SUPPLEMENTARY APPENDIX

Integration of B-cell receptor-induced ERK1/2 phosphorylation and mutations of SF3B1 gene refines prognosis in treatment-naïve chronic lymphocytic leukemia

Chiara Cavallini,¹ Carlo Visco,² Santosh Putta,³ Davide Rossi,⁴ Elda Mimiola,⁵ Norman Purvis,⁶ Ornella Lovato,¹ Omar Perbellini,² Erika Falisi,² Monica Facco,⁷ Livio Trentin,⁷ Maria G. Romanelli,⁸ Gianpietro Semenzato,⁷ Achille Ambrosetti,⁵ Gianluca Gaidano,⁹ Giovanni Pizzolo,⁵ Alessandra Cesano¹⁰ and Maria T. Scupoli^{1,5}

¹Research Center LURM, University of Verona, Italy; ²Department of Cell Therapy and Hematology, San Bonolo Hospital, Vicenza, Italy; ³Qognit Inc., Foster City, CA, USA; ⁴Hematology, Oncology Institute of Southern Switzerland and Institute of Oncology Research, Bellinzona, Switzerland; ⁵Department of Medicine, Section of Hematology, University of Verona, Italy; ⁴Pierian Biosciences, Franklin, TN, USA; ⁵Department of Medicine, Hematology and Clinical Immunology Branch, University of Padua, Italy; ⁵Department of Neurosciences, Biomedicine and Movement Sciences, Section of Biology and Genetics, University of Verona, Italy; ⁵Department of Translational Medicine, Division of Hematology, Amedeo Avogadro University of Eastern Piedmont, Novara, Italy and ¹⁰NanoString, Inc, Seattle, WA, USA

Correspondence: mariateresa.scupoli@univr.it doi:10.3324/haematol.2016.154450

SUPPLEMENTARY DATA

METHODS

Single Cell Network Profiling (SCNP) assay and metric. PBMCs were analyzed by SCNP on a FACSCanto II cytometer (Becton Dickinson, Franklin Lakes, NJ, US), as previously described. Fixed and permeabilized cells were stained with cocktails of fluorochromeconjugated antibodies: CD3-APCCy7 (clone UCHT1; BioLegend, San Diego, CA, US), CD5-V450 (clone L17F12; BD Biosciences, San Jose, CA, US), CD14-APCCy7 (clone M5E2; BioLegend), CD19-PE (clone SJ25C1; eBioscience, San Diego, CA, US), cPARP-AlexaFluor647 (clone F21-852; BD Biosciences), p-ERK-AlexaFluor488 (Thr202/Tyr204; clone D13.14.4E; Cell Signalling Technology, Danvers, MA, US). Dead cells and debris were excluded on the basis of forward scatter, side scatter, and cPARP signals. All analyses were gated on CLL cells, which were identified as CD3/CD14 negative cells co-expressing CD5 and CD19. The raw instrument fluorescence intensities were converted into calibrated intensity metrics (equivalent number of reference fluorophores, ERF). The anti-IgM-induced ERK1/2 phosphorylation (anti-IgM→pERK1/2) was measured using the Uu metric, the Mann-Whitney U statistic comparing the ERF values of the modulated and unmodulated wells. The prespecified anti-IgM-p-ERK1/2 Uu cut-off of 0.66 was used to separate patients into groups, based on our previous validation study.1

Genetic analysis. Cytogenetic abnormalities were evaluated by fluorescence *in situ* hybridization (FISH), according to the hierarchical risk model of FISH anomalies.² The mutation hot spots of the *NOTCH1*, *SF3B1*, *TP53*, *BIRC3*, *and MYD88* genes were analyzed by PCR amplification and direct sequencing of high-molecular-weight genomic DNA, as previously described.³

NOTCH1 mutations were detected in 17/146 (12%) cell samples. Thirteen patients of 17 (76%) carried the highly recurrent c.7544_7545delCT deletion, 1/17 (6%) the c.7462C>T mutation and 1/17 (6%) c.7593_7594insC mutation; 2/17 (12%) lesions were non-sense events. *SF3B1* genetic lesions were carried by cells from 19/146 (13%) patients. Missense mutations in the *SF3B1* hotspots affected codons 625 (p.R625L, n=1), 626 (p.N626I and p.N626Y, n=2), 700 (p.K700E, n=10), 704 (p.I704F, n=1), 740 (p.G740E, n=2), 742 (p.G742D, n=1), 760 (p.E760Q, n=1), 765 (p.Y765C, n=1). *TP53* gene mutations were detected in 11/146 (8%) cell samples, with 1 patient harboring 2 mutations.

Statistical analysis. Clinical and biological features between patients were compared with Fisher's exact test, χ^2 test or Student's t-test, as appropriate. Two-sample Wilcoxon signed-rank sum test was used to compare ERK1/2 phosphorylation in unmodulated and anti-lgM-modulated samples. Mann-Whitney test was used to compare phosphorylation of ERK1/2 in patients grouped on the basis of standard prognostic factors (*IGHV* mutational status and CD38 expression). Kruskal-Wallis test was used to compare phosphorylation of ERK1/2 in patients grouped on the basis of cytogenetic alterations [del(13q), trisomy 12, del(11q), del(17p)].

Time-to-first-treatment (TTFT) was calculated from the date of diagnosis to the date of initial therapy. Patients who did not receive any treatment during follow-up were censored at their last follow-up date. TTFT curves estimated using the Kaplan-Meier method for the respective groups of patients were compared using the log-rank χ^2 test. Univariate and bivariate models for TTFT were generated using Cox proportional hazards regression. Differences between

data were considered statistically significant for *P* values ≤0.05. Graphing and statistical analyses were performed using GraphPad Prism software (GraphPad Software Inc., La Jolla, CA, US) or StatView (Abacus Concepts, Berkeley, CA, US).

References.

- 1. Cesano A, Perbellini O, Evensen E, et al. Association between B-cell receptor responsiveness and disease progression in B-cell chronic lymphocytic leukemia: results from single cell network profiling studies. Haematologica. 2013;98(4):626-634
- 2. Döhner H, Stilgenbauer S, Benner A, et al. Genomic aberrations and survival in chronic lymphocytic leukemia. N Engl J Med. 2000;343(26):1910-1916
- 3. Fabbri G, Rasi S, Rossi D, et al. Analysis of the chronic lymphocytic leukemia coding genome: role of NOTCH1 mutational activation. J Exp Med. 2011;208(7):1389-1401

Supplementary Table S1. Characteristics of CLL patients¹

	Entire patient set (n=152)	Binet stage A patients (n=112)	P value Entire set <i>vs</i> Binet A
Age at the diagnosis median (range), years	66 (28 to 90)	66 (36 to 89)	NS
Gender male female	102 (67%) 50 (33%)	76 (68%) 36 (32%)	NS
Stage Binet A Binet B Binet C unavailable	112 (74%) 30 (20%) 8 (5%) 2 (1%)	112 (100%) - - -	NA
IGHV ² mutated unmutated unavailable	75 (49%) 59 (39%) 18 (12%)	58 (52%) 39 (35%) 15 (13%)	NS
CD38 ³ negative positive	112 (74%) 40 (26%)	87 (78%) 25 (22%)	NS
Cytogenetic del(13q) trisomy 12 del(11q) del(17p) normal unavailable	66 (43%) 18 (12%) 14 (9%) 8 (5%) 42 (28%) 11 (7%)	51 (45%) 13 (12%) 8 (7%) 7 (6%) 30 (27%) 8 (7%)	NS
Follow up median (range), months	50 (0 to 120)	50 (0 to 120)	NS
Disease progression, requiring treatment during follow up unavailable	63 (41%) 4 (3%)	33 (29%) 2 (2%)	NA
TTFT median (range), months	16 (0 to 120)	25 (0 to 120)	NS

¹Patients were enrolled from January 2006 to December 2013 in a regional prospective registry (CLL Veneto Patients were enrolled from January 2006 to December 2013 in a regional prospective registry (CLL veneto project) at the Hematology Units of Verona (Azienda Ospedaliera Universitaria Integrata), Vicenza (San Bortolo Hospital), and Padua (Azienda Ospedaliera).

²IGHV sequencing utilized a 2% cut-off to discriminate unmutated (UM) from mutated (M) IGHV.

³CD38 was determined using a 30% cut-off.

NS: not significant
NA: not applicable

Supplementary Table S2. Comparison of early versus advanced clinical stage, and treated versus untreated patients

	Early <i>vs</i>	advanced clinical sta	age	Untreated vs treated patients			
	Binet A n, (%)	Binet B-C n, (%)	P value	Untreated n, (%)	Treated n, (%)	P value	
Anti-IgM→p-ERK >0.66 Uu	66/112, (58%)	25/38, (66%)	NS	41/85, (48%)	49/63, (78%)	0.0003	
IGHV unmutated	39/97, (40%)	19/36, (53%)	NS	19/73, (26%)	39/60, (65%)	< 0.0001	
CD38 positive	25/112, (22%)	14/38, (37%)	NS	13/85, (15%)	26/63, (41%)	0.0006	
NOTCH1 mutated	10/107, (9%)	4/37, (11%)	NS	7/81, (9%)	7/61, (11%)	NS	
SF3B1 mutated	8/107, (7%)	11/37, (30%)	0.0014	4/81, (5%)	15/61, (24%)	0.0009	
TP53 mutated	10/107, (9%)	1/37, (3%)	NS	3/81, (4%)	7/61, (11%)	NS	
Del(13q)	51/104, (49%)	15/37, (40%)	NS	44/79, (56%)	22/60, (37%)	0.0392	
Trisomy 12	13/104, (12%)	5/37, (13%)	NS	8/79, (10%)	10/60, (17%)	NS	
Del(11q)	8/104, (8%)	6/37, (16%)	NS	4/79, (5%)	11/60, (18%)	0.0244	
Del(17p)	7/104, (7%)	1/37, (3%)	NS	2/79, (2%)	5/60, (8%)	NS	

NS: not significant

Supplementary Table S3. Gene mutation distribution in selected CLL subset

		NOTCH1			SF3B1			TP53		TP	TP53 + del(17p)	
CLL subset	wt n, (%)	mut. n, (%)	P value	wt n, (%)	mut. n, (%)	P value	wt n, (%)	mut. n, (%)	P value	wt- <i>TP53</i> and not <i>del(17p)</i> n, (%)	m- <i>TP53</i> and/or del(17p) n, (%)	P value
Anti-IgM→p-ERK1/2 >0.66 Uu	73/131, (56%)	11/15, (73%)	SN	73/127, (57%)	17/19, (89%)	0.0098	80/135, (59%)	10/11, (91%)	0.0514	72/121, (59%)	11/14, (79%)	SN
IGHV unmutated	51/119, (43%)	8/11, (73%)	S Z	43/111, (39%)	16/19, (84%)	0.0003	52/121, (43%)	7/9, (78%)	SZ SZ	ı	ı	Ϋ́Z
CD38 positive	32/131, (24%)	8/15, (53%)	0.0291	32/127, (25%)	8/19, (42%)	NS	34/135, (35%)	6/11, (54%)	NS	1	-	NA
Del(13q)	52/122, (43%)	5/13, (38%)	SN	54/116, (46.5%)	7/19, (37%)	SN	59/126, (47%)	2/9, (22%)	SN	ı	ı	NA
Trisomy 12	13/122, (11%)	4/13, (31%)	9090'0	16/116, (14%)	1/19, (5%)	NS	15/126, (12%)	2/9, (22%)	NS	ı	_	NA
Del(11q)	13/122, (11%)	1/13, (8%)	SN	11/116, (9%)	0/19,	NS	12/126, (10%)	2/9, (22%)	NS	1	-	NA
Del(17p)	7/122, (6%)	1/13, (8%)	SN	8/116, (7%)	0/19,	NS	5/126, (4%)	3/9, (33%)	0.0099	1		NA

NS: not significant; NA: not applicable

Supplementary Table S4A. Selected Cox proportional hazards models for TTFT in the entire patient set (n=125)

Variable 1	HR (CI 95%) ¹	<i>P</i> value var1	Variable 2	HR (CI 95%) ¹	P value var2
Anti-IgM→p- ERK1/2	1.296 (0.643-2.614)	0.46	NA	NA	NA
IGHV unmutated	2.785 (1.428-5.434)	0.002	NA	NA	NA
CD38 positive	1.245 (0.642-2.552)	0.56	NA	NA	NA
SF3B1 mutated	2.304 (1.204-4.405)	0.01	NA	NA	NA
Anti-IgM→p- ERK1/2	1.653 (0.854-3.198)	0.13	IGHV unmutated	3.300 (1.855-5.847)	< 0.0001
Anti-IgM→p- ERK1/2	1.922 (1.033-3.566)	0.03	SF3B1 mutated	3.731 (2.016-6.896)	0.0001

¹HR: Hazard Ratio; CI: Confidence Interval

Supplementary Table S4B. Selected Cox proportional hazards models for TTFT in Binet stage A patients (n=90)

Variable 1	HR (CI 95%) ¹	P value var1	Variable 2	HR (CI 95%) ¹	P value var2
Anti-IgM→p- ERK1/2	1.071 (0.334-2.587)	0.89	NA	NA	NA
IGHV unmutated	5.617 (1.538-15.384)	0.0008	NA	NA	NA
CD38 positive	1.683 (0.547-2.452)	0.19	NA	NA	NA
TP53 mutated	2.145 (0.833-5.556	0.11	NA	NA	NA
SF3B1 mutated	3.355 (1.230-9.174)	0.01	NA	NA	NA
Anti-IgM→p- ERK1/2	1.523 (0.605-3.837)	0.37	IGHV unmutated	6.578 (2.724-15.873)	< 0.0001
Anti-IgM→p- ERK1/2	2.437 (0.976-6.089)	0.05	SF3B1 mutated	5.319 (2.053-13.698)	0.0006

¹HR: Hazard Ratio; CI: Confidence Interval