Deficiency in mouse hyaluronidase 2: a new mechanism of chronic thrombotic microangiopathy

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SUPPLEMENTAL DATA

SUPPLEMENTAL METHODS

.*Platelet phenotype*. Platelet-rich fractions were prepared as described³⁴ using ACD (acid, citrate, dextrose) and apyrase (0,5 U/ml) as anticoagulants. ADP (10 μM) and thrombin (1 U/ml) were used to produce platelet partial and total activation, respectively. Platelet morphology was examined using electronic microscopy (JEOL and Philips Tecnai 10) observation as described elsewhere.³⁴ In addition, the amount of platelet microparticles was measured using a BD Accuri C6 flow cytometer as described elsewhere.³⁴The following reagents were used: Megamix Beads (Biocytex), PE-rat anti mouse CD41 (BD Pharmingen, 558040), and FITC-Annexin V (BD Biosciences).

Splenectomy. Control and Hyal2-/- mice were anesthetized intraperitonally using a mixture of medetomidin (Medetor® 1mg/ml, Virbac Animal Health Belgium, Wavre, Belgium; 0.25 mg/kg) and ketamine HCI (Ceva®, Brussel, Belgium; 40 mg/kg). The mice were placed on a heating table to maintain body temperature during all the procedure. After asepsis, a small incision was made in the left flank region, and the spleen was gently removed. One month after splenectomy, mice were injected with sulfo-NHS-LC-biotin to measure RBC survival.

Anti-C5 administration. Mice received biweekly intraperitonal injections of 750 μg anti-mouse C5 monoclonal antibodies (BB5.1) or control isotype-matched antibodies (both a kind gift of Dr Paul Tamburini, Alexion Pharmaceuticals, Cheshire, CT, USA) for 3 weeks. Blood samples were then collected to evaluate blood parameters and to confirm that treatment with anti-C5 led to decreased complement activity, using a hemolytic assay as described.²⁰

Glycocalyx staining in myocardial microvessels. Endothelial glycocalyx was fixed and stained with Alcian blue 8GX as described elsewhere. Briefly, anesthetized mice were cannulated retrogradely at the aorta/brachiocephalic trunk bifurcation and the vena cava was transected. Three successive solutions were perfused at a constant pressure of 33 ± 5 mmHg: first, calcium-free cardioplegic solution (BSA 0.1% for 3 min); second, phosphate buffered fixative (pH 7.4) containing 30 mM MgCl₂ for 2 min; third, the same solution + 0.05% Alcian Blue 8GX (Sigma) for 30 min. Left ventricular wall was then cut into 2-mm segments. Samples were fixed as usual for transmission microscopy which was performed using a Tecnai instrument.

Additional serum assays. Serum creatinine was measured using Crea+ (Roche Diagnostics), urine creatinine using Urine Mouse Creatinine Assay Kit (Crystal Chem Inc., Downers Grove, IL, USA), and urine albumin using Mouse Albumin Elisa Kit (Aviva Systems Biology, San Diego, CA, USA).

SUPPLEMENTAL REFERENCES

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SUPPLEMENTAL TABLES
Supplemental Table 1. Corpuscular indices in inbred mice.

	Control	Hyal2 ^{-/-}	P
	(N=5)	(N=5)	
RBC (10 ⁶ cells/µl)	9.05 ± 0.23	7.23 ± 0.48	*
Hb (g/dl)	12.8 ± 0.38	11.4 ± 0.68	NS
HCT (%)	38.4 ± 1.06	34.3 ± 1.23	*
Ret (%)	3.75 ± 0.28	16.5 ± 2.87	**
MCV (fL)	42.4 ± 0.57	48.0 ± 1.73	*
MCH (pg)	14.2 ± 0.23	15.9 ± 0.26	*
MCHC (g/dl)	33.4 ± 0.40	33.2 ± 1.02	NS
CHCM (g/dl)	34.2 ± 0.56	34.6 ± 0.98	NS
CH (pg)	14.6 ± 0.13	16.5 ± 0.18	*
RDW (%)	13.8 ± 0.29	15.4 ± 0.93	NS
HDW (g/dl)	2.60 ± 0.07	3.40 ± 0.12	*

RBC, red blood cell count; *Hb*, hemoglobin; *HCT*, hematocrit; *Ret*, reticulocytes; *MCV*, mean corpuscular volume; *MCH*, mean corpuscular hemoglobin; *MCHC*, mean corpuscular hemoglobin concentration (calculated); *CHCM*, corpuscular hemoglobin concentration mean; *CH*, corpuscular hemoglobin content; *RDW*, red cell volume distribution width; *HDW*, hemoglobin concentration distribution width. Comparisons using Mann Whitney test. *NS*, non significant; *, P < 0.05; **, P < 0.01.

Supplemental Table 2. Weight of various organs in male mice.

	Control	Hyal2 ^{-/-}	P
	(N=10)	(<i>N</i> =10)	
Animal (g)	33.1 ± 1.8	33.8 ± 1	NS
Heart (g)	0.158 ± 0.007	0.182 ± 0.006	*
Liver (g)	1.560 ± 0.098	1.553 ± 0.046	NS
Spleen (g)	0.095 ± 0.007	0.274 ± 0.022	***
Left Kidney (g)	0.220 ± 0.012	0.215 ± 0.015	NS
Right Kidney	0.224 ± 0.011	0.227 ± 0.012	NS
(9)			

Each value represents the mean ± SEM. P values for the differences between control and Hyal2-/- mice were calculated using unpaired Student's t-tests. NS, non significant; *, P<0.05; ***, P<0.001. Female *Hyal2-/-* mice had similar alterations compared with male *Hyal2-/-* mice (data not shown).

Supplemental Table 3. Effect of anti-C5 antibodies.

	Before injections		After injections						
				Control mAb		Anti-C5 mAb			
	Control	Hyal2 ^{-/-}		Control	Hyal2-/-		Control	Hyal2-/-	
	(<i>N</i> =6)	(<i>N</i> =14)	P	(N=3)	(<i>N</i> =7)	P	(<i>N</i> =3)	(<i>N</i> =6)	P
WBC	4.97	6.54	NS	6.47	5.27	NS	5.85	5.99	NS
(10³/µl)	± 0.90	± 0.87	INO	± 1.34	± 0.57	INO	± 0.76	± 0.35	140
RBC	9.57	7.83	**	9.72	7.52	*	10.02	7.26	*
(10 ⁶ /µI)	± 0.18	± 0.25		± 0.46	± 0.52		± 0.25	± 0.32	
5. 7									
PLT	1237	416	***	1174	507	*	1459	426	*
(10³/µl)	± 57	± 40		± 75	± 90		± 193	± 101	
Ret	2.57	10.48	***	2.61	12.00	*	2.72	11.74	*
(%)	± 0.13	± 0.69		± 0.12	± 1.58		± 0.30	± 0.76	

Blood parameters before and after 6 biweekly ip injections of control (isotype-matched) or anti-C5 monoclonal antibodies (mAb) in *control* and *Hyal2*-/- mice. One mouse was excluded from the active group because its plasma hemolytic activity was not decreased by the anti-C5 mAb. *WBC*, white blood cell count; *RBC*, red blood cell count; *PLT*, platelet count; *Ret*, reticulocytes. Each value represents the mean ± SEM. *P* values for the differences between *control* and *Hyal2*-/- mice were calculated using Mann-Whitney test; *NS*, non significant; *, *P*<0.05; **, *P*<0.01; ***, *P*<0.001.

Supplemental Table 4. Serum creatinine and urine albumin-to-creatinine ratio in outbred mice.

	6 months (<i>N</i> =	8)	11 months (<i>N</i> =6)			
	Control	Hyal2⁻ [/] -	P	Control	Hyal2 ^{-/-}	P
Serum creatinine (mg/dl)	0.41 ± 0.02	0.39 ± 0.02	NS	0.41 ± 0.03	0.44 ± 0.03	NS
Urine alb/creat (g/mol)	3.80 ± 0.29	4.43 ± 0.78	NS	4.66 ± 0.36	5.99 ± 1.21	NS

Statistical comparisons using unpaired t test. NS, non significant.

SUPPLEMENTAL FIGURE LEGENDS

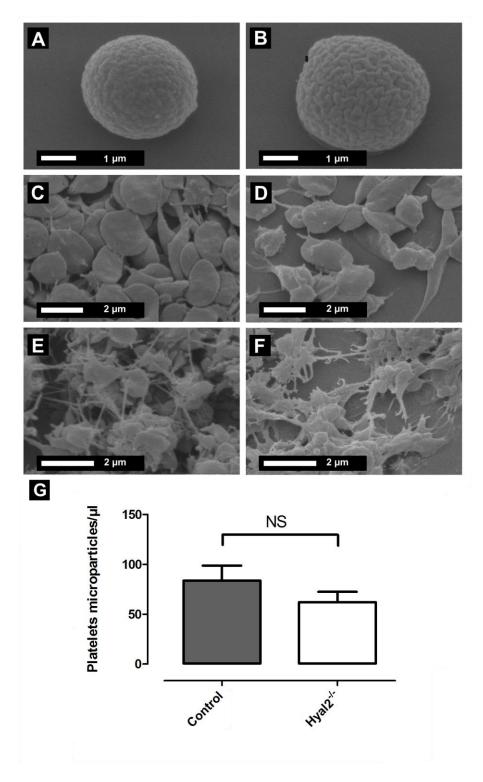
Supplemental Figure 1. Platelet Phenotype. (A-F) Platelet morphology observed using scanning electronic microscopy. (A-B) Morphological discoid appearance of washed platelets in *control* (A) and *Hyal2*-/- (B) mice. (C-F) Morphological changes of washed platelets during ADP (C-D) or thrombin (E-F) induced aggregation in *control* (C and E) and *Hyal2*-/- (D and F) mice. (G) Flow cytometric measurement of platelets microparticules in *control* (N=5) and *Hyal2*-/- (N=6) mice. Means ± SEM are shown. There is no difference (NS) between genotypes.

Supplemental Figure 2. Effect of splenectomy on RBC half-life. Control and Hyal2^{-/-} outbred male mice were splenectomized. One month post-surgery, they were injected with sulfo-NHS-LC-biotin and 5 μl of blood were taken at the indicated time points. The percentage of biotinylated RBCs (biotin+ RBCs) was calculated as the ratio of positive cells to all RBCs in flow cytometry. The RBC survival of splenectomized *control* mice (▲) and Hyal2^{-/-} mice (△) is very close to the survival of endogenous RBCs in the respective, non-operated animals (control, ●; Hyal2^{-/-}, ○). Each value represents the mean ± SEM of 3 mice in each group. There was no significant difference between RBC survival in splenectomized and non-splenectomized mice (two-way ANOVA).

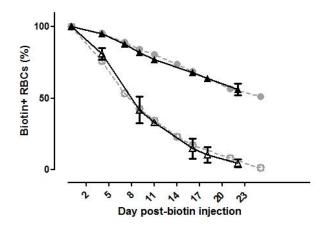
Supplemental Figure 3. Glycocalyx morphology in microvessels. Endothelial glycocalyx (stained in black using Alcian Blue 8GX) was observed using transmission electronic microscopy in myocardial microvessels. Glycocalyx was structurally intact in *Hyal2*-/- (B & D) compared with *control* (A & C) mice

SUPPLEMENTAL FIGURES

Supplemental Figure 1



Supplemental Figure 2



Supplemental Figure 3

