

A randomized phase II trial of azacitidine +/- epoetin- β in lower-risk myelodysplastic syndromes resistant to erythropoietic stimulating agents

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Supplemental Data

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Supplementary informations:

Exclusion criteria:

Patients with any of the following: creatinine clearance < 30 ml/min, serum bilirubin > 40 µmol/L, cardiac failure defined by NYHA dyspnea > 2, HIV infection, planned hematopoietic stem cell transplantation, prior treatment with chemotherapy or a hypomethylating agent, were excluded.

Treatment regimen:

Delayed administration of AZA was allowed in case of predefined hematological toxicity until day 42. AZA dose level reduction was planned in case of persistent cytopenia after day 42, defined in the absence of MDS progression, as > grade 2 non-hematological toxicity, or hematopoietic toxicity leading to hospitalization. Marrow samples were obtained before study entry, after 4, 6 and 18 cycles, and in case of persistent cytopenia, as defined above. Quality of life (QOL), using FACTan and EQ5D questionnaires, was systematically assessed before randomization and after 4, 6 and 18 cycles. Results of QOL studies, currently not available, will not be presented here.

Conventional cytogenetic and SNP- A karyotyping:

Raw data were analyzed using the Genotyping Console version 4.1 software (Affymetrix). As no germline DNA was available, the following criteria were chosen to define SNP-A abnormality: (i) Copy number aberration (CNA) of more than 1 megabases (Mb) and with less than 50% overlap with germline copy number variants (CNVs) reported in the Database of Genomic Variants (<http://dgv.tcag.ca/dgv/app/home>) (ii) Size and location criteria (telomeric > 8.7 Mb and interstitial, > 25 Mb) were used for identification of somatic UPD as previously described [12]. Subsequently, all CNA and UPD were validated by visual inspection and annotated based on the hg19 human genome assembly.

Mutational analysis:

Massively parallel sequencing on ABI's SOLiD v4 system (Life Technologies, Carlsbad, CA) or Personal Genome Machine® (PGM™) platform (Life Technologies®) of SureSelect (Agilent, Santa Clara, CA, USA) captured or Ion Ampliseq™ (Life Technologies®) amplified target sequences, respectively. Not previously reported variants were confirmed by direct Sanger sequencing for the following genes: *ASXL1*, *CBL*, *DNMT3A*, *IDH2*, *RUNX1* and *TET2* using primers described elsewhere [21].

Sample size justification and statistical analysis:

For continuous variables, values were expressed as median and interquartile range [IQR] and categorical variables were reported as count or percentage and 95% confidence interval (CI95%).

The primary efficacy endpoint was achievement of RBC-TI after 6 cycles, ie major erythroid response (HI-E) response according to IWG 2000 criteria. Such an erythroid response (according to IWG 2000 criteria and IWG 2006) was considered as a binary variable with comparisons across randomization groups based on the exact Fisher test. Missing responses were considered as failures. The search for predictive factors of response was based on multivariate logistic regression. The duration of response was studied in responding patients, computed from the date of response to the date of the first observed event such as relapse, failure, progression, or death. Overall survival was defined as the time between randomization and death. IPSS progression free survival was calculated from the date of randomization to the date of the progression of IPSS. Distributions of the response time, overall survival and IPSS progression free survival were estimated by the Kaplan – Meier method, where patients free of the events of interest were treated as right censored observations at the time of their last follow –up. Differences in these distributions were tested by the log rank test. A Cox model was used for assessing prognostic factors of overall survival. All statistical tests were two-sided, and considered significant for P values < 0.05. Analysis was performed using the SAS 9.3 software (SAS Inc, Cary, NC) and R version 3.0.2 (The R Foundation for Statistical Computing)

Participating centers:

CHU d'Amiens (Dr Gruson), CHU d'Angers (Pr Hunault-Berger), Hôpital Avicenne APHP (Pr Gardin), CH d'Avignon (Dr Slama), Hôpital du Kremlin-Bicetre APHP (Dr Tertian), CH de Boulogne-sur-mer (Dr Choufi), CHU de Caen (Dr Cheze), Hôpital Cochin APHP (Pr Dreyfus), CHU de Dijon (Dr Bastié), CHU de Limoges (Dr Gourin), CHU de Lyon (Pr Wattel), IPC Marseille (Pr Vey), CHU de Nancy (Pr Guerci), CHU de Nantes (Dr Delaunay), CHU de Nice (Dr Legros), Hôpital Saint Antoine APHP (Dr Isnard), Hôpital Saint Louis APHP (Dr Raffoux), CH de Perpignan (Dr Sanhes), CHU de Poitiers (Dr Roy), Centre Henri Becquerelle de Rouen (Dr Stamatoulas), CHU de Strasbourg (Dr Ame), Hôpital Saint Vincent de Paul Lille (Pr Rose), Toulouse (Pr Beyne Rauzy)

Supplementary Tables

Table S1. List of sequenced genes

function	Genes
epigenetic modifiers	<i>TET2, IDH1, IDH2, ASXL1, EZH2, DNMT3A, DNMT3B, KDM6A, SUZ12</i>
splicing	<i>SF3B1, SRSF2, U2AF1, U2AF2, PRFP40b, ZRSR2, SF1, SF3A1</i>
signaling	<i>KIT, CBL, BRAF, JAK2, KRAS, NRAS, FLT3, PTPN11, CSF1, CDKN2A, PTEN</i>
Transcription	<i>RUNX1, GATA1, GATA2, TP53, ETV6, WT1</i>

Table S2. Details of SNP-array karyotyping abnormalities

Case	Karyotype	nb of lesions	Type of lesion	Chromosome	Cytoband start	Cytoband end	Size (Mb)
1	normal	1	UPD	14	q31.1	q32.33	27918
2	normal	1	loss*	7	q22.1	q22.1	2141
3	normal	1	gain*	2	q12.3	q14.3	16291
4	normal	1	UPD	4	q13.3	q35.2	117021
5	normal	1	loss*	21	q22.12	q22.12	1434
6	normal	2	loss*	4	q24	q24	1050
			UPD	13	q13.2	q34	76250
7	normal	2	gain*	12	all		132000
			loss*	13	q14.13	q22.1	27855
8	normal	3	UPD	10	q25.3	q26.3	16918
			UPD	7	q21.11	q36.3	73479
			UPD	4	q22.1	q35.2	101558
9	failure	1	loss*	4	q24	q24	2282
10	47,XY,+8 [20]	1	gain	8	all		146298
11	47,XY,+8 [20]	1	gain	8	all		146298
12	47,XX,+8 [8] / 46,XX [12]	2	gain	8	all		146298
			UPD	2	p25.3	p11.2	87089
13	47,XX,+8 [14] / 46,XX [8]	3	gain*	19	p13.3	p13.3	1075
			gain	8	all		146298
			UPD	4	q22.1	q35.2	100084
14	47,XX,+8 [14] / 46,XX [6]	1	gain	8	all		146298
15	47,XX,+8 [8] / 46,XX [12]	1	gain	8	all		146298
16	47,XY,+8 [11] / 45,X,-Y [4]	1	gain	8	all		146298
17	46,X,-Y,+8 [20]	2	gain	8	all		146298
			loss	Y	all		57773
18	45,X,-Y [13] / 46,idem,+8 [7]	2	loss	Y	all		57773
			gain	8	all		146298
19	47,XX,del(5q),+8 [20]	2	loss	5	q22.1	q33.3	45420
			gain	8	all		146298
20	45,X,-Y [24] / 46,XY [1]	1	loss	Y	all		57773

21	45,X,-Y [20]	2	loss	Y	all		57773
			UPD	4	q21.1	q35.2	77073
22	46,XY,del(13q) [26] / 45,X,-Y [3] / 46,XY [1]	2	UPD	14	q32.2	q32.33	8798
			loss	13	q12.3	q21.1	28336
23	46,XY,del(5q) [22]	1	loss	5	q14.3	q23.3	36774
24	46,XX,del(5q)(q13q32) [25]	1	loss	5	q14.3	q33.2	70349
25	46,XX,del(5)(q13q33) [6] / 46,idem,del(1)(p34) [8]	2	loss	5	q14.3	q34	78000
			loss	1	p36.11	p35.3	20655
26	46,XX,del(13)(q12q21) [9] / 46,XX,del(5)(q31q33) [5] / 46,XX [15]	3	loss*	1	p36.13	p36.11	10364
			loss	5	q31.1	q33.3	26396
			loss	13	q13.1	q21.1	24530
27	46,XY,del(20q) [23]	1	loss	20	q11.22	q13.2	17438
28	46,XY,del(20q) [18] / 47,idem,+mar [2]	1	loss	20	q11.22	q13.31	22930
29	46,XY,del(20)(q12) [17] / 46,XY [3]	1	loss	20	q11.21	q13.12	15100
30	47,XY,add(9)(q22),+9 [8] / 47,idem,del(20)(q12) [10] / 46,XY [2]	2	gain	9	all p arm		66482
			loss	20	q11.22	q13.13	16567
31	46,XY,del(12)(p11p13) [17] / 46,XY [3]	2	UPD	11	p15.5	p13	32813
			loss	12	p13.32	p12.3	14613
32	46,XY,add(3)(p11),t(8;12)(q12-13;p13),del(9)(q13q33),del(17)(q21) [20]	4	loss*	12	q14.3	q14.3	1302
			loss*	5	q22.1	q22.2	2181
			loss	3	p21.31	p11.1	42823
			loss	9	q21.11	q33.1	48549
33	46,XX,del(11)(q21q22) [16] / 46,XX [4]	1	loss	11	q21	q24.1	27280

* CNA not detected by conventional cytogenetics

Table S3. Description of gene mutations

Gene	Type of mutation	Base and amino acid changes	Number of mutations
ASXL1	frameshift	NM_015338:exon12:c.1900_1922del:p.E635fs	2
ASXL1	frameshift	NM_015338:exon12:c.1934dupG:p.G646fs	2
ASXL1	frameshift	NM_015338:exon12:c.1919_1929del:p.A640fs	1
CBL	missense	NM_005188:exon9:c.G1259A:p.R420Q	1
CBL	frameshift	NM_005188:exon9:c.1380_1382dup:p.D460dup	1
DNMT3A	frameshift	NM_022552:exon8:c.935delC:p.S312fs	1
DNMT3A	nonsense	NM_022552:exon9:c.C1066T:p.Q356X	1
DNMT3A	frameshift	NM_022552:exon14:c.1571delG:p.C524fs	1
DNMT3A	missense	NM_022552:exon14:c.G1628T:p.G543V	1
DNMT3A	nonsense	NM_022552:exon17:c.C1966T:p.Q656X	1
DNMT3A	frameshift	NM_022552:exon17:c.2043delC:p.I681fs	1
DNMT3A	missense	NM_022552:exon20:c.C2374T:p.R792C	1
DNMT3A	missense	NM_022552:exon20:c.C2395A:p.P799T	1
DNMT3A	frameshift	NM_022552:exon22:c.2532delA:p.K844fs	1
DNMT3A	nonsense	NM_022552:exon22:c.C2536T:p.Q846X	1
DNMT3A	missense	NM_022552:exon23:c.G2645A:p.R882H	1
DNMT3A	missense	NM_022552:exon23:c.G2645C:p.R882P	1
ETV6	missense	NM_001987:exon2:c.C145T:p.R49C	1
EZH2	missense	NM_001203247:exon7:c.C677A:p.A226D	1
EZH2	nonsense	NM_001203247:exon19:c.C2149T:p.Q717X	1
GATA1	frameshift	GATA1:NM_002049:exon2:c.31_32insC:p.T11fs	1
IDH2	missense	NM_002168:exon4:c.G419A:p.R140Q	1
JAK2	missense	NM_004972:exon14:c.G1849T:p.V617F	4

KDM6A	frameshift	NM_021140:exon27:c.3919_3920insG:p.R1307fs	1
KIT	missense	NM_001093772:exon8:c.A1256C:p.D419A	1
PRPF40B	missense	NM_001031698:exon24:c.C2491G:p.R831G	1
RUNX1	frameshift	NM_001001890:exon5:c.885_886insTC:p.S295fs	1
SF3B1	missense	NM_012433:exon14:c.G1866C:p.E622D	8
SF3B1	missense	NM_012433:exon14:c.A1865T:p.E622V	1
SF3B1	missense	NM_012433:exon14:c.C1873T:p.R625C	4
SF3B1	missense	NM_012433:exon14:c.C1986A:p.H662Q	4
SF3B1	missense	NM_012433:exon14:c.A1997G:p.K666R	3
SF3B1	missense	NM_012433:exon14:c.G1998C:p.K666N	1
SF3B1	missense	NM_012433:exon15:c.A2098G:p.K700E	38
SF3B1	missense	NM_012433:exon16:c.G2225A:p.G742D	1
SF3B1	missense	NM_012433:exon16:c.G2233C:p.A745P	1
TET2	frameshift	NM_001127208:exon3:c.2290dupG:p.Q764fs	1
TET2	frameshift	NM_001127208:exon10:c.4317_4318insG:p.K1439fs	1
TET2	frameshift	NM_001127208:exon11:c.5055delT:p.N1685fs	1
TET2	frameshift	NM_001127208:exon11:c.5318_5319insC:p.F1773fs	1
TET2	frameshift	NM_001127208:exon11:c.5324delG:p.S1775fs	1

Gene	Type of mutation	Base and amino acid changes	Number of mutations
TET2	frameshift	NM_001127208:exon3:c.1143delC:p.F381fs	1
TET2	frameshift	NM_001127208:exon3:c.1927_1928insCA:p.S643fs	1
TET2	frameshift	NM_001127208:exon3:c.3202delC:p.Q1068fs	1
TET2	frameshift	NM_001127208:exon4:c.3412delC:p.Q1138fs	1
TET2	frameshift	NM_001127208:exon11:c.5225_5229del:p.1742_1743del	1
TET2	frameshift	NM_001127208:exon7:c.3945delC:p.D1315fs	1
TET2	frameshift	NM_001127208:exon3:c.3347_3348insA:p.I1116fs	1
TET2	frameshift	NM_001127208:exon7:c.3811_3812insG:p.C1271fs	1
TET2	missense	NM_001127208:exon11:c.C5597G:p.P1866R	1
TET2	missense	NM_001127208:exon11:c.T5615C:p.L1872P	1
TET2	missense	NM_001127208:exon11:c.T5692C:p.S1898P	1
TET2	missense	NM_001127208:exon3:c.A1118G:p.Q373R	1
TET2	missense	NM_001127208:exon3:c.G3404A:p.C1135Y	2
TET2	missense	NM_001127208:exon6:c.C3655T:p.H1219Y	1
TET2	missense	NM_001127208:exon7:c.G3866T:p.C1289F	1
TET2	missense	NM_001127208:exon8:c.A4034G:p.Y1345C	1
TET2	missense	NM_001127208:exon8:c.T3998A:p.M1333K	1
TET2	missense	NM_001127208:exon9:c.C4100T:p.P1367L	1
TET2	missense	NM_001127208:exon9:c.C4138T:p.H1380Y	1
TET2	missense	NM_001127208:exon9:c.G4054A:p.E1352K	1
TET2	missense	NM_001127208:exon9:c.G4097A:p.R1366H	1
TET2	nonframeshift	NM_001127208:exon6:c.3641_3642insGGA:p.R1214delinsRE	1
TET2	nonsense	NM_001127208:exon3:c.148_149insTCT:p.G50delinsVX	1
TET2	nonsense	NM_001127208:exon3:c.C1630T:p.R544X	1

TET2	nonsense	NM_001127208:exon3:c.C1852T: p.Q618X	1
TET2	nonsense	NM_001127208:exon3:c.C1876T : p.Q626X	1
TET2	nonsense	NM_001127208:exon3:c.C2050T:p.Q684X	1
TET2	nonsense	NM_001127208:exon3:c.C2200T:p.Q734X	1
TET2	nonsense	NM_001127208:exon3:c.C2227T:p.Q743X	1
TET2	nonsense	NM_001127208:exon3:c.C2428CT:p.Q810X	1
TET2	nonsense	NM_001127208:exon3:c.G1692A:p.W564X	1
TET2	nonsense	NM_001127208:exon3:c.G2995T:p.E999X	1
TET2	splice site	NM_001127208:exon6:c.3595-2A>T	1
TP53	missense	NM_000546:exon5:c.A478T:p.M160L	1
U2AF 1	missense	NM_001025203:exon2:c.C101T:p.S34F	2
U2AF 1	missense	NM_001025203:exon6:c.A470C:p.Q157P	1
ZRSR 2	frameshift	NM_005089:exon8:c.708delC:p.F236fs	1
ZRSR 2	frameshift	NM_005089:exon11:c.1116_1117insT:p.D372fs	1
ZRSR 2	missense	NM_005089:exon8:c.C571T:p.H191Y	1

Table S4: azacitidine reports that included lower risk MDS

Report	N (%) of lower risk pts)	Transfusion dependent patients before AZA	ESA failure before AZA	AZA number of days	Transfusion independency after AZA	Hematologic improvement after AZA	Median response duration
Musto, 2010	74(100%)	83,8%	58%	5d in 39%	NR	45.9%	6 months
Lyons, 2009	151(63%)	48% 43% 50%	NR	5-2-2 d 5-2-5 d 5d	44% 44% 46%	47% 44% 50%	473 d 387 d not reached
Silverman,2002	191(23%)	63%	NR	7d	45%	47% 35%**	9 months
Tobiasson,2014	30 (100%)	100%	100%	5 d	17%	30%	5 months
Falantes, 2015	27	96%	NR	5 d in 37%	NR	40,7%	
Fili, 2013	32	81%	69%	5 d	NR	41%	10 months

NR: not reported

** 5 days schedule after an initial observation period