

## GATA2 regulates differentiation of bone marrow-derived mesenchymal stem cells

Mayumi Kamata,<sup>1</sup> Yoko Okitsu,<sup>1</sup> Tohru Fujiwara,<sup>1,2</sup> Masahiko Kanehira,<sup>1</sup> Shinji Nakajima,<sup>1</sup> Taro Takahashi,<sup>1</sup> Ai Inoue,<sup>1</sup> Noriko Fukuhara,<sup>1</sup> Yasushi Onishi,<sup>1</sup> Kenichi Ishizawa,<sup>1</sup> Ritsuko Shimizu,<sup>3</sup> Masayuki Yamamoto,<sup>4</sup> and Hideo Harigae<sup>1,2</sup>

<sup>1</sup>Departments of Hematology and Rheumatology; <sup>2</sup>Molecular Hematology/Oncology; <sup>3</sup>Molecular Hematology; and <sup>4</sup>Medical Biochemistry, Tohoku University Graduate School of Medicine, Sendai, Japan

---

©2014 Ferrata Storti Foundation. This is an open-access paper. doi:10.3324/haematol.2014.105692

Manuscript received on February 12, 2014. Manuscript accepted on August 7, 2014.

Correspondence: harigae@med.tohoku.ac.jp

## **Supplementary Methods**

### **Antibodies**

Allophycocyanin (APC)-labeled human CD14, phycoerythrin (PE)-labeled human CD29, PE-labeled human CD34, fluorescein isothiocyanate (FITC)-labeled mouse/human CD44, FITC-labeled human CD45, FITC-labeled human CD90 and FITC-labeled mouse Sca-1 antibodies were purchased from BD Biosciences.

APC-labeled anti-human CD105, PE-labeled mouse CD29, FITC-labeled mouse CD11b, FITC labeled mouse CD34 and PE-labeled mouse CD45 antibodies were purchased from eBioscience.

### **Cytokine treatment**

BM-MSCs were differentiated into adipocyte-lineage cells, with the presence of cytokines as follows: 100 ng/mL each of recombinant human bone morphogenetic protein 4 (rhBMP4), human transforming growth factor- $\beta$ 1 (rhTGF $\beta$ 1), human interleukin (IL)-1 $\beta$ /IL-1F2z (rhIL-1 $\beta$ ), human IL-6 (rhIL-6), human interferon- $\gamma$  (rhINF $\gamma$ ), human tumor necrosis factor (TNF)- $\alpha$  (rhTNF $\alpha$ ), or human IL-17A (rhIL-17A). All cytokines were obtained from R&D Systems.

### **Cell cycle fractionation**

In a 6-well plate,  $8.0 \times 10^4$  human BM-MSCs were seeded and cultured for 24 hours.

On the next day, control or anti-GATA2 siRNAs were transfected. After 48 hours, the

cells were collected and stained in a nucleic acid staining solution (NASS; 0.1 M

phosphate-citrate buffer, pH 4.8 with 0.9 % NaCl, 0.5 % bovine serum albumin, 0.02 %

saponin) supplemented with 1  $\mu$ M/mL 7-aminoactinomycin D (7-AAD, Imgenex,

SanDiego, CA) for 20 minutes. The cells were washed with PBS and were further

stained in the NASS supplemented with 10 ng/mL pyronin Y (Sigma Aldrich, St. Louis,

MO) for 5 minutes at 4°C. The cell cycle fractions were identified and quantified using

FACSArisII cell sorter and BD FACSDiva software (BD Biosciences, San Jose, CA).

Cells in G0 and G1 phases were determined as 7-AAD<sup>lo</sup>/pyronin Y<sup>lo</sup> and

7-AAD<sup>lo</sup>/pyronin Y<sup>int/hi</sup>, respectively.

## **Supplementary Figure Legends**

### **Supplementary Figure 1. Phenotyping of mouse BM-MSCs.**

(A) BM-MSCs were induced with iCre to delete the DNA binding domain of GATA2 using the Cre-loxP system. (B) Differentiation of GATA2<sup>fl</sup> BM-MSCs. Typical adipocytes contained oil drops that were stained with Oil Red O. (C) BM-MSCs from GATA2<sup>fl</sup> mice expressed cell-surface antigen markers for BM-MSCs. The data was a representative of 2 independent BM-MSC lines.

### **Supplementary Figure 2. Phenotyping of human BM-MSCs.**

(A) Differentiation of human BM-MSCs. Typical adipocytes contained oil drops that were stained with Oil Red O. (B) Human BM-MSCs expressed cell-surface antigens, characteristic of BM-MSCs. The data was a representative of 3 independent BM-MSC lines.

### **Supplementary Figure 3. Quantitative RT-PCR-based validation analysis for *GATA2* knockdown.**

The siRNA-mediated *GATA2* suppression was confirmed using quantitative RT-PCR.



The expression of *GATA2* relative to that of *GAPDH* was calculated (expressed as mean  $\pm$  SD,  $n = 3$ ). Asterisk,  $p < 0.05$ .

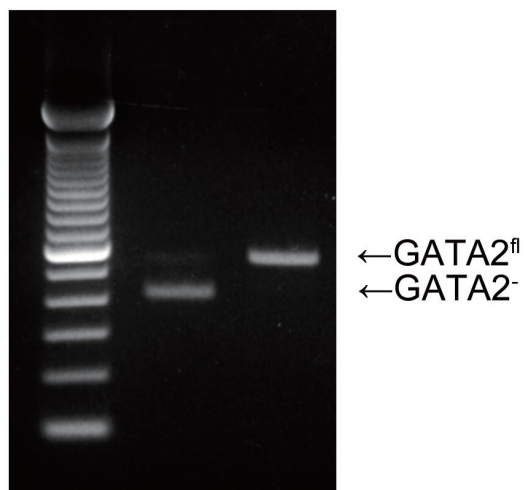
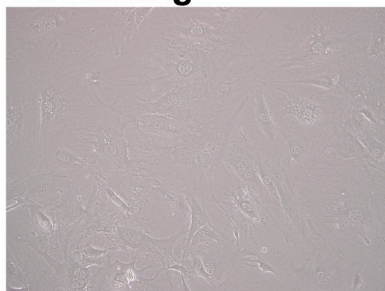
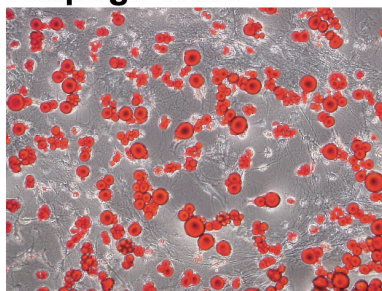
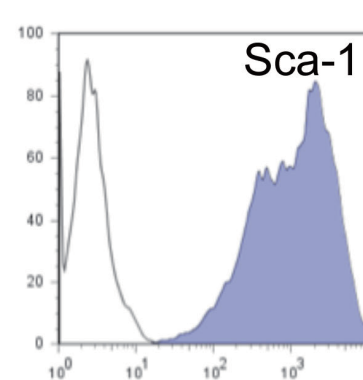
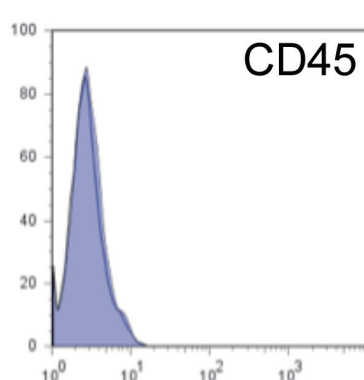
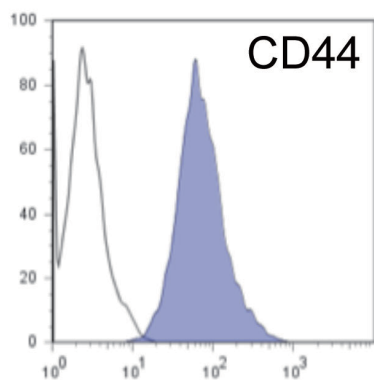
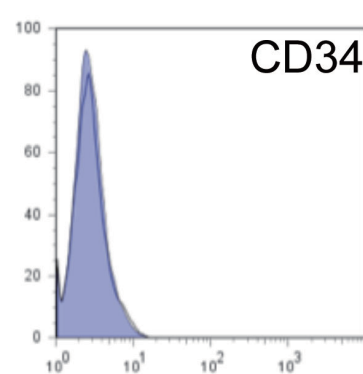
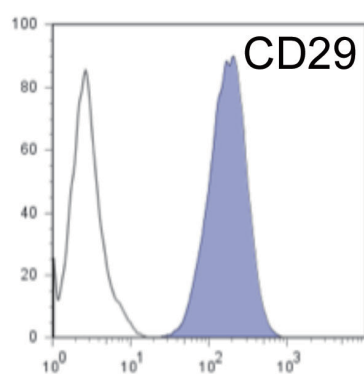
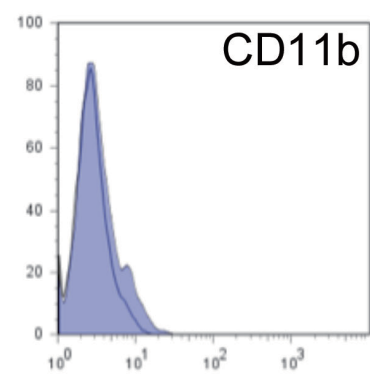
**Supplementary Figure 4. Increase of G1 phase cells with decreased GATA2 expression in BM-MSCs.**

(A) Representative cell-cycle profile in human BM-MSCs treated with control or anti-GATA-2 siRNA, based on 7-AAD and pyronin Y staining. (D) Percentage of cells in G0 and G1 stages of the cell cycle (expressed as mean  $\pm$  SD,  $n = 3$ ). The average percentages of G0 and G1 phase cells were significantly decreased and increased, respectively, by GATA2 knockdown ( $p < 0.05$ ).

**Supplementary Figure 5. Effects of various cytokines on adipocyte differentiation and GATA2 expression in BM-MSCs.**

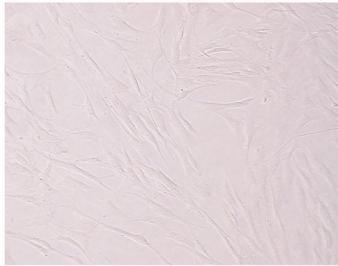
Human BM-MSCs were cultured with various cytokines (final concentration, 100 ng/mL) for 16 days, and samples were harvested on days 0, 4, 8, 12, and 16. Oil-drop formation and the expression of *GATA2* and *aP2* were analyzed using quantitative RT-PCR. (A) rhTNF $\alpha$ , (B) rhINF $\gamma$ , (C) rhIL-1 $\beta$ , (D) rhIL-6, (E) IL-17A, (F) rhTGF $\beta$ 1, (G)

rhBMP4. Data was expressed as mean  $\pm$  SD ( $n = 3$ ). Asterisk,  $p < 0.05$ .

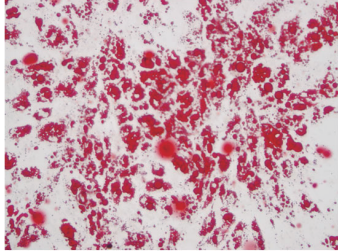
**A****B****Unstaining****Adipogenesis****C**

**A**

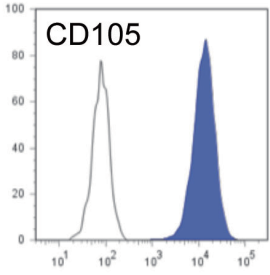
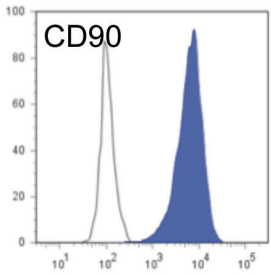
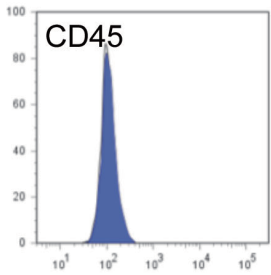
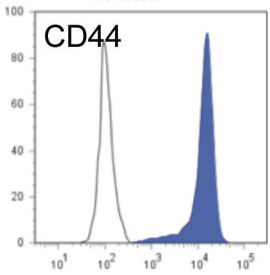
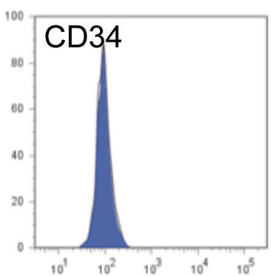
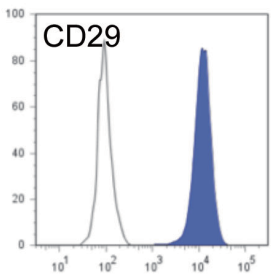
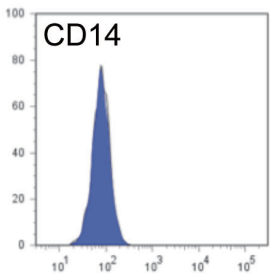
**Unstaining**

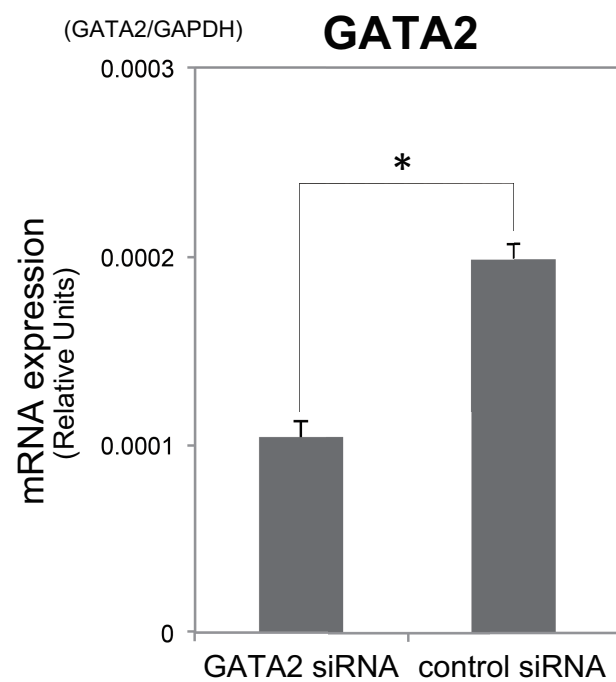


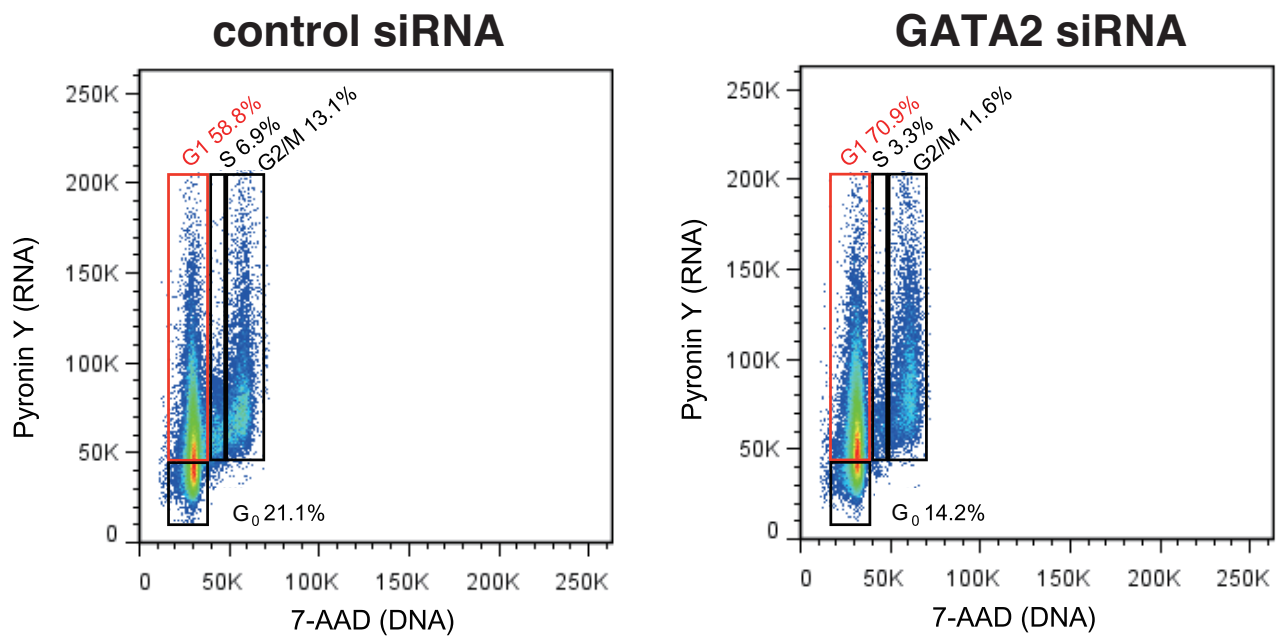
**Adipogenesis**



**B**





**A****B**

	G0 stage (%)	G1 stage (%)
control siRNA	23.0 ± 4.0	57.7 ± 3.5
GATA2 siRNA	15.1 ± 1.4	69.9 ± 1.7



