

Financial and other disclosures provided by the author using the ICMJE (www.icmje.org) Uniform Format for Disclosure of

Competing Interests are available with the full text of this paper at www.haematologica.org.

References

1. Parmar K, Mauch P, Vergilio JA, Sackstein R, Down JD. Distribution of hematopoietic stem cells in the bone marrow according to regional hypoxia. *Proc Natl Acad Sci USA*. 2007;104(13):5431-6.
2. Simsek T, Kocabas F, Zheng J, Deberardinis RJ, Mahmoud AI, Olson EN, et al. The distinct metabolic profile of hematopoietic stem cells reflects their location in a hypoxic niche. *Cell Stem Cell*. 2010;7(3):380-90.
3. Mendez-Ferrer S, Michurina TV, Ferraro F, Mazloom AR, MacArthur BD, Lira SA, et al. Mesenchymal and haematopoietic stem cells form a unique bone marrow niche. *Nature*. 2010;466(7308):829-34.
4. Mendez-Ferrer S, Lucas D, Battista M, Frenette PS. Haematopoietic stem cell release is regulated by circadian oscillations. *Nature*. 2008;452(7186):442-7.
5. Takubo K, Nagamatsu G, Kobayashi CI, Nakamura-Ishizu A, Kobayashi H, Ikeda E, et al. Regulation of glycolysis by Pdk functions as a metabolic checkpoint for cell cycle quiescence in hematopoietic stem cells. *Cell Stem Cell*. 2013;12(1):49-61.
6. Norddahl GL, Pronk CJ, Wahlestedt M, Sten G, Nygren JM, Ugale A, et al. Accumulating mitochondrial DNA mutations drive premature hematopoietic aging phenotypes distinct from physiological stem cell aging. *Cell Stem Cell*. 2011;8(5):499-510.
7. Romero-Moya D, Bueno C, Montes R, Navarro-Montero O, Iborra FJ, Lopez LC, et al. Cord blood-derived CD34+ hematopoietic cells with low mitochondrial mass are enriched in hematopoietic repopulating stem cell function. *Haematologica*. 2013;98(7):1022-9.
8. Notta F, Doulatov S, Laurenti E, Poeppl A, Jurisica I, Dick JE. Isolation of single human hematopoietic stem cells capable of long-term multilineage engraftment. *Science*. 2011;333(6039):218-21.
9. Nibley WE, Spangrude GJ. Primitive stem cells alone mediate rapid marrow recovery and multilineage engraftment after transplantation. *Bone Marrow Transplantation*. 1998;21(4):345-54.
10. Martinez C, Urbano-Ispizua A, Rozman C, Marin P, Rovira M, Sierra J, et al. Immune reconstitution following allogeneic peripheral blood progenitor cell transplantation: comparison of recipients of positive CD34+ selected grafts with recipients of unmanipulated grafts. *Exp Hematol*. 1999;27(3):561-8.
11. Isern J, Martín-Antonio B, Ghazanfari R, Martín AM, López JA, del Toro R, et al. Self-renewing human bone marrow mesospheres promote hematopoietic stem cell expansion. *Cell Rep*. 2013. pii: S2211-1247(13)00165-4.
12. Suda T, Takubo K, Semenza GL. Metabolic regulation of hematopoietic stem cells in the hypoxic niche. *Cell Stem Cell*. 2011;9(4):298-310.
13. Jang YY, Sharkis SJ. A low level of reactive oxygen species selects for primitive hematopoietic stem cells that may reside in the low-oxygenic niche. *Blood*. 2007;110(8):3056-63.
14. Ito K, Hirao A, Arai F, Matsuoka S, Takubo K, Hamaguchi I, et al. Regulation of oxidative stress by ATM is required for self-renewal of hematopoietic stem cells. *Nature*. 2004;431(7011):997-1002.
15. Tothova Z, Kollipara R, Huntly BJ, Lee BH, Castrillon DH, Cullen DE, et al. FoxOs are critical mediators of hematopoietic stem cell resistance to physiologic oxidative stress. *Cell*. 2007;128(2):325-39.
16. Oguro H, Iwama A, Morita Y, Kamijo T, van Lohuizen M, Nakauchi H. Differential impact of Ink4a and Arf on hematopoietic stem cells and their bone marrow microenvironment in Bmi1-deficient mice. *J Exp Med*. 2006;203(10):2247-53.
17. Liu J, Cao L, Chen J, Song S, Lee IH, Quijano C, et al. Bmi1 regulates mitochondrial function and the DNA damage response pathway. *Nature*. 2009;459(7245):387-92.
18. Moreno-Loshuertos R, Acin-Perez R, Fernandez-Silva P, Movilla N, Perez-Martos A, Rodriguez de Cordoba S, et al. Differences in reactive oxygen species production explain the phenotypes associated with common mouse mitochondrial DNA variants. *Nat Genet*. 2006;38(11):1261-8.
19. De Benedictis G, Rose G, Carrieri G, De Luca M, Falcone E, Passarino G, et al. Mitochondrial DNA inherited variants are associated with successful aging and longevity in humans. *Faseb J*. 1999;13(12):1532-6.
20. Zhang J, Asin-Cayuela J, Fish J, Michikawa Y, Bonafe M, Olivieri F, et al. Strikingly higher frequency in centenarians and twins of mtDNA mutation causing remodeling of replication origin in leukocytes. *Proc Natl Acad Sci USA*. 2003;100(3):1116-21.
21. Yao YG, Kajigaya S, Feng X, Samsel L, McCoy JP, Jr., Torelli G, et al. Accumulation of mtDNA variations in human single CD34(+) cells from maternally related individuals: Effects of aging and family genetic background. *Stem Cell Res*. 2013;10(3):361-70.
22. Ito K, Hirao A, Arai F, Takubo K, Matsuoka S, Miyamoto K, et al. Reactive oxygen species act through p38 MAPK to limit the lifespan of hematopoietic stem cells. *Nat Med*. 2006;12(4):446-51.
23. Chen C, Liu Y, Liu R, Ikenoue T, Guan KL, Liu Y, et al. TSC-mTOR maintains quiescence and function of hematopoietic stem cells by repressing mitochondrial biogenesis and reactive oxygen species. *J Exp Med*. 2008;205(10):2397-408.
24. Gurumurthy S, Xie SZ, Alagesan B, Kim J, Yusuf RZ, Saez B, et al. The Lkb1 metabolic sensor maintains haematopoietic stem cell survival. *Nature*. 2010;468(7324):659-63.
25. Gan B, Hu J, Jiang S, Liu Y, Sahin E, Zhuang L, et al. Lkb1 regulates quiescence and metabolic homeostasis of haematopoietic stem cells. *Nature*. 2010;468(7324):701-4.
26. Nakada D, Saunders TL, Morrison SJ. Lkb1 regulates cell cycle and energy metabolism in haematopoietic stem cells. *Nature*. 2010;468(7324):653-8.
27. Basu S, Broxmeyer HE, Hangoc G. Peroxisome proliferator-activated-gamma coactivator-1alpha-mediated mitochondrial biogenesis is important for hematopoietic recovery in response to stress. *Stem Cells Dev*. 2013;22(11):1678-92.
28. Forristal CE, Winkler IG, Nowlan B, Barbier V, Walkinshaw G, Levesque JP. Pharmacologic stabilization of HIF-1alpha increases hematopoietic stem cell quiescence in vivo and accelerates blood recovery after severe irradiation. *Blood*. 2013;121(5):759-69.
29. Frolova O, Samudio I, Benito JM, Jacamo R, Kornblau SM, Markovic A, et al. Regulation of HIF-1alpha signaling and chemoresistance in acute lymphocytic leukemia under hypoxic conditions of the bone marrow microenvironment. *Cancer Biol Ther*. 2012;13(10):858-70.
30. Skrtic M, Srikanthadevan S, Jhas B, Gebbia M, Wang X, Wang Z, et al. Inhibition of mitochondrial translation as a therapeutic strategy for human acute myeloid leukemia. *Cancer Cell*. 2011;20(5):674-88.

Circulating microparticles in children with sickle cell anemia: a heterogeneous procoagulant storm directed by hemolysis and fetal hemoglobin

Anna Falanga,¹ and Alice Trincheri^{1,2}

¹Division of Immunohematology and Transfusion Medicine, Hospital Papa Giovanni XXIII, Bergamo; ²Department of Internal Medicine, University of Pavia, Medical School, Pavia, Italy

E-mail: annafalanga@yahoo.com doi:10.3324/haematol.2013.085779

Chronic hemolytic anemias are made up of sickle cell anemia (SCA), beta (β)-thalassemia, paroxysmal nocturnal hemoglobinuria, autoimmune hemolytic anemia, and unstable hemoglobinopathies. They are associated with a high thrombotic risk. In SCA patients, a high

rate of both venous and arterial thrombosis (deep vein thrombosis, pulmonary embolism, stroke, pregnancy-related venous thromboembolism) has been reported.¹ Interestingly, these subjects commonly present with laboratory features of a subclinical hypercoagulable state,² charac-

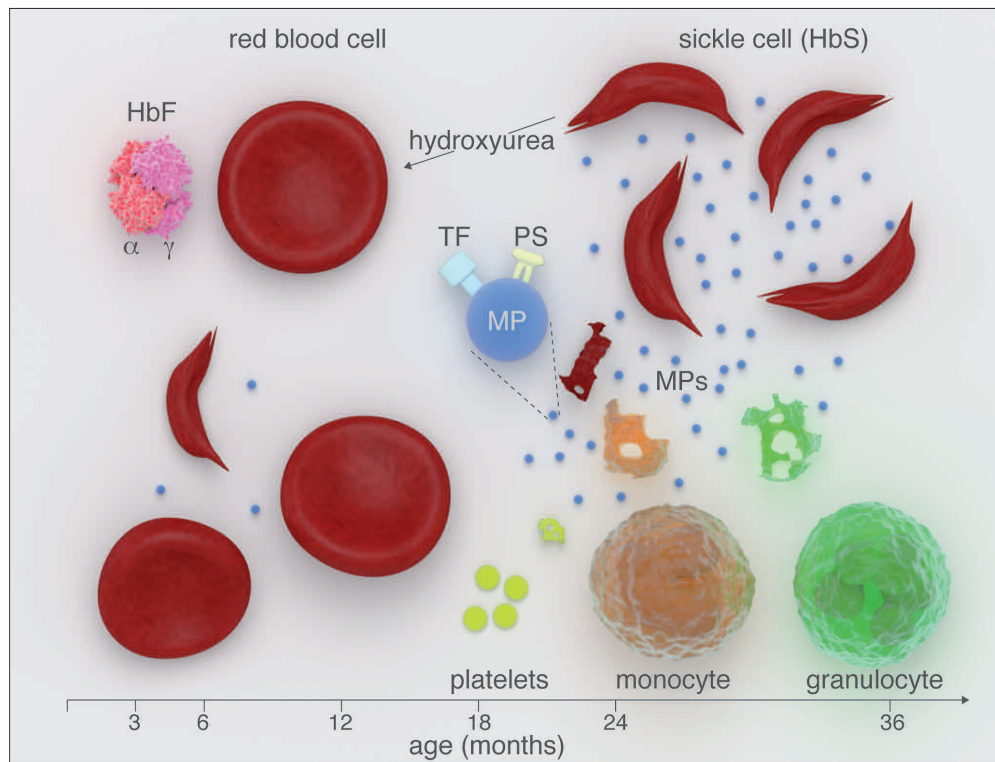


Figure 1. Mixed microparticles in SCA children act at the interface between hemolysis and clotting activation. Microparticles (MPs) of different cellular origin (e.g. red blood cells, platelets, monocytes, granulocytes) are released upon hemolysis, blood cell apoptosis, and activation. Hemolysis increases when the physiological switch from fetal (HbF) to adult hemoglobin occurs and sickle cells (characterized by HbS) are produced. *Vice versa*, hemolysis decreases when hydroxyurea therapy is started, with subsequent upregulation of HbF ($\alpha_2\gamma_2$) levels. On their surface MPs expose procoagulant phospholipids (i.e. phosphatidylserine, PS) and proteins (i.e. Tissue Factor, TF), and activate other blood cells (e.g. platelets, monocytes) and hemostasis. HbF levels inversely correlate to circulating MPs and regulate specific MP patterns, particularly those platelet-derived.

terized by increased plasma levels of markers of thrombin generation, e.g. D-Dimer, thrombin-antithrombin complex (TAT), prothrombin Fragment 1+2 (F1+2), during the non-crisis 'steady state' and, in particular, during acute pain episodes. In fact, hemolysis is a known procoagulant condition³ due to the release of cell-free plasma hemoglobin and the depletion of nitric oxide.⁴ Additional biological mechanisms of hemolysis-associated hypercoagulation include: red blood cell membrane abnormalities leading to exposure of anionic procoagulant phospholipids (i.e. phosphatidylserine), endothelial dysfunctions with overexpression of cell adhesion molecules, platelet activation, thrombocytosis following functional hyposplenism or surgical splenectomy.^{2,4,6}

Importantly in this scenario is the occurrence in SCA patients of increased levels of circulating microparticles (MPs).⁷ MPs are vesicles of less than 1 μm resulting from the shedding of activated or apoptotic blood and vascular cell membranes. Among their various functions, MPs exert procoagulant actions through the expression on their surface of procoagulant phospholipids (i.e. phosphatidylserine) and proteins (i.e. Tissue Factor).⁸ Elevated circulating MPs are found in different clinical conditions at high thrombotic risk, e.g. diabetes mellitus, atherosclerosis, acute coronary syndrome and myocardial infarction, sepsis, antiphospholipid syndrome, malignancy.⁹⁻¹² Specifically, in SCA, MPs of erythrocyte origin produced during hemolysis carry negative niches that activate the intrinsic phase of blood coagulation (tenase and prothrombinase) leading to thrombin generation.¹³ A significant correlation between the total number of MPs and the levels of markers of hypercoagulability (i.e. D-dimer, TAT, and F1+2) has been repeatedly demonstrated in SCA patients.^{7,13-15} In this condition, MPs

likely represent the interface between hemolysis and blood clotting activation. However, as shown for the first time by the study of Nébor *et al.* published in this Journal,¹⁶ in SCA children there is a variety of MPs which originate not only from erythrocyte, but from virtually all blood cells, mainly platelets. In contrast to the situation for adult patients,¹³⁻¹⁵ only limited data are available regarding MP characterization in SCA children.⁵ Nébor *et al.* found that, although platelet-derived and erythrocyte-derived MPs were the most common type in this condition, all other cell origins, e.g. monocyte-, granulocyte-, and endothelial cell-derived MPs, were represented. Interestingly, the age-related reduction in HbF levels during childhood was associated with an increase in MP levels, particularly those from platelets and monocytes, and to a lesser extent those from erythrocytes. While confirming the already known inverse relation between HbF concentration and MPs formation⁵ and thrombin generation,¹³ these data show for the first time the specific cellular patterns involved in the process. In the same way, in this population the reactivation of fetal hemoglobin (HbF) synthesis (which impairs HbS polymerization) induced by hydroxyurea, the current standard therapy option in SCA, correlated with the reduction in plasma levels of MPs, particularly those of platelet and erythrocyte origin. Attempts to standardize the methodology for the isolation, analysis and count of MPs have been shown to have limitations and these tests can be influenced by many different factors, from blood collection up to gate analysis.¹⁷ However, the data published here open up new perspectives on how all blood cellular compartments are involved in the clotting activation associated to SCA and, possibly, to all hemolytic anemias. In these circumstances, different subtypes of MPs act as messengers between hemolysis and

the hemostatic system activation. This also expands our vision of the possible mechanism(s) involved in the hemolytic crisis brought on by other comorbid conditions, such as sepsis. Along the same lines, there is evidence that the HbF levels, an important regulatory mechanism of SCA severity and hemolysis, govern MP concentration by acting on specific MP subtypes.

We can imagine that, in SCA children, a storm of various (mainly platelet-derived) procoagulant MPs takes place with chronic hemolysis and is driven by HbF levels (Figure 1).

Anna Falanga is Head of the Division of Immunohematology/Transfusion Medicine and the Hemostasis and Thrombosis Center at the Department of Oncology-Hematology of Hospital Papa Giovanni XXIII, in Bergamo, Italy. Her main clinical and research interest is pathogenesis and management of thrombotic and hemorrhagic disorders associated to hematologic malignancies. Alice Trincheri is in her 5th year at the Specialty School at the University of Pavia Medical School, Italy, and is a fellow of the Hemostasis and Thrombosis Center of Hospital Papa Giovanni XXIII, Bergamo, Italy.

Financial and other disclosures provided by the author using the ICMJE (www.icmje.org) Uniform Format for Disclosure of Competing Interests are available with the full text of this paper at www.haematologica.org.

References

- Ataga KI, Cappellini MD, Rachmilewitz EA. Beta-thalassaemia and sickle cell anaemia as paradigms of hypercoagulability. *Br J Haematol.* 2007;139(1):3-13.
- Ataga KI, Brittain JE, Desai P, May R, Jones S, Delaney J, et al. Association of coagulation activation with clinical complications in sickle cell disease. *PLoS one.* 2012;7(1):e29786.
- Cappellini MD. Coagulation in the pathophysiology of hemolytic anemias. *Hematology Am Soc Hematol Educ Program.* 2007;74-8.
- Villagra J, Shiva S, Hunter LA, Machado RF, Gladwin MT, Kato GJ. Platelet activation in patients with sickle disease, hemolysis-associated pulmonary hypertension, and nitric oxide scavenging by cell-free hemoglobin. *Blood.* 2007;110(6):2166-72.
- Setty BN, Kulkarni S, Rao A K, Stuart MJ. Fetal hemoglobin in sickle cell disease: relationship to erythrocyte phosphatidylserine exposure and coagulation activation. *Blood.* 2000;96(3):1119-24.
- Setty BNY, Betal SG, Zhang J, Stuart MJ. Heme induces endothelial tissue factor expression: potential role in hemostatic activation in patients with hemolytic anemia. *J Thromb Haemost.* 2003;6(12):2202-9.
- Shet AS, Aras O, Gupta K, Hass MJ, Rausch DJ, Saba N, et al. Sickle blood contains tissue factor-positive microparticles derived from endothelial cells and monocytes. *Blood.* 2003;102(7):2678-83.
- Lacroix R, Dignat-George F. Microparticles as a circulating source of procoagulant and fibrinolytic activities in the circulation. *Thromb Res.* 2012;129:S27-9.
- Nieuwland R, Berckmans RJ, McGregor S, Böing AN, Romijn FP, Westendorp RG, et al. Cellular origin and procoagulant properties of microparticles in meningococcal sepsis. *Blood.* 2000;95(3):930-5.
- Zwicker JI, Liebman HA, Neuberger D, Lacroix R, Bauer KA, Furie BC, et al. Tumor-Derived Tissue Factor – Bearing Microparticles Are Associated With Venous Thromboembolic Events in Malignancy Tumor-Derived Tissue Factor – Bearing Microparticles Are Associated With Venous Thromboembolic Events in Malignancy. *Clin Cancer Res.* 2009;15(22):6830-40.
- Trappenburg MC, Van Schilfgaarde M, Marchetti M, Spronk HM, Ten Cate H, Leyte A, et al. Elevated procoagulant microparticles expressing endothelial and platelet markers in essential thrombocythemia. *Haematologica.* 2009;94(7):911-8.
- Falanga A, Tartari CJ, Marchetti M. Microparticles in tumor progression. *Thromb Res.* 2012;129:S132-6.
- Gerotziakas GT, Van Dreden P, Chaari M, Galea V, Khaterchi A, Lionnet F, et al. The acceleration of the propagation phase of thrombin generation in patients with steady-state sickle cell disease is associated with circulating erythrocyte-derived microparticles. *Thromb Haemostasis.* 2012;107(6):1044-52.
- Van Beers EJ, Schaap MCL, Berckmans RJ, Nieuwland R, Sturk A, Van Doornaal FF, et al. Circulating erythrocyte-derived microparticles are associated with coagulation activation in sickle cell disease. *Haematologica.* 2009;94(11):1513-9.
- Westerman M, Pizzey A, Hirschman J, Cerino M, Weil-Weiner Y, Ramotar P, et al. Microvesicles in haemoglobinopathies offer insights into mechanisms of hypercoagulability, haemolysis and the effects of therapy. *Br J Haematol.* 2008;142(1):126-35.
- Nébor D, Romana M, Santiago R, Vachery N, Picot J, Broquere C, et al. Fetal hemoglobin and hydroxycarbamide modulate both plasma concentration and cellular origin of circulating microparticles in sickle cell anemia children. *Haematologica.* 2013;98(6):862-7.
- Lacroix R, Robert S, Poncelet P, Kasthuri RS, Key NS, Dignat-George F. Standardization of platelet-derived microparticle enumeration by flow cytometry with calibrated beads: results of the International Society on Thrombosis and Haemostasis SSC Collaborative workshop. *J Thromb Haemost.* 2010;8(11):2571-4.

Are ongoing trials on hematologic malignancies still excluding older subjects?

Antonio Cherubini,^{1,2} Francesca Pierri,³ Beatrice Gasperini,² Elisa Zengarini,² Annarita Cerenzia,² Elisabetta Bonifacio,⁴ Flavio Falcinelli,⁴ and Fabrizia Lattanzio⁵

¹Geriatrics, Italian National Research Center on Aging (IRCCS-INRCA), Ancona; ²Institute of Gerontology and Geriatrics, Department of Clinical and Experimental Medicine, University of Perugia Medical School; ³Department of Economy, Finance and Statistics, University of Perugia; ⁴Institute of Hematology, Department of Hematology and Clinical Oncology, University of Perugia, Ospedale S. Maria della Misericordia, Perugia; ⁵Scientific Direction, Italian National Research Center on Aging (INRCA-IRCCS), Ancona, Italy

E-mail: a.cherubini@inrca.it doi:10.3324/haematol.2013.087601

Hematologic malignancies are diseases that mainly affect older subjects. Multiple myeloma,¹ myelodysplastic syndromes² and chronic myeloid leukemia³ are common in advanced age. Nevertheless, there is evidence that older patients with hematologic malignancies have often been excluded from clinical trials (CTs).^{4,5} Their exclusion prevents clinicians from obtaining information concerning the efficacy and safety of treatments in older patients and might represent an important barrier to the treatment of these patients.⁶ Published literature reflects tri-

als performed some years before their publication. It is not known whether older individuals are gradually being included in more trials as a consequence of the aging of the population and of the recommendation provided by Regulatory Agencies, e.g. FDA and ICH, to include older individuals in CTs.^{7,8} The aims of this study were to assess the presence and the extent of underrepresentation of older individuals in ongoing CTs on hematologic malignancies registered in an online open-access CT registry maintained by the World Health Organization (WHO), and to evaluate