

# Silenced B-cell receptor response to autoantigen in a poor-prognostic subset of chronic lymphocytic leukemia

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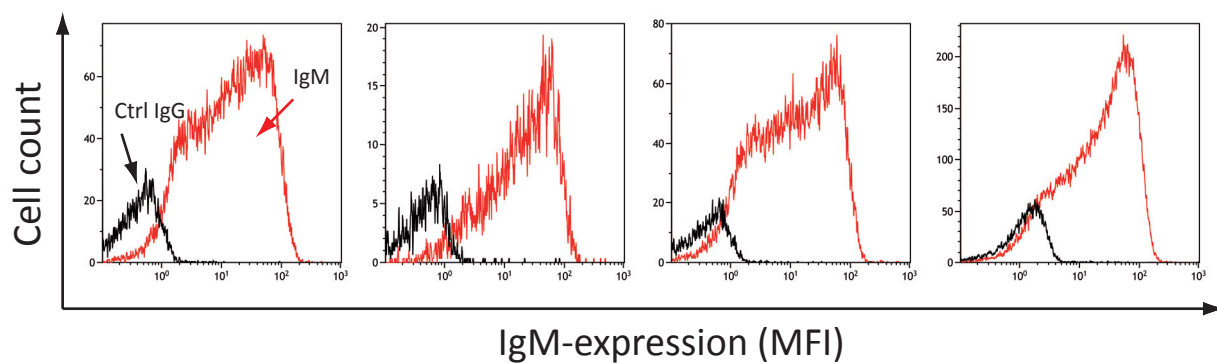
## Bergh et al. Supplementary Tables and Figures

**Supplementary Table 1. Antibodies and reagents**

Ab reactivity	Conjugate	Species/Clone	Company	Method used
CD5	PerCpCy5.5	Mouse/UCHT2	BioLegend	FC <sup>3</sup>
CD5	PE	Mouse/UCHT2	BD Biosciences	FC
CD19	PE-cy7	Mouse/HIB19	BioLegend	FC
CD19	PeCy5	Mouse/HIB19	BioLegend	FC
BrdU	FITC	Mouse/B44	BD Biosciences	FC
IgM	RPE	Mouse/M15/8	ABD Serotec	FC
CD36	PE	Mouse/CB38	BD Biosciences	FC
<b>CD36</b>	<b>APC/Cy7</b>	<b>Mouse/336213</b>	<b>BioLegend</b>	<b>FC</b>
SR-PSOX (CXCL16) <sup>1</sup>	-	Rat/256213	R&D systems	FC
SR-B1 <sup>2</sup>	-	Rabbit/Polyclonal	LifeSpan BioSciences	FC
CD25	APC	Mouse/BC96	BD Biosciences	FC
CD86	PE	Mouse/IT2.2	BD Biosciences	FC
Ig kappa light chain	-	Mouse/G20-193	BD Biosciences	FC
IgD	-	Mouse	DAKO	FC
IgM F(ab') <sub>2</sub>	-	Goat/Polyclonal	Southern Biotechnology	Stimuli
IgM	HRP	Rabbit/Polyclonal	DAKO	ELISA
IgM	ALP	Goat/Polyclonal	Sigma-Aldrich	Luminescence ELISA
Phospho-p44/42 MAPK (ERK1/2)(Thr202/Tyr204)	-	Rabbit	Cell Signaling Technology	WB <sup>4</sup>
P44/42 MAPK (ERK1/2)	-	Rabbit	Cell Signaling Technology	WB
Rabbit Ig	HRP	Goat/Polyclonal	DAKO	WB
<b>Other reagents</b>				
CpG oligodeoxynucleotide (ODN 10104, stimulatory CpG type B, human specific)	-		Coley Pharmaceutical Group and InVivogen	Stimuli
nLDL	Biotin		Gift from Sohvi Hörkkö	Stimuli/FC
MDA-LDL	Biotin		Gift from Sohvi Hörkkö	Stimuli/FC/IF <sup>5</sup>
MDA-BSA	Biotin		Gift from Sohvi Hörkkö	FC/IF
AnnexinV	Pacific blue or FITC		BioLegend/BD Biosciences	FC
Streptavidin	Alexa488		Molecular Probes	IF/FC
Fluo-4 AM	-		Invitrogen	FC

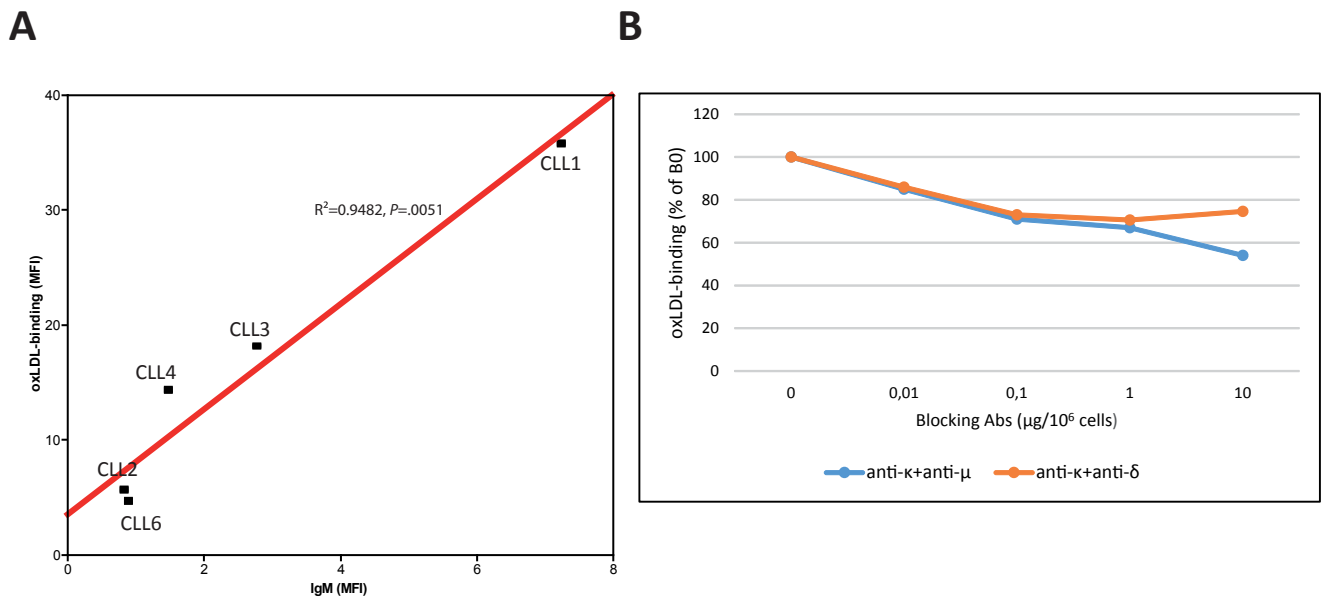
<sup>1</sup> SR-PSOX, Scavenger receptor for PhosphatidylSerine and Oxidized lipoproteins, <sup>2</sup> SR-B1, Scavenger receptor class B member 1, <sup>3</sup> FC, Flow cytometer, <sup>4</sup> WB, Western blot, <sup>5</sup> IF, Immunofluorescence

## Supplementary Figure 1



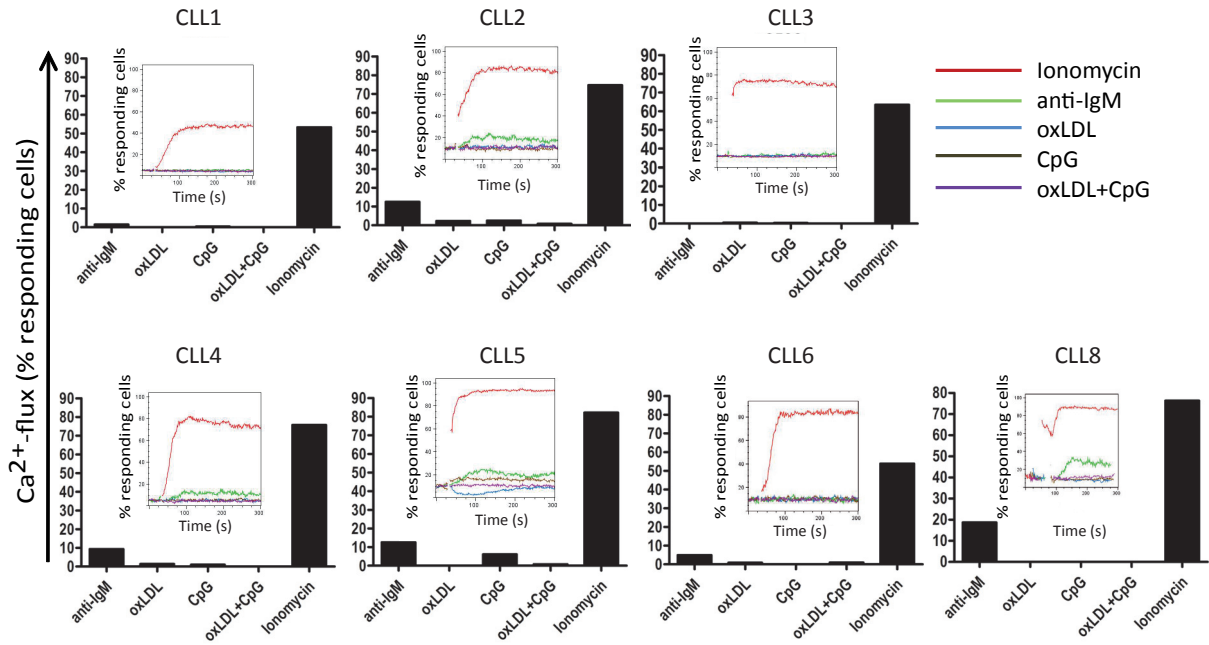
**Figure 1. Surface expression of IgM in normal B-cells.** IgM surface expression was analyzed in CD19<sup>+</sup> B-cells from four healthy individuals. (Mean MFI = 17; range 12-25).

## Supplementary Figure 2



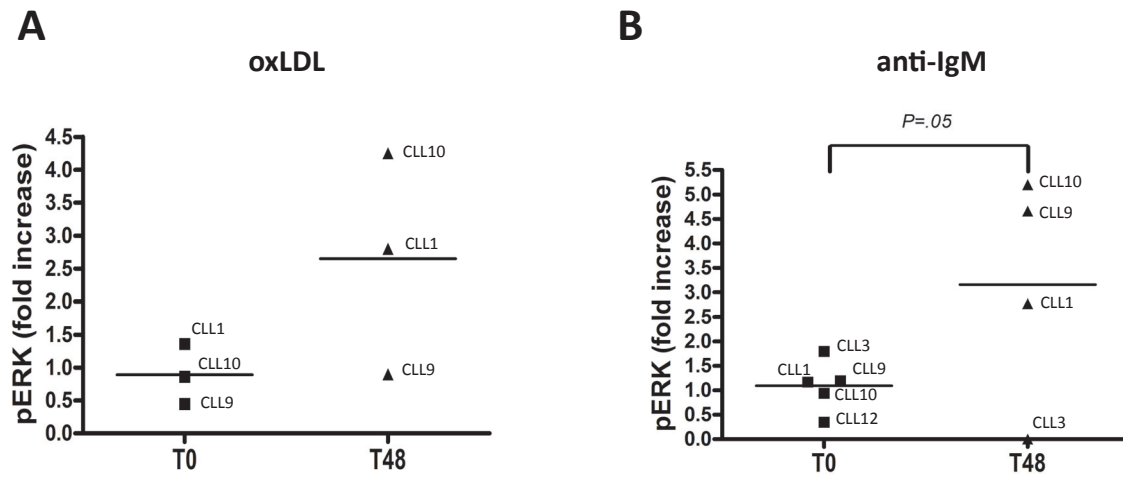
**Figure 2. Cell surface receptor analysis. (A)** The level of oxLDL-binding correlated with the level of sIgM-expression ( $R^2 = 0.948$ ,  $P = .005$ ,  $n = 5$ ) **(B)** oxLDL-binding was blocked in a dose-dependent manner by anti-IgM (anti- $\mu$  + anti- $\kappa$ ) or anti-IgD (anti- $\delta$  + anti- $\kappa$ ) Abs.

## Supplementary Figure 3



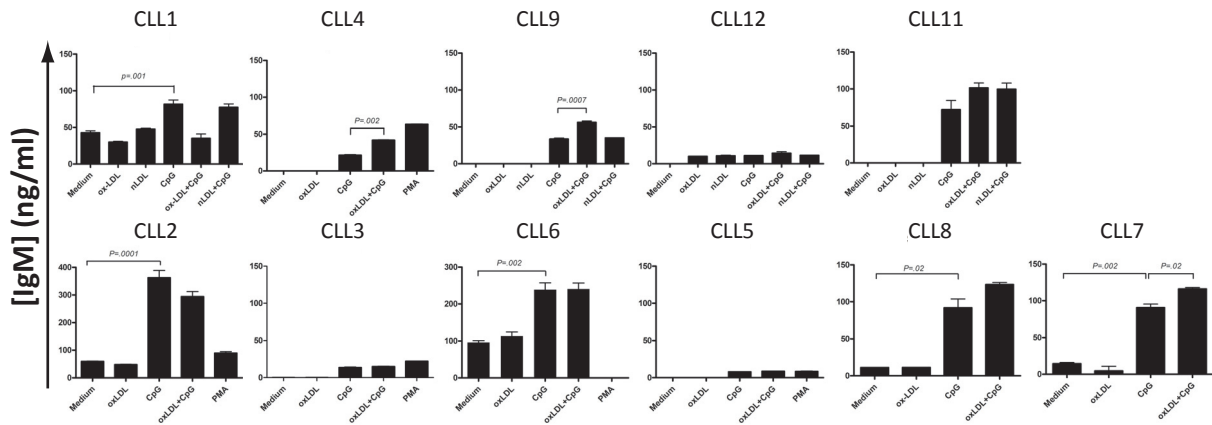
**Figure 3. oxLDL ligation does not induce  $Ca^{2+}$  - flux.** Subset #1 lymphocytes from 7 CLL cases were analyzed for  $Ca^{2+}$  -release. Cells were loaded with the  $Ca^{2+}$  sensitive dye Fluo-4 AM and analyzed by flow cytometry before and after addition of anti-IgM  $F(ab')_2$  (20  $\mu$ g/mL), oxLDL (50  $\mu$ g/mL) and/or CpG (10  $\mu$ g/mL). The calcium ionophore ionomycin was used as positive control. The diagrams show percent responding cells of total viable cells. The insert in each diagram show flow images for  $Ca^{2+}$  -flux responses.

## Supplementary Figure 4



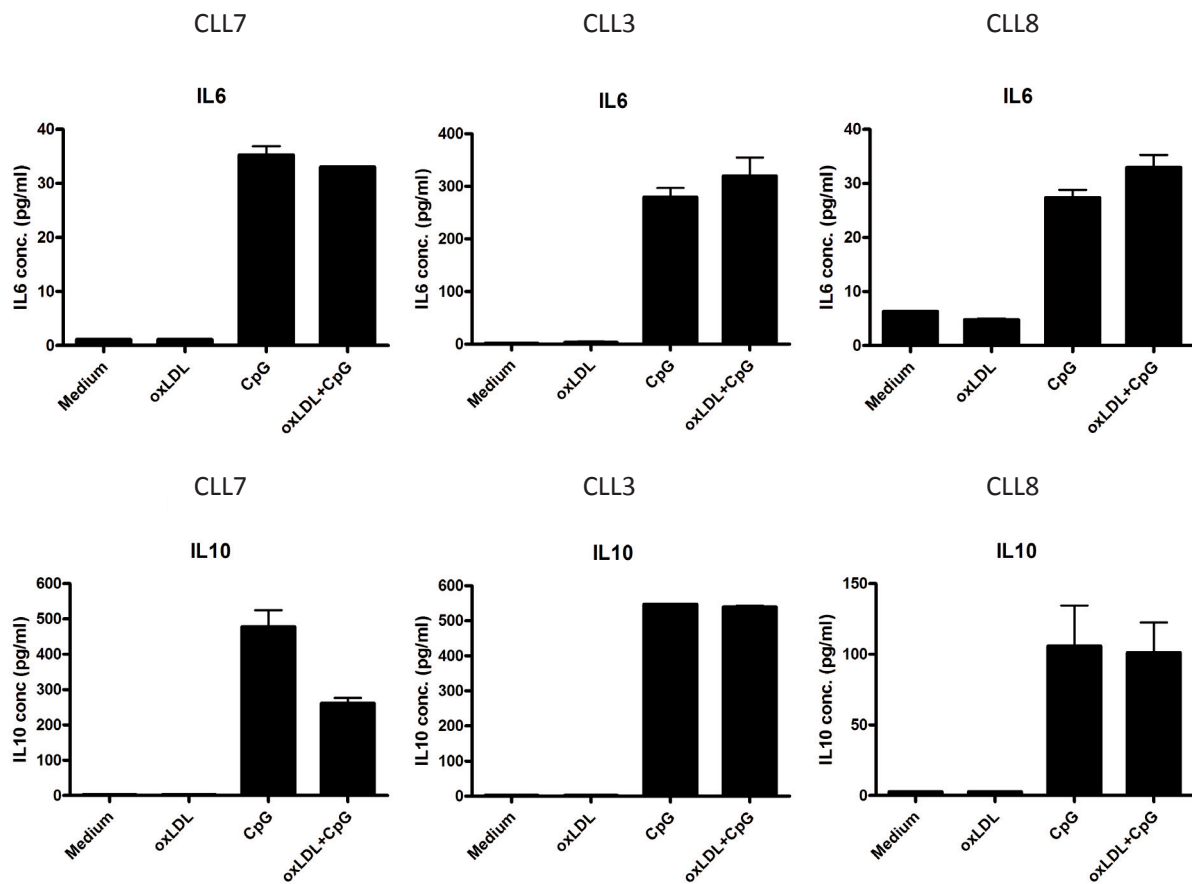
**Figure 4. Constitutive phosphorylation of ERK1/2 at basal level.** Cells were cultured for 48 hours and ERK1/2 phosphorylation was analyzed by immunoblotting immediately (T0) and after 48 hours (T48). Cells were stimulated at both time points with 50  $\mu\text{g}/\text{mL}$  oxLDL or 10  $\mu\text{g}/\text{mL}$  anti-IgM  $\text{F}(\text{ab}')_2$  for 5 minutes. The diagram show data on pERK/total ERK fold-increase. **(A)** after oxLDL exposure; **(B)** after anti-IgM exposure

## Supplementary Figure 5



**Figure 5. IgM release from CLL subset #1 cells upon BcR and TLR stimulation.** PBMC (45-98% CD5<sup>+</sup>/CD19<sup>+</sup>) from 11 subset #1 patients were cultured in the presence or absence of nLDL (25  $\mu$ g/mL), oxLDL (25  $\mu$ g/mL), anti-IgM F(ab')<sub>2</sub> (10  $\mu$ g/mL), PMA (50 ng/mL) and/or CpG (5  $\mu$ g/mL) for 72 hours. Culture supernatants were collected and analyzed for IgM content in ELISA. Data are expressed as mean  $\pm$  SEM.

## Supplementary Figure 6

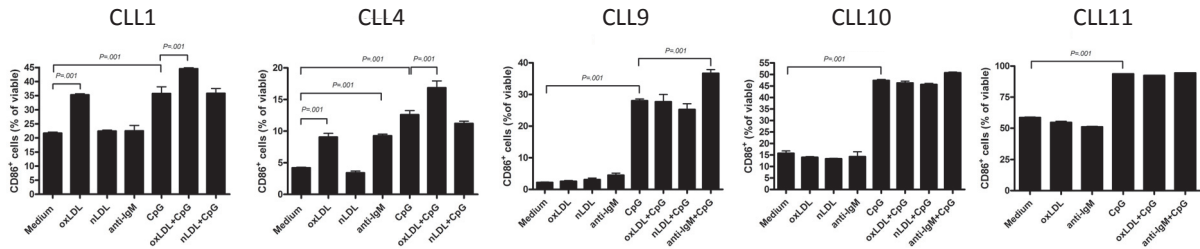


**Figure 6. Cytokine release from CLL subset #1 cells upon BcR and TLR stimulation.** CLL cells from three subset #1 CLL patients were cultured in the presence or absence of oxLDL (25  $\mu\text{g}/\text{mL}$ ), nLDL (25  $\mu\text{g}/\text{mL}$ ), anti-IgM F(ab')<sub>2</sub> (10  $\mu\text{g}/\text{mL}$ ), and/or CpG (5  $\mu\text{g}/\text{mL}$ ) for 72 hours. Culture supernatants were collected and analyzed for 10 different cytokines in Luminex-assay. Data for IL6 and IL10 are presented and the concentrations are expressed as mean  $\pm$ SEM.

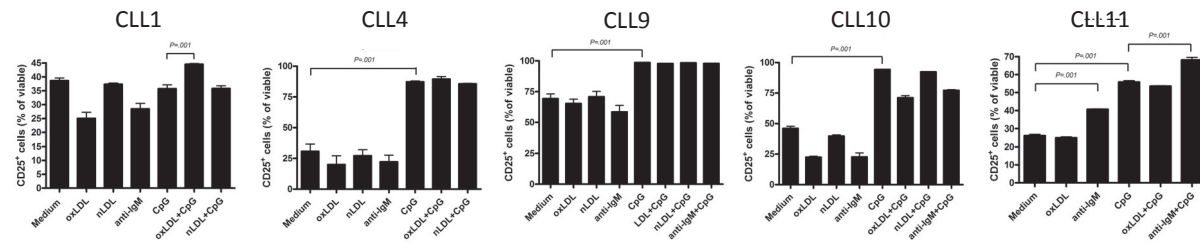


# Supplementary Figure 7

**A**



**B**



**Figure 7. Surface expression of CD25 and CD86 after BcR and TLR stimulation.** CLL subset #1 cells from five patients were cultured in the presence or absence of oxLDL (25  $\mu\text{g}/\text{mL}$ ), nLDL (25  $\mu\text{g}/\text{mL}$ ), anti-IgM F(ab')<sub>2</sub> (10  $\mu\text{g}/\text{mL}$ ), and/or CpG (5  $\mu\text{g}/\text{mL}$ ) for 24 hours. Surface expression of CD86 (**A**) and CD25 (**B**) was measured with flow cytometry. Data are expressed as mean  $\pm$  SEM.