

Therapeutic effect of androgen therapy in a mouse model of aplastic anemia produced by short telomeres

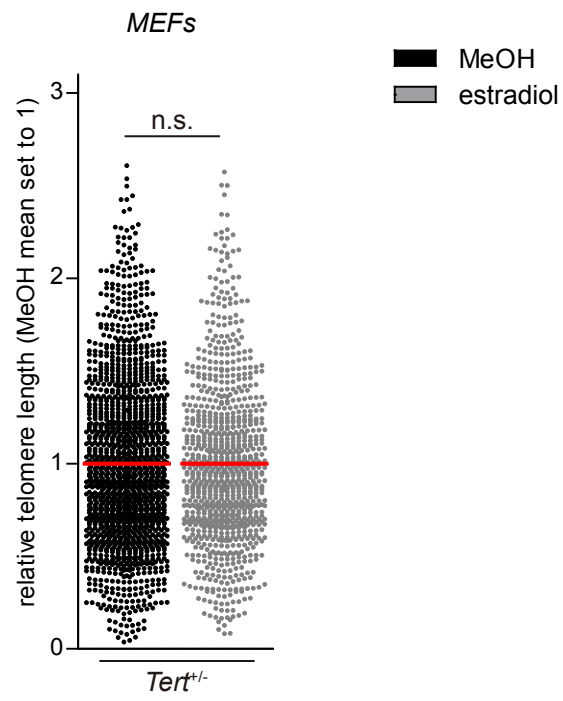
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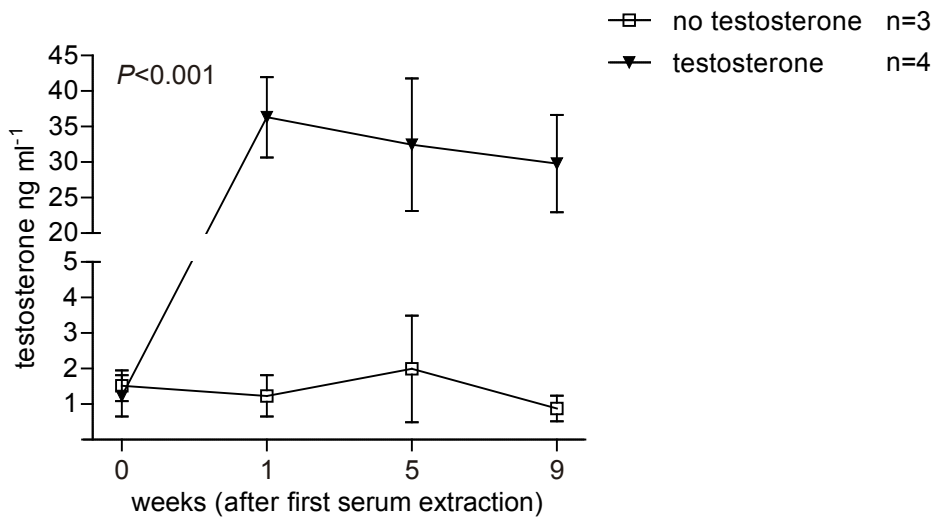
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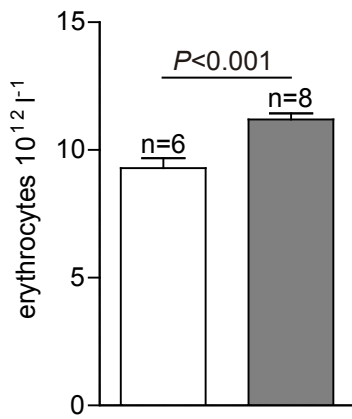
1 **Supplementary Figure 1 | Q-FISH analysis in *Tert*^{+/-} MEFs.** Relative TL (arbitrary
2 units of fluorescence) of *Tert*^{+/-} MEFs incubated with 1 μ M estradiol for 4 passages. n =
3 899 telomeres for estradiol and n = 1576 telomeres for methanol treatment. Two-sided
4 Student's *t*-test was used for statistical analysis. n.s. = not significant.

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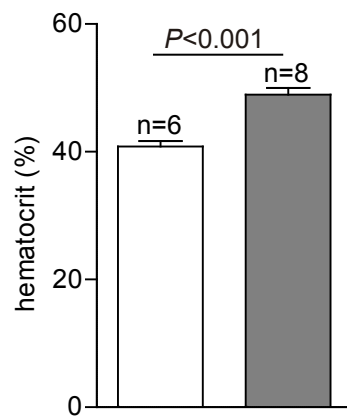
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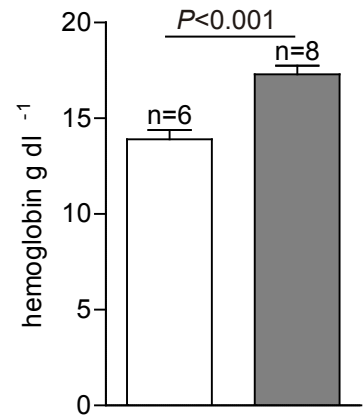
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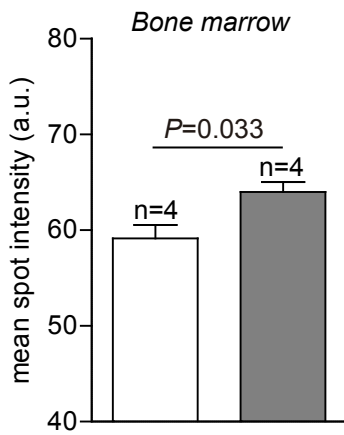
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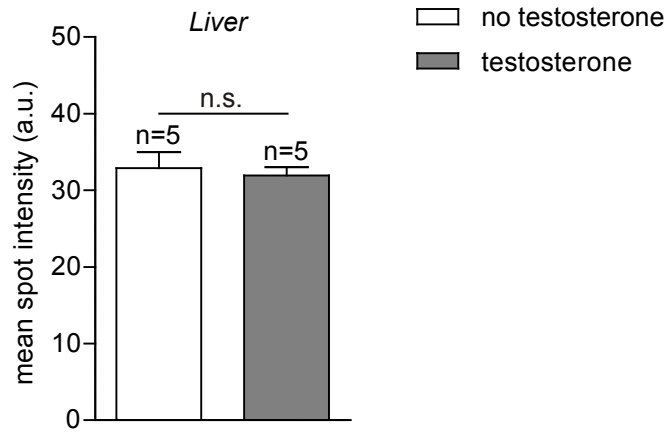
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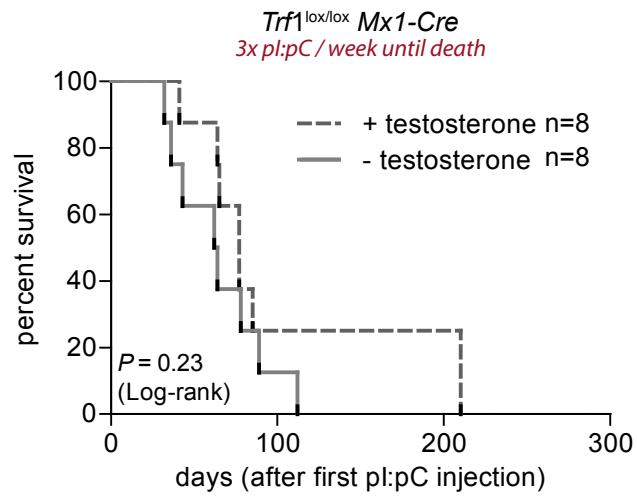
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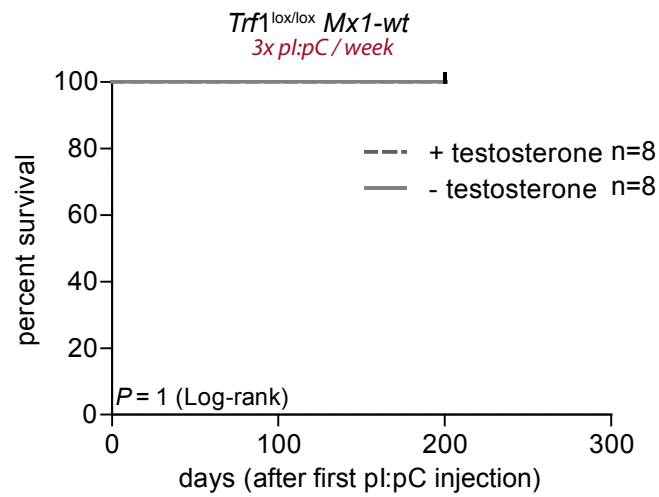
1 **Supplementary Figure 2 | Blood analysis of pl:pC treated mice after testosterone**
2 **administration.** (A) Time course of serum testosterone concentration in mice implanted
3 with a slow release testosterone pellet and mice without. Serum for ELISA analysis was
4 extracted at indicated time points. (B) Erythrocyte count, (C) hematocrit and (D)
5 hemoglobin levels in mice with or without testosterone treatment. (E) TL determined in
6 Q-FISH analysis on bone marrow cross sections from mice with or without testosterone
7 therapy represented as mean telomere spot intensity (arbitrary units of fluorescence,
8 a.u.). (F) Q-FISH TL analysis represented as mean telomere spot intensity in liver tissue
9 section from the same mice as in (E). n = number of mice. Graphs show mean values,
10 error bars indicate s.e.m. Two-way ANOVA test was used for statistical analysis in
11 figure A, two-sided Student's *t*-test was used for figures B-F. *P*-values are indicated.

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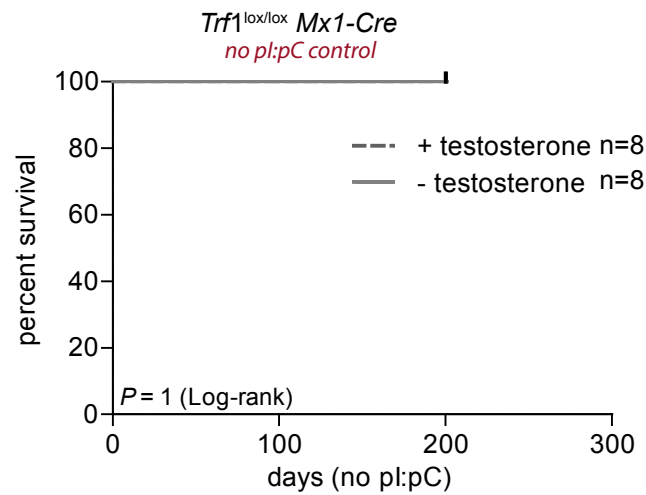
A



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1 **Supplementary Figure 3 | Survival of pl:pC treated control mice.** (A) Kaplan-Meier
2 survival curve of mice treated with pl:pC 3 times per week until death. In addition, mice
3 were treated with or without testosterone as indicated. (B) Kaplan-Meier survival curve
4 of *Trf1^{lox/lox} Mx1-wt* mice treated with pl:pC 3 times per week. In addition, mice were
5 treated with or without testosterone as indicated. Mice were sacrificed after 200 days.
6 (C) Kaplan-Meier survival curve of *Trf1^{lox/lox} Mx1-Cre* mice not treated with pl:pC. Mice
7 were treated with or without testosterone as indicated. Mice were sacrificed after 200
8 days. n = number of mice. Log-rank (Mantel-Cox) test was used for statistical analysis.
9 *P*-values are depicted.

1 **Supplementary Tables**

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Group	Genotype	Treatment
E1	<i>Trf1</i> ^{lox/lox} <i>Mx1-Cre</i>	2 pl:pC injection / week (continuous); +testosterone
C1	<i>Trf1</i> ^{lox/lox} <i>Mx1-Cre</i>	2 pl:pC injection / week (continuous); no testosterone
E2	<i>Trf1</i> ^{lox/lox} <i>Mx1-Cre</i>	3 pl:pC injection / week (continuous); +testosterone
C2	<i>Trf1</i> ^{lox/lox} <i>Mx1-Cre</i>	3 pl:pC injection / week (continuous); no testosterone
E3	<i>Trf1</i> ^{lox/lox} <i>Mx1-Cre</i>	3 pl:pC injection / week (for 4 weeks); + testosterone after pl:pC
C3	<i>Trf1</i> ^{lox/lox} <i>Mx1-Cre</i>	3 pl:pC injection / week (for 4 weeks); no testosterone
C4	<i>Trf1</i> ^{lox/lox} <i>Mx1-Cre</i>	no pl:pc; +testosterone
C5	<i>Trf1</i> ^{lox/lox} <i>Mx1-Cre</i>	no pl:pc; no testosterone
C6	<i>Trf1</i> ^{lox/lox} <i>Mx1-wt</i>	3 pl:pC injection / week (continuous); +testosterone
C7	<i>Trf1</i> ^{lox/lox} <i>Mx1-wt</i>	3 pl:pC injection / week (continuous); no testosterone

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11 **Supplementary Table 1: Experimental and control groups of mice subjected to**
 12 **this study.** After irradiation (12Gy) and bone marrow transplantation a total of 80 mice
 13 were stratified into 10 groups of 8 mice each which were subjected to different
 14 treatments as indicated. E denotes experimental group and C denotes control group.

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Primer name	Primer sequence (5'-3')
Actin-Forward	GGCACCACACCTTCTACAATG
Actin-Reverse	GTGGTGGTGAAGCTGTAG
Tert-Forward	GGATTGCCACTGGCTCCG
Tert-Reverse	TGCCTGACCTCCTCTTGTGAC

Supplementary Table 2: qPCR primers used in this work.

1 **Supplementary Methods:**

2 **Bone marrow transplantation, pl:pC and testosterone treatment.**

3 10 weeks old *Trf1lox/lox Mx1-Cre* and *Trf1lox/lox Mx1-wt* mice were used as bone
4 marrow donors for transplantation into 8 weeks old lethally (12Gy) irradiated wild-type
5 mice as previously described (Samper et al., 2002). A total of 2 million cells were
6 transplanted via tail vein injection at a donor:recipient ratio of 1:8 and mice were left for
7 a latency period of 30 days to allow bone marrow reconstitution. To induce *Cre*
8 expression, mice were intraperitoneally injected with polyinosinic-polycytidylic acid
9 (pl:pC; Sigma-Aldrich) (15 ug/g body weight). For androgen therapy mice were
10 subcutaneously implanted with a 90-days testosterone slow release pellet (Innovative
11 Research of America). After 90 days into treatment, testosterone pellets were renewed.
12 To control for potential adverse affects of pl:pC and testosterone we included mice that
13 were untreated, treated with testosterone alone or mice without *Cre*. For details see
14 Supplementary Table_1.

15

16 **Telomere measurement**

17 For Q-FISH analysis tissues sections or metaphases were post fixed in 4%
18 formaldehyde for 5 min, washed 3 x 5 min in PBS and incubated at 37°C for 15 min in
19 pepsin solution (0.1% Porcine Pepsin, Sigma; 0.01M HCl, Merck). After another round
20 of washes and fixation as mentioned above, slides were dehydrated in a 70%–90%–
21 100% ethanol series (5 min each). Slides were 10 min air-dried and 30 µl of telomere
22 probe mix added to each slide (10mM TrisCl pH 7, 25mM MgCl₂, 9mM citric acid,

1 82mM Na₂HPO₄, 70% deionized formamide (Sigma), 0.25% blocking reagent (Roche)
2 and 0.5 mg/ml Telomeric PNA probe (Panagene)), a cover slip added and slides
3 incubated for 3 min at 85 °C, and for further 2 h at room temperature in a wet chamber
4 in the dark. Slides were washed 2 x 15 min in 10mM TrisCl pH 7, 0.1% BSA in 70%
5 formamide under vigorous shaking, then 3 x 5 min in TBS 0.08% Tween20, and then
6 incubated in a 40,6-diamidino-2-phenylindole (DAPI) bath (4 mg/ml 1 DAPI (Sigma) in
7 PBS) before mounting samples in Vectashield (VectorTM). Confocal image were
8 acquired as stacks every 0.5 μm for a total of 1.5 μm using a Leica SP5-MP confocal
9 microscope and maximum projections were done with the LAS-AF software. Telomere
10 signal intensity was quantified using Definiens software.

11 HT-Q-FISH on peripheral blood leukocytes was done using 120–150 μl blood extracted
12 from the facial vein. Red blood cells were lysed (Erythrocyte lysis buffer, Qiagen) and
13 30–90 k leukocytes were plated in duplicate into clear-bottom, black-walled 96-well
14 plates pre-coated for 30 min with 0.001% poly-L-lysine. Plates were incubated at 37°C
15 for 2 h and fixed with methanol/acetic acid (3:1, v/v) 2 x 10 min and then overnight at -
16 20°C. Fixative was removed, plates dried for at least 1 h at 37°C and samples were
17 rehydrated in PBS. Plates were then subjected to a standard Q-FISH protocol (see
18 above) using a telomere-specific PNA-CY3 probe; DAPI was used to stain nuclei. Sixty
19 images per well were captured using the OPERA (Perkin Elmer) High-Content
20 Screening system. TL values were analysed using individual telomere spots (>10,000
21 telomere spots per sample). The average fluorescence intensities of each sample were
22 converted into kilobase using L5178-R and L5178-S cells as calibration standards,

1 which have stable TLs of 79.7 and 10.2 kb, respectively. Samples were analysed in
2 duplicate, or triplicate in the case of calibration standards.

3