Chem. 1999;274(30):20826-20832.

- 3. Silvestri L, Nai A, Pagani A, Camaschella C. The extrahepatic role of TFR2 in iron homeostasis. Front Pharmacol. 2014;5:93.
- 4. Worthen CA, Enns CA. The role of hepatic transferrin receptor 2 in the regulation of iron homeostasis in the body. Front Pharmacol. 2014;5:34.
- 5. Lane DJ, Merlot AM, Huang ML, et al. Cellular iron uptake, trafficking and metabolism: key molecules and mechanisms and their roles in disease. Biochim Biophys Acta. 2015;1853(5):1130-1144.
- 6. Ginzburg Y, Rivella S. β-thalassemia: a model for elucidating the dynamic regulation of ineffective erythropoiesis and iron metabolism. Blood. 2011;118(16):4321-4330.
- 7. Li H, Rybicki AC, Suzuka SM, et al. Transferrin therapy ameliorates disease in beta-thalassemic mice. Nat Med. 2010;16(2):177-182.
- 8. Gelderman MP, Baek JH, Yalamanoglu A, et al. Reversal of hemochromatosis by apotransferrin in non-transfused and transfused Hbbth3/+ (heterozygous b1/b2 globin gene deletion) mice. Haematologica. 2015;100(5):611-622.
- 9. Gardenghi S, Ramos P, Marongiu MF, et al. Hepcidin as a therapeutic tool to limit iron overload and improve anemia in β-thalassemic mice. J Clin Invest. 2010;120(12):4466-4477.
- 10. Bartnikas TB, Andrews NC, Fleming MD. Transferrin is a major determinant of hepcidin expression in hypotransferrinemic mice. Blood. 2011;117(2):630-637.
- 11. Ganz T. Systemic iron homeostasis. Physiol Rev. 2013;93(4):1721-1741.
- 12. Babitt JL, Lin HY. The molecular pathogenesis of hereditary hemochromatosis. Semin Liver Dis. 2011;31(3):280-292.
- 13. Kautz L, Jung G, Valore EV, Rivella S, Nemeth E, Ganz T. Identification of erythroferrone as an erythroid regulator of iron metabolism. Nat Genet. 2014;46(7):678-684.
- 14. Nai A, Lidonnici MR, Rausa M, et al. The second transferrin receptor regulates red blood cell production in mice. Blood. 2015;125(7):1170- 1179.
- 15. Forejtnikovà H, Vieillevoye M, Zermati Y, et al. Transferrin receptor 2 is a component of the erythropoietin receptor complex and is required for efficient erythropoiesis. Blood. 2010;116(24):5357-5367.
- 16. Pagani A, Vieillevoye M, Nai A, et al. Regulation of cell surface transferrin receptor-2 by iron-dependent cleavage and release of a soluble form. Haematologica. 2015;100(4):458-465.
- 17. Chiabrando D, Vinchi F, Fiorito V, Mercurio S, Tolosano E. Heme in pathophysiology: a matter of scavenging, metabolism and trafficking across cell membranes. Front Pharmacol. 2014;5:61.
- 18. Vinchi F, De Franceschi L, Ghigo A, et al. Hemopexin therapy improves cardiovascular function by preventing heme-induced endothelial toxicity in mouse models of hemolytic diseases. Circulation. 2013;127(12):1317-1329.
- 19. Schaer DJ, Vinchi F, Ingoglia G, Tolosano E, Buehler PW. Haptoglobin, hemopexin, and related defense pathways-basic science, clinical perspectives, and drug development. Front Physiol. 2014 2014;5:415.

Jekyll and Hyde: the role of heme oxygenase-1 in erythroid biology

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There is currently a great deal of excitement regard-
ing the role of stem cell niches that regulate
hematopoietic stem cell self-renewal and differen-
tiation. It should be noted, however, that the original ing the role of stem cell niches that regulate hematopoietic stem cell self-renewal and differentiation. It should be noted, however, that the original description of a hematopoietic niche actually occurred in 1958 when the French hematologist, Marcel Bessis, described erythroblastic islands. ¹ The island was characterized by developing erythroblasts surrounding a central macrophage and, based on careful structural observations, Bessis and colleagues made a number of interesting inferences about the role of central macrophages in erythropoiesis. It was suggested that the macrophage functions as a "nurse" cell, providing iron to developing erythroblasts for heme synthesis and, furthermore, that the extruded nuclei produced at the end of erythroid differentiation are phagocytized by these central macrophages. 2 These concepts proved prescient as they have been supported by a number of recent findings which have shown that the macrophage-erythroblast interactions mediated by a large number of adhesion molecules are essential for the highly regulated process of erythroblast proliferation and survival which is necessary for the production of two million reticulocytes per second. 3-6 In this context the recent findings that *in vivo* depletion of erythroblastic island macrophages blocks erythroblast proliferation and maturation fully validates the central role of macrophages in regulating erythropoiesis. 7

Apart from playing an important role in the genesis of red blood cells within the bone marrow, macrophages of the reticulocyte endothelial system in general and spleen in particular play a critical role in quality control by removing senescent and damaged red cells from the circulation. 8-10 Thus different macrophage subsets play a dual

role in both the production of red cells and in the elimination of senescent normal red cells and pathological red cells. This important symbiotic interrelationship between erythroid and macrophage biology is receiving increasing attention in hematology research since the findings of the studies have direct relevance to our understanding of both normal and disordered erythropoiesis.

In this issue of Haematologica, Fraser and colleagues describe exciting new findings regarding a key role for heme-oxygenase-1 in both regulating erythroid differentiation and in mediating clearance of circulating red cells through its effect on macrophages. ¹¹ The work of Fraser *et al.* documents that heme-oxygenase-1 deficiency adversely affects steady-state erythropoiesis in murine bone marrow due to a diminished ability of erythroblasts to form erythroblastic islands. The reduction in erythroblastic islands was the result of decreased numbers of the subtype of bone marrow macrophages involved in island formation. These observations reinforce the concept of an essential requirement of a specific subset of macrophages for the formation of bone marrow erythroblastic islands and that island formation is necessary to sustain normal bone marrow erythropoiesis. Interestingly, the decreased erythropoiesis in the bone marrow led to increased erythropoiesis in the spleen, a common compensatory response in the murine system in which the spleen is the major erythropoietic organ that responds to stress erythropoiesis.

While heme-oxygenase-1 deficiency had a negative effect on bone marrow erythropoiesis, it had a positive effect on red cell life span in circulation as a result of compromised ability of the macrophages of the reticuloendothelial system to remove senescent red cells. It thus appears that heme-oxygenase-1 plays the role of Dr. Jekyll by increasing red cell life span in circulation and also plays the part of Mr. Hyde by decreasing bone marrow erythropoiesis.

The work of Fraser *et al.* represents an important step in our understanding of the complex interplay between erythroid and macrophage biology in the regulation of red cell production and destruction. In particular, it brings to our attention the previously unsuspected and distinct roles of heme-oxygenase-1 in murine erythroid biology through its action on macrophages. However, many questions remain. How does heme-oxygenase-1 deficiency account for the observed microcytosis and decreased hemoglobin content of red cells? Is there perturbation of iron homeostasis due to dysregulation of hepcidin production?12 Importantly, do the reported findings using the murine system account for the hematologic phenotype noted in the very rare cases of human heme-oxygenase-1 deficiency?13,14

What then are the implications of these current findings? One is that heme-oxygenase-1 may play a much broader role in erythroid biology than previously suspected and likely plays a role in a number of human red cell disorders. A second implication is that there is clearly a complex interplay of cell-cell interactions in regulating various biological functions. Finally, the work of Fraser *et al.* gives us a valuable impetus to further explore the complex role of macrophages in various aspects of erythroid biology.

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References

- 1. Bessis M. L'ilot erythroblastique. Unite functionelle de la moelle osseuse. Rev Hematol. 1958;13(1):8-11.
- 2. Bessis MC, Breton-Gorius J. Iron metabolism in the bone marrow as seen by electron microscopy: a critical review. Blood. 1962;19(6):635- 663.
- 3. Rhodes MM, Kopsombut P, Bondurant MC, et al. Adherence to macrophages in erythroblastic island enhances erythroblast proliferation and increases erythrocyte production by a different mechanism than erythropoietin. Blood. 2008;111(3):1700-1708.
- 4. Chasis JA, Mohandas N. Erythroblastic islands: niches for erythropoiesis. Blood. 2008;112(3):470-478.
- 5. Jacobsen RN, Perkins AC, Levesque J-P. Macrophages and regulation of erythropoiesis. Curr Opin Hematol. 2015;22(3):212-219.
- 6. Korolnek T, Hamza I. Macrophages and iron trafficking at birth and death of red cell. Blood. 2015; March 16 [Epub ahead of print]
- 7. Jacobsen RN, Forristal CE, Raggatt LJ, et al. Mobilization with granulocyte colony-stimulating factor blocks medullary erythropoiesis by depleting F4/80+VCAM1+CD169+ER-HR+Ly6G+ erythroid island macrophages in the mouse. Exp Hematol. 2014;42(7):547-561.
- 8. Clark MR. Senescence of red blood cells: progress and problems. Physiol Rev. 1988;68(2):503-554.
- 9. Arashiki N, Kimata N, Manno S, et al. Membrane peroxidation and methemoglobin formation are both necessary for band 3 clustering: mechanistic insights into erythrocyte senescence. Biochemistry. 2013;52(34):5760-5769.
- 10. Safeukui I, Buffet P, Delpaine G, et al. Quantitative assessment of sensing and sequestration of spherocytic erythrocytes by human spleen: implications for understanding clinical variability of membrane disorders. Blood. 2012;120(2):424-430.
- 11. Fraser ST, Midwinter RG, Coupland LA, et al. Heme oxygenase-1 deficiency alters erythroblastic island formation, steady-state erythropoiesis and red blood cell lifespan in mice. Haematologica. 2015; 100(5):601-610.
- 12. Kim A, Nemeth E. New insights into iron regulation and erythropoiesis. Curr Opin Hematol. 2015;22(3):199-205.
- 13. Yachie A, Niida Y, Wada T, et al. Oxidative stress caused enhanced endothelial injury in human heme oxygenase-1 deficiency. J Clin Invest. 1999;103(1):129-135.
- 14. Radhakrishnan N, Yadav SP, Sachdeva A, et al. Human heme oxygenase-I deficiency presenting with hemolysis, nephritis, and asplenia. J Pediatr Hematol Oncol. 2011;33(1):74-78.

Personalized medicine in myelodysplastic syndromes: wishful thinking or already clinical reality?

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General concepts of managing patients with myelodysplastic syndromes

This editorial will start with the important (and true) premise that the term myelodysplastic syndromes (MDS) covers a group of heterogeneous and complex hematologic disorders primarily found within the older population. In fact, its diversity makes the disease challenging and "truly personalized", not only in terms of diagnostics but also in carrying out clinical decision-making. The heterogeneity of MDS manifests in the individual patient as a disease ranging from an indolent condition with a considerable life expectancy to forms approaching the aggressiveness of acute myeloid leukemia (AML). A risk-adapted treatment strategy is, therefore, mandatory for a disease showing such a highly variable clinical course. Prognostic factors may be subdivided into those related to the patient's general char-

acteristics and health condition and those related to the MDS disease itself. During the past 15 years, treatment has been stratified according to the International Prognostic Scoring System (IPSS) risk score; i.e. into "lower-risk" MDS (low/int-1, LR-MDS), where correction of cytopenia was the main objective, and "higher-risk" MDS (int-2/high, HR-MDS), where the reduction or delay of progression or AML evolution and prolonged survival was the objective. More recently, a revised version of the IPSS has been introduced (IPSS-R) subdividing patients into 5 risk groups with different outcomes in terms of AML evolution and survival. Using this new IPSS-R, one-quarter of LR-MDS per classical IPSS were re-classified as having a higher risk, and may potentially require more intensive treatment, while on the other hand a substantial subset of HR-MDS patients per classical IPSS were re-classified as lower risk suggesting that IPSS-R can refine the scoring of an individual MDS patient. Nevertheless, it is still a subject of controversy as to how