SUPPLEMENTARY APPENDIX

Prognostic impact of the absence of biallelic deletion at the *TRG* locus for pediatric patients with T-cell acute lymphoblastic leukemia treated on the Medical Research Council UK Acute Lymphoblastic Leukemia 2003 trial

Nadine Farah, ¹ Amy A Kirkwood, ² Sunniyat Rahman, ¹ Theresa Leon, ¹ Sarah Jenkinson, ¹ Rosemary E. Gale, ¹ Katharine Patrick, ³ Jeremy Hancock, ⁴ Sujith Samarasinghe, ⁵ David C. Linch, ¹ Anthony V Moorman, ⁶ Nicholas Goulden, ⁵ Ajay Vora^{3,5} and Marc R. Mansour

¹Department of Haematology, UCL Cancer Institute, London; ²Cancer Research UK & UCL Cancer Trials Centre, UCL, London; ³Department of Haematology, Sheffield Children's Hospital; ⁴Bristol Genetics Laboratory, Southmead Hospital; ⁵Department of Haematology, Great Ormond Street Hospital and ⁶Leukaemia Research Cytogenetics Group, Northern Institute for Cancer Research, Newcastle, UK

Correspondence: m.mansour@ucl.ac.uk doi:10.3324/haematol.2017.185801

SUPPLEMENTARY INFORMATION

Patient cohort

Diagnostic samples from previously untreated T-ALL patients enrolled on the MRC UKALL2003 trial were analyzed. The trial, registered at http://www.controlled-trials.com under the ISRCTN number 07355119, opened in October 2003 for patients aged 1-18 years. The upper age limit of the trial was increased in September 2006 to 20 years and in June 2008 to 25 years of age. Samples obtained at diagnosis from 152 of the 393 (39%) patients with T-ALL were available for analysis. Ethical approval for the trial was obtained previously from the Scottish Multi-Centre Research Ethics Committee, and samples were collected with informed consent according to the Declaration of Helsinki.

Details of the trial protocol are as published¹ and are outlined in Supplementary Figures S5-6. At trial entry, patients with National Cancer Institute (NCI) standard risk (<10 years of age and white cell count [WCC] <50x10 9 /L) were assigned to regimen A while NCI high risk patients (≥10 years and/or WCC ≥50x10 9 /L) received regimen B. Patients <16 years with a slow early response (≥25% blasts in the day 15 or 8 bone marrow for regimens A or B respectively) and all patients with high risk cytogenetics (*KMT2A* [*MLL*] and *TCF3-HLF* fusions, near haploidy, low hypodiploidy, and iAMP21) were assigned to regimen C. MRD was evaluated by real-time quantitative PCR analysis of immunoglobulin and T-cell receptor gene rearrangements with a quantitative range of 0·01% as defined by the European MRD Study Group.² Patients with undetectable MRD at the end of induction (EOI, day 29) and before interim maintenance were classified as MRD low risk, as were

those who had detectable EOI MRD (<0.01%) but undetectable MRD before the start of interim maintenance. MRD low risk patients were eligible for treatment reduction randomization. Patients with EOI MRD \geq 0.01% were classified as MRD high risk and were eligible for treatment intensification randomization (Figure S5).

Materials and Methods

Samples

Whole-Genome-Amplified (WGA) diagnostic genomic DNA (gDNA) from patient samples was as previously prepared.³

qPCR assay

The quantitative polymerase chain reaction (qPCR) assay was as previously described,⁴ and evaluated the presence or absence of an amplicon within the intron between the V and J regions of the *TRG* locus at chromosome 7p14. The primers (Forward: 5'-CATCCTCACTTTCCTGCTTCTTC-3'; Reverse: 5'-CCAAGGTGAATCCCTACATGCT-3') amplified an 87 base pair (bp) amplicon 5089 bp from the 3' end of the *TRGV11* pseudoexon and 10123 bp away from the 5' end of *TRGJP1* exon. The reference gene *ANLN* lies 1.9Mb downstream of *TRG* at 7p15-14, and the primers (Forward: 5'-AAATTCTGCCCTTTGCTTGTTT-3'; Reverse: 5'-GAAAGCAACCACAGAGAATATGTAAGTAA-3') amplified an 89 bp product.

25μl PCR reactions were set up as follows: 0.5μl of each forward and reverse primer (either *TRG* or *ANLN*) was added at 0.2μM, 12.5μl 2x concentrated ready-to-use FastStart Universal SYBR Green Master reaction mix (ROX) (Roche, 2008), 2μl genomic DNA template, and 9.5μl nuclease-free water. Samples were analyzed

in triplicate for each primer pair. The reactions were run on a Mastercycler epgradient S thermocycler (Eppendorf) at 95°C for 10mins, followed by 40 cycles of 95°C for 15 seconds and 60°C for 1 min for each cycle. CT (cycling time) values were obtained from the Mastercycler ep Realplex software.

Standard curves using dilutions of gDNA from the non-leukemic cell line HEK293T that does not have a rearrangement of the *TRG* locus showed that the primer pair efficiencies were very similar to each other (E=1.84 and 1.817 for *TRG* and *ANLN* respectively). The *ANLN* primer pair efficiencies were also validated using gDNA from Jurkat cells, a non-early T cell precursor cell line that does have a rearrangement of the *TRG* gene.

Mean and standard deviation (SD) between the CT values of each sample for each primer pair were calculated. If the SD between CT values of the replicates for the PCR reaction of the reference gene, *ANLN*, was >0.5, then the reactions were discarded and all the reactions for that patient sample were repeated.

TRG:ANLN fold change was calculated according to the comparative $\Delta\Delta$ CT method using HEK293T gDNA as the calibrator. Each run included the positive control (HEK293T gDNA) and a negative control (nuclease-free water) for each primer pair. The SD of the HEK293T CT values for each of the primer pairs showed minimal variability across the runs (n=14; *TRG*: mean CT value 23.7, range 23.4-23.9, SD 0.1; *ANLN*: mean CT value 23.8, range 23.6-23.9, SD 0.1). Therefore, the CT results of the HEK293T sample of each run was used as the calibrator value for the fold change calculations for samples on that run.

Comparison of WGA and non-WGA gDNA fold change

TRG:ANLN fold change calculated by the method above using WGA gDNA samples was validated using the same assay in the corresponding non-WGA gDNA samples available from 26 patients, which included patients with fold changes that covered the entire range (mean fold change 0.01–0.94). There was good agreement between the fold change results from the 2 types of samples (R²=0.92; Figure S2A), with a bias of -0.023 and an agreement interval from -0.201 to 0.155, in which 95% of the differences between the 2 fold changes should lie (Figure S2B). One patient with ABD and three patients with non-ABD in the WGA samples were found to be indeterminate by qPCR assay in the corresponding non-WGA samples. However, none of the patients had their ABD status change from ABD to non-ABD, and viceversa, when comparing the WGA and the corresponding non-WGA results.

Validation of fold change using a different reference gene

The fold change results that were indeterminate by the qPCR assay above (TRG:ANLN fold change 0.26-0.49) were validated using a different reference gene encoding the mitochondrial protein Cytochrome c Oxidase Assembly factor 1 (COA1), which lies 4MB upstream of the TRG locus, to abrogate the possibility that abnormalities of the ANLN reference gene itself may have contributed to the indeterminate fold change calculation. The COA1 primers (Forward: 5'-GGAAAACTGGGTTGCAGGAG-3'; Reverse: 5'-AGAAGACCCAGCTTGCTTCT-3') amplified a 105bp product.

TRG and COA1 assays were performed in triplicate using the same PCR reaction reagents, calibrator and conditions as described above and the TRG:COA1 fold

change calculated. Standard curves using dilutions of gDNA from HEK293T cells were set up using the *COA1* primers and the efficiency was comparable to that of the *TRG* primers (E=1.96 and E=1.84 for *COA1* and *TRG* respectively).

The TRG:COA1 fold change led to the same ABD status assignment as that of the TRG:ANLN fold change for 7 patients that had informative TRG:ANLN fold change results (data not shown). The TRG:COA1 fold change results for 15 patients with indeterminate TRG:ANLN did not change the ABD assignment from ABD to non-ABD or vice versa, although 3 patients had fold changes in this assay that varied from those of the TRG:ANLN assay only across the thresholds of 0.26-0.49 of the indeterminate range (Figure S2C). As a result of the inconclusive fold change results from both the TRG:ANLN and TRG:COA1 assays for these samples, these patients were assigned the ABD indeterminate status.

SNP array analysis

WGA gDNA samples were previously analyzed using the Infinium CytoSNP-850K Beadchip array (Illumina, Essex, UK).³ Log-R intensities and B-allele frequencies for each of the 62 SNP markers across the region hg19 chr7:38288270-38385938 which includes the *TRG* locus were assessed independently by 4 individuals and the *TRG* locus deletion status was scored as no or heterozygous deletion (ABD), or homozygous deletion (non-ABD).

Statistical Analysis

Survival curves were plotted using Kaplan-Meier analysis for overall survival (time from the start of treatment until death), event-free survival (time to relapse,

secondary tumor or death, whichever came first) and relapse-free survival (time to relapse in those who achieved remission). Patients who died in remission were censored. Those who did not have an event were censored at the date last seen. Comparisons between groups were carried out using Cox regression and the log rank test. Differences between the median Day 29 MRD results in the ABD and non-ABD groups was compared using the Wilcoxon Mann-Whitney test. Statistical analysis was conducted using STATA version 14.2 (STATACORP, Texas), MRD scatter plot drawn using GraphPad Prism 6 (GraphPad Software, Inc, California).

References

- 1. Qureshi A, Mitchell C, Richards S, Vora A, Goulden N. Asparaginase-related venous thrombosis in UKALL 2003- re-exposure to asparaginase is feasible and safe. Brit J Haematol. 2010; 149(3): 410-413.
- 2. van der Velden VHJ, Cazzaniga G, Schrauder A, et al. Analysis of minimal residual disease by Ig/TCR gene rearrangements: guidelines for interpretation of real-time quantitative PCR data. Leukemia. 2007; 21(4): 604-611.
- 3. Jenkinson S, Kirkwood AA, Goulden N, Vora A, Linch DC, Gale RE. Impact of PTEN abnormalities on outcome in pediatric patients with T-cell acute lymphoblastic leukemia treated on the MRC UKALL2003 trial. Leukemia. 2016; 30(1): 39-47.
- 4. Gutierrez A, Dahlberg SE, Neuberg DS, et al. Absence of biallelic TCRgamma deletion predicts early treatment failure in pediatric T-cell acute lymphoblastic leukemia. J Clin Oncol. 2010; 28(24): 3816-3823.
- 5. Vora A, Goulden N, Wade R, et al. Treatment reduction for children and young adults with low-risk acute lymphoblastic leukaemia defined by minimal residual disease (UKALL 2003): A randomized controlled trial. Lancet Oncol. 2013; 14(3): 199-209.
- 6. O'Connor D, Enshaei A, Bartram J, et al. Genotype-Specific Minimal Residual Disease Interpretation Improves Stratification in Pediatric Acute Lymphoblastic Leukemia. J Clin Oncol. 2018; 36(1):34–43.

Supplementary Tables

Table S1: Comparison of the baseline characteristics, treatment allocation and response of T-ALL patients that were (n=152) or were not (n=241) included in the study.

		Patients without					
	Patients with samples		P				
	N=152	samples N=241	r				
Baseline characteristics	N-132	111-241					
Sex, N (%)	447 / 77 0)	100/007)	0.42				
Male	117 (77.0)	168 (69.7)	0.12				
Female	35 (23.0)	73 (30.3)					
WBC count (x10 ⁹ /L), N (%)		()					
<50	48 (31.6)	95 (39.4)	0.12				
≥50	104 (68.4)	146 (60.6)					
Age, N (%)							
<10 years	82 (53.9)	107 (44.4)	0.07				
≥10 years	70 (46.1)	134 (55.6)					
NCI risk group, N (%)							
Standard	19 (12.5)	40 (16.6)	0.27				
High	133 (87.5)	201 (83.4)					
CNS disease at diagnosis, N (%)							
No	144 (94.7)	225 (93.4)	0.58				
Yes	8 (5.3)	16 (6.6)					
Final Treatment Allocation and							
Response							
Final treatment given, N (%)							
Α	14 (9.2)	30 (12.4)	0.25				
В	87 (57.2)	118 (49.0)					
С	51 (33.6)	93 (38.6)					
Slow early Response, N (%)	· · · · · · · · · · · · · · · · · · ·						
No	97 (63.8)	148 (61.4)	0.55#				
Yes	37 (24.3)	55 (22.8)					
Unknown*	18 (11.8)	38 (15.8)					
MRD, N (%)							
Negative (<0.01% positive cells)	50 (32.9)	70 (29.0)	0.25 [§]				
Positive (≥0.01% positive cells)	67 (44.1)	97 (40.2)					
Indeterminate $^{\Psi}$, ,	, ,					
	35 (23.0)	74 (30.7)					

P values derived using Chi-squared tests. *Patients without bone marrow results at day 8 or 15 (assumed to have rapid early response for treatment escalation decision) and two patients with conflicting slow early response data and bone marrow results. #Excluding the unknown group: P = 0.92. §Excluding the indeterminate group: P = 0.89. $^{\Psi}$ Includes samples where there were either no targets for MRD assessment or the targets were not sensitive enough, and samples where MRD was either not analysed or was unevaluable due to the sample being inadequate or having missing data. WBC, white blood cell count; NCI, National Cancer Institute, CNS, central nervous system; MRD, minimal residual disease.

Table S2: Time-to-event outcomes comparing the groups with and without samples for ABD analysis.

			4	_	
		Events/n	HR (95% CI)	P	5-year rate (95% CI)
EFS					
	Without samples	59/241	1.00	0.075	76.9% (71.0 – 81.8)
	With samples	27/152	0.66(0.42 - 1.05)		84.2% (77.4 – 89.1)
RFS					
	Without samples	37/239	1.00	0.41	84.9% (79.4 – 89.0)
	With samples	20/151	0.80(0.46 - 1.38)		87.2% (80.7 – 91.6)
OS					
	Without samples	43/241	1.00	0.074	82.8% (77.4 – 87.0)
	With samples	18/152	0.61(0.35 - 1.05)		89.5% (83.4 – 93.4)

P values calculated using the log rank test. EFS, Event-Free Survival; RFS, Relapse-Free Survival; OS, Overall Survival; HR, Hazard Ratio; CI, Confidence Interval.

Table S3. Comparison of baseline characteristics, treatment allocation, response and molecular characterisation of patients with definitive and indeterminate qPCR results for the ABD assay.

	qPCR result	Indeterminate qPCR result	Р
	N=133	N=19	P
Baseline characteristics			
Sex, N (%)			
Male	104 (78.2)	13 (68.4)	0.38*
Female	29 (21.8)	6 (31.6)	
WBC count (x10 ⁹ per L), N (%)			
<50	39 (29.3)	9 (47.4)	0.11
≥50	94 (70.7)	10 (52.6)	
Age, N (%)			
<10 years	72 (54.1)	10 (52.6)	0.90
≥10 years	61 (45.9)	9 (47.4)	
NCI risk group, N (%)			
Standard	14 (10.5)	5 (26.3)	0.066*
High	119 (89.5)	14 (73.7)	
CNS disease at diagnosis, N (%)			
No	125 (94.0)	19 (100.0)	0.60*
Yes	8 (6.0)	0	
Final Treatment Allocation and			
Response			
Final treatment given, N (%)			
A	11 (8.3)	3 (15.8)	0.53*
В	76 (57.1)	11 (57.9)	
С	46 (34.6)	5 (26.3)	
Slow early Response, N (%)			
No	85 (63.9)	12 (63.2)	>0.99 ^{*,§}
Yes	32 (24.1)	5 (26.3)	
Unknown [#]	16 (12.0)	2 (10.5)	
MRD, N (%)			
Negative (<0.01% positive cells)	41 (30.8)	9 (47.4)	$0.34^{*,\Psi}$
Positive (≥0.01% positive cells)	61 (45.9)	6 (31.6)	
Indeterminate $^{\theta}$	31 (23.3)	4 (21.1)	
Molecular Characterisation			
NOTCH1/FBXW7, N (%)			
Wild type	47 (35.3)	6 (31.6)	0.20*
Single Mutant	50 (37.6)	11 (57.9)	
Double Mutant	36 (27.1)	2 (10.5)	
PTEN, N (%)		. ,	
Wild type	94 (75.8)	17 (94.4)	0.12*
Mutant	30 (24.2)	1 (5.6)	
RAS, N (%)	` '	` ,	
Wild type	120 (90.2)	17 (89.5)	>0.99
Mutant	13 (9.8)	2 (10.5)	

P values derived using Chi-squared tests unless otherwise indicated. *Fisher's exact test. #Patients without bone marrow results at day 8 or 15 (assumed to be RER for treatment escalation) and one patient with conflicting slow early response data and bone marrow results. \$Excluding the unknown group: P > 0.99. *Excluding the indeterminate group: P = 0.15. *Includes samples where there were either no targets for MRD assessment or the targets were not sensitive enough, and samples where MRD was either not analysed or was unevaluable due to the sample being inadequate or having missing data. WBC, white blood cell count; NCI, National Cancer Institute, CNS, central nervous system; MRD, minimal residual disease.

Table S4: Time-to-event outcomes comparing the groups with definitive and indeterminate qPCR results for the ABD assay.

		Events/n	HR (95% CI)	Р	5-year rate (95% CI)
EFS					
	qPCR result Indeterminate qPCR	24/133 3/19	1.00 0.86 (0.26 – 2.87)	0.81	84.2% (76.8 – 89.4) 84.2% (58.7 – 94.6)
RFS					
	qPCR result Indeterminate qPCR	17/132 3/19	1.00 1.22 (0.36 – 4.17)	0.75	87.6% (80.6 – 92.2) 84.2% (58.7 – 94.6)
os					
	qPCR result Indeterminate qPCR	16/133 2/19	1.00 0.85 (0.20 – 3.71)	0.83	89.5% (82.9 – 93.6) 89.5% (64.1 – 97.3)

P values calculated using the log rank test. EFS, Event-Free Survival; RFS, Relapse-Free Survival; OS, Overall Survival; HR, Hazard Ratio; CI, Confidence Interval.

Table S5: Time-to-event outcomes comparing the ABD and non-ABD groups.

		Events/n	HR (95% CI)	Р	5-year rate (95% CI)
EFS					
	Non-ABD	17/110	1.00	0.09	85.4% (79.5 – 92.3)
	ABD	7/23	2.12 (0.88 – 5.12)		77.3% (55.4 – 90.3)
RFS					
	Non-ABD	13/110	1.00	0.42	88.8% (81.1 – 93.5)
	ABD	4/22	1.58 (0.52 – 4.86)		81.8% (58.5 – 92.8)
OS					
	Non-ABD	12/110	1.00	0.37	90.0% (82.7 – 94.3)
	ABD	4/23	1.67 (0.54 – 5.17)		87.0% (64.8 – 95.6)

P values calculated using the log rank test. EFS, Event-Free Survival; RFS, Relapse-Free Survival; OS, Overall Survival; HR, Hazard Ratio; CI, Confidence Interval.

Table S6: Relapse-free survival comparing the ABD and non-ABD groups treated on Regimens $\mbox{\sc A/B}$ and C.

	Events/n	HR (95% CI)	Р	5-year rate (95% CI)
Regimen A/B				
Non-ABD	7/75	1.00	0.31	91.9% (82.8 – 96.3)
ABD	0/11	-		100%
Regimen C				
Non-ABD	6/35	1.00	0.21	82.1% (64.4 – 91.6)
ABD	4/11	2.19 (0.62 - 7.78)		63.6% (29.7 – 84.5)

P values calculated using the log rank test. HR, Hazard Ratio; CI, Confidence Interval.

Supplementary Figure Legends

Figure S1. Kaplan-Meier survival curves of the patients included or not included in the study. (A) Overall Survival, (B) Event-Free Survival, (C) Relapse-Free Survival.

Figure S2. Validation of the TRG:ANLN qPCR assay. (A) Comparison of results obtained using WGA and Non-WGA samples from 26 patients with available material. (B) Bland-Altman plot showing the differences between the TRG:ANLN WGA and non-WGA results plotted against the mean of each pair of fold changes. The shaded area shows the agreement interval in which 95% of the differences are expected to lie. (C) TRG:ANLN and TRG:COA1 fold changes for diagnostic WGA samples from 15 patients with indeterminate ABD status by TRG:ANLN fold change.

Figure S3. Kaplan-Meier survival curves of the patients with definitive and indeterminate qPCR results. (A) Overall Survival, (B) Event-Free Survival, (C) Relapse-Free Survival.

Figure S4. Kaplan-Meier survival curves grouped according to ABD status for patients on different treatment regimens. Relapse-Free Survival for patients on (A) Regimen C, (B) Regimen A or B, and (C) those with positive MRD regardless of treatment regimen.

Figure S5. Schematic diagram of the UKALL2003 trial protocol. (A) Treatment intensity decision points, (B) Randomization arms of the trial based on MRD

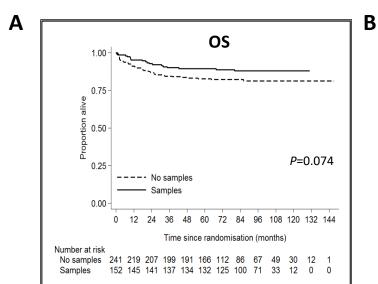
analysis. Slow Early Response defined as ≥25% blasts in the Day 15 bone marrow for Regimen A and Day 8 marrow for Regimen B; High Risk Cytogenetics includes *KMT2A* [*MLL*] and *TCF3-HLF* fusions, near haploidy, low hypodiploidy, and iAMP21; MRD positive defined as ≥0.01% positive cells and MRD negative as <0.01% positive cells; MRD indeterminate includes samples with no targets for MRD assessment or targets were present but not sensitive enough, and samples where MRD was either not analysed or was not evaluable due to the sample being inadequate or having missing data. Abbreviations: ALL, Acute Lymphoblastic Leukemia; MRD, Minimal Residual Disease; WCC, White Cell Count.

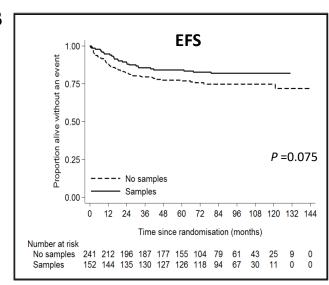
Figure S6. Schematic diagram of the UKALL2003 treatment regimens A, B and C. Regimen A: 3 drug induction with vincristine, dexamethasone, asparaginase followed by consolidation with daily mercaptopurine and central nervous system (CNS)-directed therapy with weekly intrathecal methotrexate. Interim Maintenance (IM): daily mercaptopurine, weekly methotrexate, monthly vincristine and corticosteroid pulses; Delayed Intensification (DI): asparaginase, vincristine, dexamethasone, doxorubicin, cyclophosphamide and cytarabine; Continuing Therapy: oral mercaptopurine and methotrexate, monthly vincristine, corticosteroid pulses, and intrathecal methotrexate every 3 months. Regimen B: 4 drug induction: daunorubicin in addition to dugs used in regimen A induction. Consolidation phase as in Regimen A with the addition of BFM, Berlin Frankfurt Munster (4 weeks of cyclophosphamide and cytarabine). Regimen C: Augmented consolidation by addition of 4 doses of vincristine and 2 doses of pegylated asparaginase during BFM consolidation. Capizzi maintenance as interim maintenance consisted of escalating doses of intravenous methotrexate without

folinic acid rescue, and vincristine and pegylated asparaginase. Abbreviations: MRD, minimal residual disease; CNS, central nervous system; IM, interim maintenance; DI, delayed intensification; BFM, Berlin Frankfurt Munster.

Further treatment regimen details available at Vora et al⁵ and O'Connor et al.⁶

Figure S1





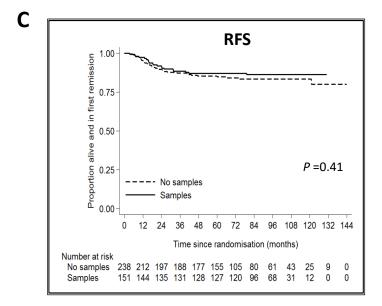


Figure S2

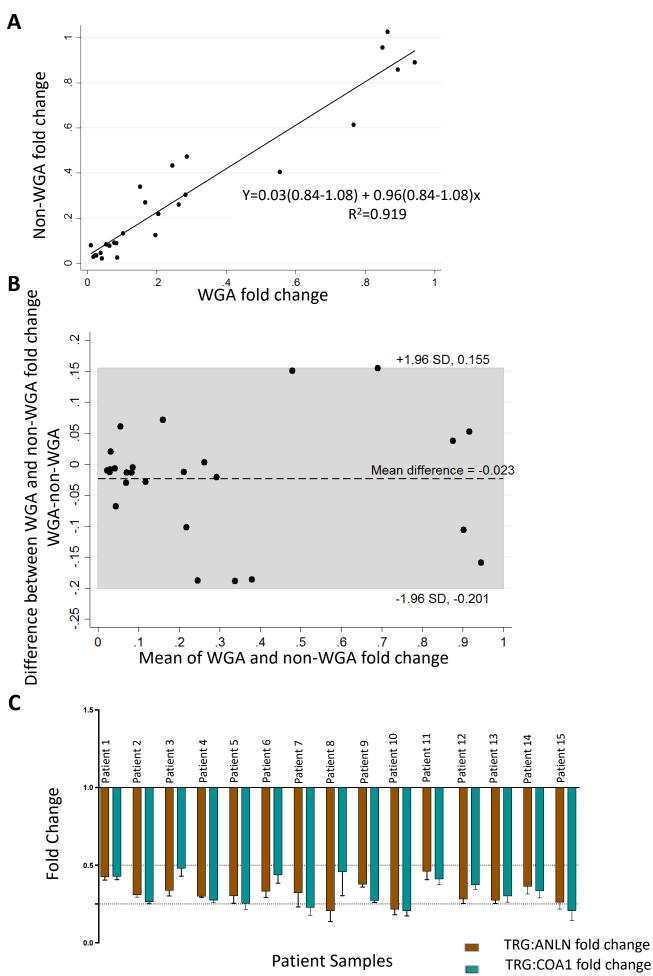
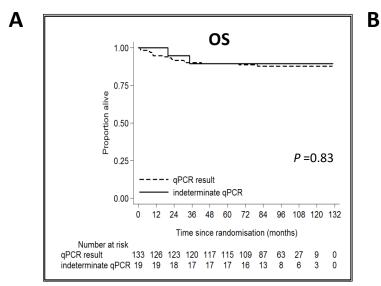
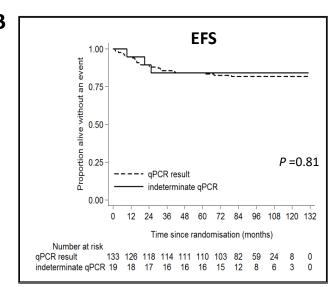


Figure S3





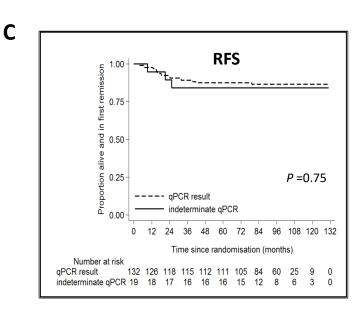
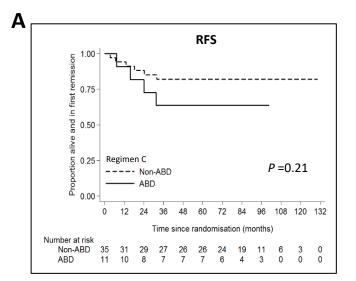
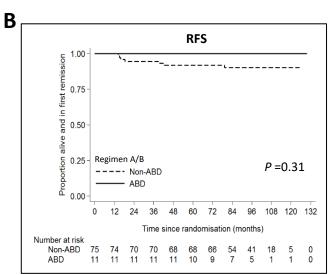


Figure S4





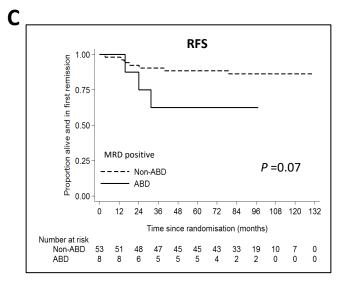
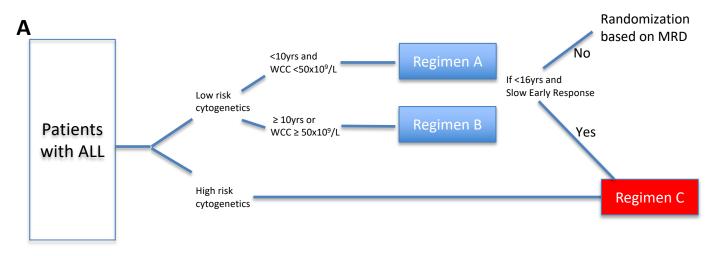
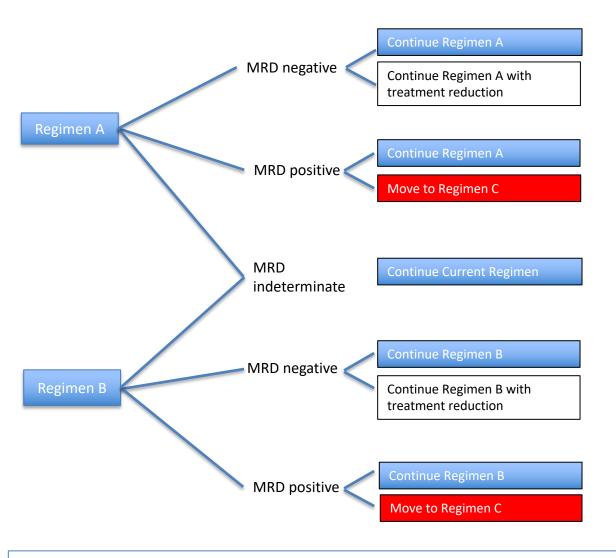


Figure S5

В



Treatment Intensity Decision points



Randomization based on MRD

Figure S6

