Mutation status of essential thrombocythemia and primary myelofibrosis defines clinical outcome

Julia Asp,^{1,2} Björn Andréasson,^{3,4} Ulrika Hansson,⁵ Carina Wasslavik,² Johanna Abelsson,³ Peter Johansson,^{3,4} and Lars Palmqvist^{1,2}

¹Department of Clinical Chemistry and Transfusion Medicine, Institute of Biomedicine, the Sahlgrenska Academy, University of Gothenburg; ²Department of Clinical Chemistry, Sahlgrenska University Hospital, Gothenburg; ³Haematology and Coagulation Section, Department of Medicine, Sahlgrenska University Hospital, Gothenburg; ⁴Hematology Section, Department of Medicine, NU Hospital Group, Uddevalla; and ⁵Department of Clinical Pathology and Genetics, Sahlgrenska University Hospital, Gothenburg, Sweden

Correspondence: julia.asp@gu.se doi:10.3324/haematol.2015.138958

Mutation Status of Essential Thrombocythemia and Primary Myelofibrosis Defines Clinical Outcome

Julia Asp, Björn Andréasson, Ulrika Hansson, Carina Wasslavik, Johanna Abelsson, Peter Johansson, and Lars Palmqvist

Methods

Patients

The study was performed in accordance to the Declaration of Helsinki after ethical approval by the Research Ethics Board at the University of Gothenburg and the Sahlgrenska Academy, Gothenburg, Sweden. All patients diagnosed with ET or PMF according to the WHO 2008 classification between 2008 and 2013 in Western Sweden at the Sahlgrenska University Hospital or NU Hospital Group and reported to the Swedish national cancer register, INCA, were selected (n=186). 170 patients were included; 129 ET patients (73 females and 56 males, median age 67 years, range 27–90 years) and 41 PMF patients (21 females and 20 males, median age 70 years, range 32–85 years). 16 patients did not answer the request to participate in the study. Clinical, laboratory and outcome data was available for all patients. A critical review of diagnosis was performed in triple negative patients by re-analyzing bone marrow in addition to review of clinical data.

Mutation analysis of JAK2, CALR and MPL

Genomic DNA from blood taken at initial referral or time of diagnosis was used for analysis. *JAK2* V617F mutation status was determined for all patients. Ipsogen JAK2 MutaQuant Kit (Qiagen, Hilden, Germany) was used in the majority of cases. *CALR* mutation status was analyzed with fragment analysis in all cases negative for *JAK2* mutation.¹ Detected *CALR* mutations were verified by Sanger sequencing. The *MPL* W515 mutation was analyzed using Ipsogen MPL W515K/L MutaScreen Kit (Qiagen) in cases negative for both *JAK2* V617F and *CALR* mutations.

Screening for myeloid mutations

Genomic DNA from all patients negative for *JAK2* V617F, *CALR* and *MPL* W515K/L mutations (n=20) in addition to all ET patients with *MPL* mutations (n=3, all with the W515L mutation) and age matched controls from ET patients harboring the *JAK2* (n=18) or *CALR* (n=18) mutations were screened for additional mutations. The TruSight Myeloid Sequencing panel (Illumina, San Diego, CA, USA) was used on the MiSeq instrument (Illumina). Secondary analysis was performed by MiSeq Reporter using BWA mapper and somatic variant caller. Results were filtered using Variant Studio (Illumina). Global filtering according to 1000 genomes was set to >3%, coverage was at least 100 reads and at least 10 reads for the variant. Variants causing missense, frameshift, altered stop/initation codon, in-frame insertion/deletion or variants affecting splice site were regarded as mutations. Variants with quality >Q30 and allele frequencies of at least 5% were considered positive for mutation. In addition to patient samples, three samples of DNA extracted from blood donors or individuals with normal blood morphology were included as controls. Mutations appearing in these samples were considered as normal polymorphisms occurring in our population cohort or

sequencing artefacts and were excluded from further analysis. BAM-files from the secondary analysis were used to further analyze the regions with identified mutations by Integrative Genomics Viewer (www.broadinstitute.org). Variants in areas with difficult reads were excluded. Results were visualized using a Circos plot.²

Statistical analysis

Statistical significance was determined using Kruskal-Wallis non-parametric test or 2-tailed Fisher exact test with Analyze-it, v.2.30 (Analyse-it Software Ltd, Leeds, UK). A value of p<0.05 was considered statistically significant.

Survival analysis

The probabilities of OS were estimated by the Kaplan-Meier method and differences in survival distributions were compared with Gehan-Breslow-Wilcoxon test using GraphPad Prism, v.6.07 (GraphPad Software Inc, La Jolla, CA, USA). OS was defined as time from diagnosis to last follow-up or death from any cause. Follow up was done from diagnosis to end of study (2015-05-31) and the median follow-up time was 46 months for ET patients and 41 month for PMF patients.

References

 Klampfl T, Gisslinger H, Harutyunyan AS, et al. Somatic mutations of calreticulin in myeloproliferative neoplasms. N Engl J Med. 2013;369(25):2379-2390.
Krzywinski M, Schein J, Birol I, et al. Circos: an information aesthetic for comparative genomics. Genome Res. 2009;19(9):1639-1645.

Table S1 ET with different mutations (median valu	es and ranges)
---------------------------------------------------	----------------

	JAK2 V617F (n=82)	<i>CALR</i> (n=28)	<i>MPL</i> (n=7)	TN (n=8)
Gender (F/M)	49/33	13/15	3/4	6/2
Age	66.5 (27-90)	67.5 (49-88)	80 (70-82)*	64.5 (39-85)
Hemoglobin (g/L)	143 (117-170)**	137 (108-152)	117 (99-128)**	134.5 (113-155)
Hematocrit (EVF)	0.44 (0.37-0.52)**	0.43 (0.37-0.48)	0.38 (0.31-0.40)**	0.40 (0.35-0.47)
WBC (10 ⁹ /L)	9.4 (4.8-16.9)	8.7 (2.9-12.5)	9.0 (5.6-19.8)	9.4 (5.6-15.6)
Platelets (10 ⁹ /L)	736 (467-1776)	991 (407-1824)**	1011 (592-1888)	690 (570-2061)
EPO (2.6-18.5 IU/L)	4.9 (1.7-30.6)***	10.1 (3.3-40.8)	15.0 (6.2-26.3)**	10.9 (6.2-14.6)
Vascular complications (yes/no)	33/49	5/23 ⁺	4/3	3/5
Transformation (yes/no)	1/81	1/27	2/5+	0/8

Kruskal-Wallis non-parametric test * $p \le 0.01$, ** $p \le 0.001$, *** $p \le 0.0001$, Fischer exact test *p < 0.05

	<i>JAK2</i> V617F (n=24)	CALR (n=7)	<i>MPL</i> (n=3)	TN (n=7)
Gender (F/M)	12/12	4/3	1/2	4/3
Age	71 (41-85)	61 (48-82)	82 (73-83)	63 (50-83)
Hemoglobin (g/L)	120 (79-146)	119 (90-146)	111 (104-116)	91 (60-156)
Hematocrit (EVF)	0.38 (0.23-0.49)	0.38 (0.29-0.45)	(0.32-0.35)	(0.21-0.22)
WBC (10 ⁹ /L)	8.4 (3.2-28.0)	8.7 (5.6-42.7)	11.0 (5.3-19.8)	7.1 (1.0-17.8)
Platelets $(10^9/L)$	392 (37-1244)	878 (32-1412)	504 (384-937)	64 (5-833)
EPO (2.6-18.5 IU/L)	9.9 (3.3-578)	19.0 (10.1-30.1)	33.2 (7.4-40.7)	44.8 (4.8-81.1)
Vascular complications (yes/no)	7/17	2/5	0/3	2/5
Transformation (yes/no)	1/23	0/7	0/3	3/4*

Table S2 PMF with different mutations (median values and ranges)

Fischer exact test *p<0.05

Table S3 Findings from TruSight My Mutation load in %	eloid Sequencing panel		ET				PMF	
			TN Patient no.	CALR	JAK2	MPL	TN	MPL
			1 2 3 4 5 6 7 8	9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24	25 26 27 28 29 30 31 32 33 34 35 36 37 38 39 40 41 42 43	44 45 46 47 48 49 50 5	1 52 53 54 55 56 57 58	59
CALR (NM 004343.3) JAK2 (NM 004972.3)	c.1154 1155insTTGTC c.1849G>T	p.Lys385AsntsTer47 p.Val617Phe		24 31 38 37 35 8 41 25 17 34 26	8 10 25 29 14 17 6 33 12 9 14 6 13 35 46 22 32	32		-
MPL (NM 005373.2)	c.1514G>A	p.Ser505Asn				27		
c.15437>C p.Trp515Arg c.15437>C;1546C>G] p.Trp515Arg; Gir c.1543_1544delinsGC p.Trp515Ala	p.Trp515Arg p.Trp515Arg: Gln516Glu]				35	6		
	p.Trp515Ala				72		44	
EZH2 (NM 004456.4)	c.1544G>T c.2110+6T>G	p.Trp515Leu -			50	5 39 28		-
	c.553G>C	p.Asp185His				52		
c.965A	c.965A>G	p.Asn322Ser		49			c	_
ATRA (NW 000485.3)	c.5579A>G	p.Asn1860Ser		100			~	
APRIL DIAL OFFICE PL	c.6479T>C	p.Val2160Ala				6		_
Auto (Am_015556.5)	c.2278C>T	p.Gin760Ter	**	11				
	c.2444T>C	p.Leu815Pro	100			10		
	c.2880G>A	p.Ser9581er p.Trp960Ter		33		42		
	c.3000 3001insA	p.Thr1001AsnfsTer4				27		
KOM64 (NM 021140 2)	c.3083C>A	p.Ser1028Ter		49	9			-
	c.1527+3G>A	-	100					
	c.2240T>A	p.Leu747His p.Gki1064Tar			40			
	c.385-2A>C	partore				8		
	c.4075C>T	p.Gin1359Ter				6		<u> </u>
DNM13A (NM 022552.4)	c.1182delC	p.Leus73vai p.Asp394GlufsTer13				15		2
	c.2193 2195delCTT	p.Phe732del				25		
	c.2644C>T c.2645G>A	p.Arg882Cys p.Arg882His		19	44		38	
	c.517G>T	p.Gly173Cys					6	
	c.718G>T	p.Glu240Ter p.Glu30Ala			ea		68	
TET2 (NM 001127208.2)	c.100C>T	p.Leu34Phe	50	53	53			_
	c.1064G>A	p.Gly355Asp		50	52		51	
	c.1088L>1 c.1273delA	p.Thr425LeufsTer2	47		46			
	c.1737delA	p.Lys580AsnfsTer21				42		
	c.1897A>T	p.Met633Leu p.His800GlofsTer15				51	30	
	c.2604T>G	p.Phe868Leu			51			
	c.3454G>A	p.Gly1152Arg		×			38	
	c.4072T>G	p.Cys1358Gly		20		30		
	c.4097G>A	p.Arg1366His			9			
	c.4567C>T	p.Pro1419 Leu142UIIISLeu p.Gin1523Ter			41 34			
	c.5162T>G	p.Leu1721Trp	53	50				
	c.521C>A c.5284A>G	p.Pro174His p.ile1762Val	49	48	50 50	51		
	c.5605G>T	p.Gly1869Trp					27	
IDH1 (NM_005896.2)	c.5666C>A	p.Pro1889His p.Val71lle				8		-
	c.395G>A	p.Arg132His				27		
IDH2 (NM 002168.2)	c.419G>A	p.Arg140Gin p.Arg140Trp				39	45	
RAD21 (NM 006265.2)	c.407A>G	p.Asn136Ser					5	
STAG2 (NM 001042749.1)	c.1821+2T>A	- 				6		
SMC3 (NM 005445.3)	c.1238T>C	p.lle413Thr			5		3	_
	c.2080G>C	p.Ala694Pro					11	_
ZRSR2 (NM 005089.3)	c.1314 1315insAGCCGG	p.Gly438 Ser439insSerArg					45	_
	c.212T>A	p.Leu71Ter					10	
	c.1385G>A c.873G>A	p.Arg462Gin p.Trp291Ter				59		
SRSF2 (NM 001195427.1)	c.284C>A	p.Pro95His	40			32	31	
	c.284C>T	p.Pro95Leu				42	24	20
U2AF1 (NM 001025203.1)	c.101C>T	p.Ser34Phe		8				
ETU/6 (NML 001987 4)	c.470A>C	p.Gln157Pro		47 49			40	_
BCORL1 (NM_021946.4)	c.1508C>T	p.Pro503Leu		*/			5	_
	c.3158A>G	p.Lys1053Arg p.Gly209Ser		100	40			
BCOR (NM 001123385.1)	c.4977-4G>T	-	50	49	45	53		
	c.651G>A	p.Met217lle			48		19	
RUNX1 (NM 001754.4)	c.167T>C	p.Leu56Ser			\$4		42	
CUX1 (NM 001202543.1)	c.1702G>C	p.Ala568Pro					19	
	c.1/4+8G>A c.2014C>T	- p.Arg672Ter				99 41		
	c.295G>A	p.Val99tle			57			
	c.3466+2T>G	- n Glu131AsofsTer11					28	
	c.550G>A	p.Ala184Thr					6	
CATAL (MAL 003040)	c.640+6A>G	-	48	51	100			_
GATA2 (NM 032638.4)	c.481C>G	p.Pro161Ala		58 46	58			
	c.490G>A	p.Ala164Thr				57		
PHF6 (NM 032458.2)	c.871G>A	p.Gly291Arg					8	
CEBPA (NM 004364.3)	c.563 564insCTC	p.Pro188 Pro189insSer			7			
	c.566C>A c.568T>C	p.Pro189HIS p.Ser190Pro			8 7			
	c.584 589dupACCCGC	p.His195 Pro196dup			33			4
MYD88 (NM 001172567.1) NPM1 (NM 002520.6)	c.577C>T c.859 860insTCTG	p.Arg193Trp p.Trp288CvsfsTer12	47				36 33	-
CBLC (NM 012116.3)	c.1303C>T	p.Pro435Ser		32			33	
KIT (NM 000222.2) CSE38 (NM 156039.2)	c.1621A>C	p.Met541Leu n.Are698Cvs		49 51	50			-
	c.2093G>A	p.Arg698His	32	51				
NOTCUS DIAL OTTOTAL	c.2509G>A	p.Asp837Asn				19		<u> </u>
NUTCHI (NM 017617.3)	c.5011G>A c.6853G>A	p.vai1671lle p.Val2285lle			56	47		
CBL (NM 005188.3)	c.1186T>C	p.Cys396Arg					23	
	c.1246T>G c.1259G>A	p.Cys416Gly p.Arg420Gln					43 7	
CDKN2A (NM 001195132.1)	c.442G>A	p.Ala148Thr					48 48	
1P53 (NM 000546.5) PTEN (NM 000314.4)	c.215C>G c.768G>T	p.ero72Arg p.Glu256Asp	54	ая			9	-
SET8P1 (NM (015559.2)	c 2612T>C	n ile871Thr					9	-