## **Supplemental Methods**

Genotyping of single nucleotide polymorphisms in the promoter of HO-1 gene

The promoter region (up to 1.4 kb) was analyzed by direct sequencing. Primer sequences have been published elsewhere (1). Briefly, PCR reactions carried out in Amplitaq Gold Buffer (Applied Biosystems, Courtaboeuf, France) with 2.5 mM MgCl<sub>2</sub>, 200 µM dNTPs (Roche Diagnotics, Mannheim, Germany), 10 pmol of each oligonucleotide primers, approximately 200 ng genomic DNA and 1 U AmpliTaq Gold Taq polymerase (Applied Biosystems, Couratboeuf, France). After initial denaturation for 10 min at 95°C, 35 cycles were carried out at 95°C for 30 s, 55°C for 30 s and 72°C for 1 min. Amplicons were then prepared for sequencing by incubation with Shrimp alkaline phosphatase and exonuclease I (EXOSAP®, Amersham Biosciences, Orsay, France). Nucleotide sequencing was carried out using CEQ Dye Terminator Cycle Sequencing (Beckman Coulter, Villepinte, France) and analyzed using a capillary sequencer CEQ 8000 (Beckman Coulter). All sequence changes were confirmed on both strands.

1. Ono K, Goto Y, Takagi S, Baba S, Tago N, Nonogi H, et al. A promoter variant of the heme oxygenase-1 gene may reduce the incidence of ischemic heart disease in Japanese. Atherosclerosis. 2004 Apr;173(2):315-9.

## **Supplemental table**

eTable 1: Allele and genotype frequencies of -1135 and -413 polymorphisms

-1135 A>G Genotype AA		Severe hemophilia A patients	
Genotype AA       18 (29.5%)       46 (26.4%)         Genotype GG       19 (31.1%)       57 (30.5%)         Genotype AG       24 (34.9%)       84 (44.9%)         Allele A       60 (49.2%)       176 (47.1%)         Allele G       62 (50.8%)       198 (52.9%)         -413 A>T         Genotype AA       18 (36.7%)       62 (35.2%)         Genotype TT       19 (38.8%)       82 (46.6%)         Genotype AT       12 (24.5%)       32 (18.2%)         Allele A       55 (56.1%)       206 (58.5%)         Allele T       43 (43.9%)       146 (41.5%)		Inhibitor-positive – N (%)	Inhibitor-negative – N (%)
Genotype GG       19 (31.1%)       57 (30.5%)         Genotype AG       24 (34.9%)       84 (44.9%)         Allele A       60 (49.2%)       176 (47.1%)         Allele G       62 (50.8%)       198 (52.9%)         -413 A>T       62 (35.2%)         Genotype AA       18 (36.7%)       62 (35.2%)         Genotype TT       19 (38.8%)       82 (46.6%)         Genotype AT       12 (24.5%)       32 (18.2%)         Allele A       55 (56.1%)       206 (58.5%)         Allele T       43 (43.9%)       146 (41.5%)	-1135 A>G		
Genotype AG  24 (34.9%)  84 (44.9%)  Allele A  60 (49.2%)  176 (47.1%)  Allele G  62 (50.8%)  198 (52.9%)  -413 A>T  Genotype AA  18 (36.7%)  62 (35.2%)  Genotype TT  19 (38.8%)  82 (46.6%)  Genotype AT  12 (24.5%)  32 (18.2%)  Allele A  55 (56.1%)  206 (58.5%)  Allele T  43 (43.9%)  146 (41.5%)	Genotype AA	18 (29.5%)	46 (26.4%)
Allele A 60 (49.2%) 176 (47.1%) Allele G 62 (50.8%) 198 (52.9%)  -413 A>T  Genotype AA 18 (36.7%) 62 (35.2%)  Genotype TT 19 (38.8%) 82 (46.6%)  Genotype AT 12 (24.5%) 32 (18.2%)  Allele A 55 (56.1%) 206 (58.5%)  Allele T 43 (43.9%) 146 (41.5%)	Genotype GG	19 (31.1%)	57 (30.5%)
Allele G       62 (50.8%)       198 (52.9%)         -413 A>T       62 (35.2%)         Genotype AA       18 (36.7%)       62 (35.2%)         Genotype TT       19 (38.8%)       82 (46.6%)         Genotype AT       12 (24.5%)       32 (18.2%)         Allele A       55 (56.1%)       206 (58.5%)         Allele T       43 (43.9%)       146 (41.5%)	Genotype AG	24 (34.9%)	84 (44.9%)
Allele G       62 (50.8%)       198 (52.9%)         -413 A>T       62 (35.2%)         Genotype AA       18 (36.7%)       62 (35.2%)         Genotype TT       19 (38.8%)       82 (46.6%)         Genotype AT       12 (24.5%)       32 (18.2%)         Allele A       55 (56.1%)       206 (58.5%)         Allele T       43 (43.9%)       146 (41.5%)			
-413 A>T  Genotype AA  18 (36.7%)  62 (35.2%)  Genotype TT  19 (38.8%)  82 (46.6%)  Genotype AT  12 (24.5%)  32 (18.2%)  Allele A  55 (56.1%)  206 (58.5%)  Allele T  43 (43.9%)  146 (41.5%)	Allele A	60 (49.2%)	176 (47.1%)
Genotype AA       18 (36.7%)       62 (35.2%)         Genotype TT       19 (38.8%)       82 (46.6%)         Genotype AT       12 (24.5%)       32 (18.2%)         Allele A       55 (56.1%)       206 (58.5%)         Allele T       43 (43.9%)       146 (41.5%)	Allele G	62 (50.8%)	198 (52.9%)
Genotype TT       19 (38.8%)       82 (46.6%)         Genotype AT       12 (24.5%)       32 (18.2%)         Allele A       55 (56.1%)       206 (58.5%)         Allele T       43 (43.9%)       146 (41.5%)	-413 A>T		
Genotype AT  12 (24.5%)  32 (18.2%)  Allele A  55 (56.1%)  206 (58.5%)  Allele T  43 (43.9%)  146 (41.5%)	Genotype AA	18 (36.7%)	62 (35.2%)
Allele A 55 (56.1%) 206 (58.5%) Allele T 43 (43.9%) 146 (41.5%)	Genotype TT	19 (38.8%)	82 (46.6%)
Allele T 43 (43.9%) 146 (41.5%)	Genotype AT	12 (24.5%)	32 (18.2%)
Allele T 43 (43.9%) 146 (41.5%)			
	Allele A	55 (56.1%)	206 (58.5%)
N of nationts 61 197	Allele T	43 (43.9%)	146 (41.5%)
1) of patients of 10/	N of patients	61	187