BactAlert, absence of mycoplasms, assessed by qPCR based MycoTool
3	assay, and compliant endotoxin level (<8.67 EU/ml), assessed by PTS EndoSafe LAL assay.
4	Patient CYAD-01 met the product specifications. CYAD-01 phenotype identity (90%
5	NKG2D <sup>+</sup> ), purity (95% CD3 <sup>+</sup> ), viability (86%) and cell yield (300 x $10^6$ cells), evidenced a
6	viable, highly pure CD3 <sup>+</sup> , NKG2D <sup>+</sup> product. Viral vector safety was evaluated by Vector copy
7	number (VCN) and Replication copy number (RCR) qPCR-based assays.In vitro product
8	functionality/potency was confirmed by IFN- $\gamma$ secretion and efficient cytotoxicity effect in
9	response to NKG2DL on tumor cells.