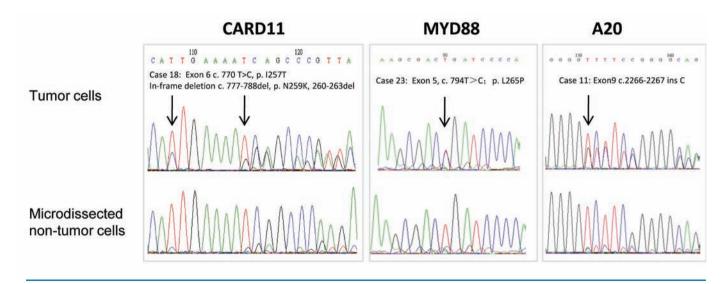
BCR and TLR signaling pathways are recurrently targeted by genetic changes in splenic marginal zone lymphomas

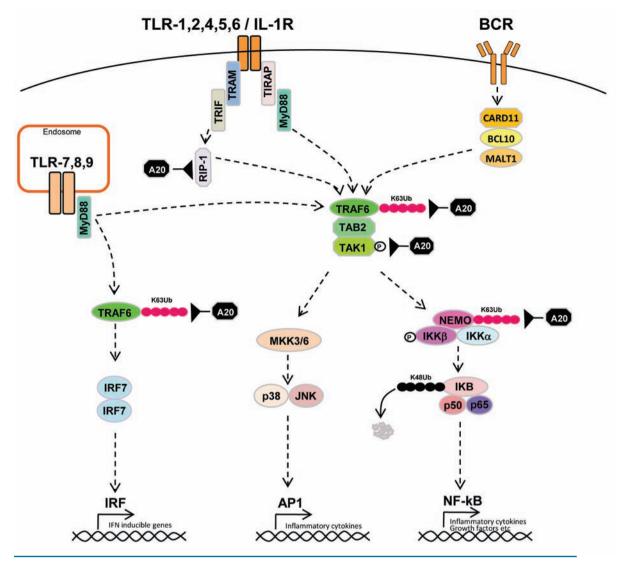
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Online Supplementary Figure S1. Examples of CARD11, MYD88 and A20 mutation seen in SMZL.



Online Supplementary Figure S2. CARD11, MYD88 and A20 in BCR and TLR signaling pathways. CARD11 links the BCR signaling to the canonical NF-kB activation pathway. MYD88 is a universal adaptor and couples TLR/IL-R signaling to the NF-kB, AP1 and IRF activation pathway. A20, an ubiquitinmodification enzyme, inactivates RIP-1, TRAF6, TAK1 and NEMO, important signaling molecules mediating NF-kB, AP1 and IRF activation.

Genes	Exon	Primer name	Sequence (5'-3')	Amplicon size (bp)	PCR annealing temperature (°C)	PCR condition			
	E5-1	Forward	GTGCCCCTCTCCACAGT	200	62				
	E3-1	Reverse	AGTACCGCTCCTGGAAGGTT]	62				
	E5-2	Forward	GAAGAAGCAGATGACGCTGA	235	62				
	E3-2	Reverse	GTCACCCTGGCGGAGTAG	233	02				
	E6	Forward	CACCCTTGGGGTATTTCAGA	210	59	7			
CARD11	Lo	Reverse	CAGGCCCTCACCTGGATG						
	E7	Forward	CCTGACCCTCTGAAACCTCCT	204	62				
	L,	Reverse	GCGATCCCCACTCCCAC	204	02				
	E8	Forward	TCGATGCGCATATTGATTTC	181	62				
		Reverse	CTGCAGGTGGTGCCTGTA	101					
	E9	Forward	CCCAAAGCAGCCTTCGTC	234	62				
	E9	Reverse	CCTGGTCCAGGTTGTTGCTGTCC	254					
	F1.1	Forward	CTCGGGGCTCCAGATTGTA	227	58				
	E1-1	Reverse	GCCGGATCTCCAAGTACTCA	327					
	F1.0	Forward	GCTGCTCTCAACATGCGAGT	2.5					
	E1-2	Reverse	GGAAAGTCAGCCTCCTCACC	317	62	PCR was carried out in a 10µl			
		Forward	CTGGATCCTGACTGTGGGTAA			reaction mixture with 5-10ng			
	E2	Reverse	GCTTCAAACACCCATGCTCT	- 281	62	template DNA and AmpliTaq			
MYD88		Forward	TCTGACCACCACCCTTGTG	2/2/		Gold 360 (Applied			
	E3	Reverse	CAGGGCAGGGCTTCATGC	- 264	62	Biosystem) master mix plus			
		Forward	GGCCCTTCCTGAAGCTATTC	<u> </u>	62	GC-enhancer according to the			
	E4	Reverse	TGGTACTGCATCCACAGTCC	270		manufacturer's instructions. The PCR conditions were			
		Forward	GTTGAAGACTGGGCTTGTCC		59	95°C for 10 min to activate the enzyme, followed by 40			
	E5	Reverse	AGGAGGCAGGGCAGAAGTA	- 292					
7 10 10 10 10 10 10 10 10 10 10 10 10 10	-	Forward	ATGAAGTGAGTGAAGGGTGGG			cycles of denaturation at			
CD79A	E5	Reverse	AGAATGTCCCAGGGAAGTGAG	326	58	95°C for 20 sec, annealing a			
		Forward	TAGGTGGCTGTCTGGTCAATG		7.22	58-62°C (depending on the			
	E5	Reverse	TGTTCTTGCAGAATGCACCTC	306	58	primer set) for 20 sec, and extension at 72°C for 30-45			
CD79B		Forward	CTGGAGACAAATGGCAGCTC	2.52		sec (depending on the			
	E6	Reverse	CACCTACGAGGTAAGGAGAGGG	362	58	amplicon size).			
	F2.4	Forward	CTGCAGGCAGCTATAGAGGAG						
	E2-1	Reverse	CGAAACTGAGGACAAAACTGG	272	58				
	F2.2	Forward	GCAATATGCGGAAAGCTGTG	200		and the state of t			
	E2-2	Reverse	GCTATCACCCAGGCAAAAGA	300	58				
	F2	Forward	TTGCTGGGTCTTACATGCAG	271	50	***			
	E3	Reverse	TTAGGGGGAAAAACCTACCC	271	58				
	E4	Forward	GGGAGTACAGGATACATTCAAGC	251	50				
	E4	Reverse	AAGGCATAAGGCTGAAAGCA	251	58				
A20	E5	Forward	ACCTAAGGGCCTCATTTTCC	275	58				
	E3	Reverse	GCAAAAAGGAAAACCCTGATG	2/3	36				
	E6	Forward	TGAGATCTACTTACCTATGGCCTT G	315	58				
		Reverse	TCAGGTGGCTGAGGTTAAAGA			-			
	E7-1	Forward	ACAGGCCTGCATTTCAGTG	282	58				
		Reverse	GGAAGGTTCCATGGGATTC						
	E7-2	Forward Reverse	GCAGGAAAACAGCGAGCA CCAAGGGCTCATAGGCTTCT	272	58				
	E7-3	Forward	ACTCCCAAAGCTGAACTCCA	304	58	7			

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		Reverse	GGGATCCAAGTGCCTTGT			
	E7-4	Forward	ACTGCCATGAAGTGCAGGAG	279	58	
		Reverse	ATCTGACTTGGAACGCTGGT			
	E7-5	Forward	TGCAGTACTTGCTTCAAAAGGA	313	58	
	Li-3	Reverse	CCACTTCACTCACGTTTGTTTT	313		
	E8	Forward	GGGGTGACCCCTATGTGGTACT	293	58	
	Lo	Reverse	CCAGTTGCTCTTCTGTCCTTTT	2,53	56	
	E9-1	Forward	GTGCTCTCCCTAAGAAATGTGAG	205	58	
	L9-1	Reverse	CTGGTTGGGATGCTGACACT	203	36	
	E9-2	Forward	CTCTGCATGGAGTGTCAGCAT	257	58	
		Reverse	GGGTTCAGAGGATAGCACCA	231		
	E2	Forward	GGCATTGACAGACTAGAGCTTC	315	58	
	LZ	Reverse	GACTGGCATCACAGTCTGC	313	36	
	E3	Forward	GGGAGAAGCAGCACACTG	272	58	
	E3	Reverse	GAAGGGAGTTCACTGTGAAGG	272	30	
	E4	Forward	CCTGTCTGAGAACCCTTTTG	200	50	
	E4	Reverse	GACCTCAACCCTCTTTCTTC	280	58	
	D5	Forward	GCTTTGTGATCTCATGTGAGATG	227	50	
	E5	Reverse	GGGATCTGAACAGGTTCTGTG	327	58	
	E(Forward	GAGGAAGCAGAGGGAGAATG	246	50	
	E6	Reverse	GAGCAGGAAGGTGGGAAG	346	58	
	F-7	Forward	CCCACCATCCTCTAGCTCAG	252	50	
	E7	Reverse	CCAGAAATCAGTGCTGCAC	252	58	
	E8 -	Forward	CAACTCATGCGATAGTGAGC	222	50	
		Reverse	GAACCTTCTACCACTGGCAC	332	58	
	E9	Forward	GGTCTTAGAGGAGCACCGAG	226	58	
		Reverse	AGCCTTTGTGCTGCTGGT	236		
	E10	Forward	TAGCCACTATTCACACACACC	265	58	
ABIN-1		Reverse	CCCCAAGGTTCAAAGCTG	265		
		Forward	CACATCCTGCAAGTGCTCTAC	200	58	
	E11	Reverse	GCTTGTTTGCTCCACAGAAC	290		
	EIA	Forward	CCAGGCAGGACAGAGAATC	202	5 0	
	E12	Reverse	CCTGAGTCACTCCCAGTGTG	302	58	
	E12	Forward	GAAGGAGTTCCCTGAGGATG	272	5 0	
	E13	Reverse	GAAAGCTCCAGCTCCCACAG	273	58	
	E14	Forward	AGGAGGCATGGGAGTCTG		58	
		Reverse	GAGGACAGGCCAGTTGC	251		
	E15	Forward	CCAGAGGGAAGCATCAG		58	
		Reverse	CACACACTGTGCATCCATC	259		
	E16	Forward	AGAGGGTGATGAGATGGGT		58	
		Reverse	CTCTCTGGAAGGTGTCTGG	328		
	E17 -	Forward	TTGTCCTGGCACAGTAGGTG		58	
		Reverse	AGGCAATGCTGGCAGATAAG	238		
		Forward	ACTGTTCCTGCACTGCATTC	i i		
	E18	Reverse	TCAGGGACTGGTGTACAAGC	272	58	

Online Supplementary Table S2. Summary of mutations found in SMZL.

Case reference	CARD11 mutation	MYD88 mutation	A20 mutation		
C08	n/a	n/a	Exon7 c.1246-1250 del AACAA		
C09	No	No	Exon4 c.553G>T, p.G185X (somatic origin confirmed)		
C10	No	No	Exon9 c.2209C>T, p. Q737X		
C11	No	No	Exon9 c.2266-2267insC (somatic origin confirmed)		
C17	Exon6 c.746A>C, p.Q249P	No	No		
C18	Exon6,c. 770 T>C, p. I257T;In-frame deletion c. Δ12bp(777-788del), p. N259K,260-263del; (both mutations occurred on the same allele and somatic origin confirmed)	No	No		
C19	Exon5 c.572A>G, p. N191S	Exon5 c.794T>C, p.L265P	No		
C20	No	Exon5 c.794T>C, p.L265P (somatic origin confirmed)	No		
C21	No	Exon5 c.794T>C, p.L265P	No		
C22	No	Exon5 c.794T>C, p.L265P	No		
C23	n/a	Exon5 c.794T>C, p.L265P (somatic origin confirmed)	n/a		
C24	No	Exon5 c.794T>C, p.L265P	No		
C28 No		No	Exon3 c.400-401 del GA (somatic origin confirmed)		
C29	n/a	n/a	Exon6 c.872 del T		

n/a: not applicable.

Online Supplementary Table S3. Univariate analysis for prognosis by the Kaplan-Meier method.

Factor		os				EFS		
ractor		2 yrs	5 yrs	P =	2 yrs	5 yrs	P =	
Age	<60 (n= 7) >=60 (n=17)	1 0.941	1 0.739	0.210	1 0.729	1 0.574	0.117	
Stage	<iv (n="24)</td" iv=""><td>1 0.958</td><td>1 0.834</td><td>0.675</td><td>1 0.781</td><td>1 0.558</td><td>0.517</td></iv>	1 0.958	1 0.834	0.675	1 0.781	1 0.558	0.517	
Arcaini prognostic index	Low (n=4) Moderate (n=16) High (n=7)	1 1 0.857	1 0.923 0.643	0.204	0.667 0.909 0.686	0.333 0.701 0.686	0.510	
Lymphocytosis	Yes (n=18) No (n= 10)	0.944 1	0.850 0.889	0.767	0.791 0.875	0.562 0.750	0.800	
Villous lymphocytes	Yes (n=15) No (n= 9)	1	1 0.800	0.121	0.917 0.833	0.625 0.833	0.641	
7q deletion	Yes (n=10) No (n=19)	1 0.947	0.857 0.806	0.689	0.714 0.821	0.714 0.559	0.200	
TP53 status	Abnormal (n=5) Normal (n=24)	1 0.958	0.800 0.834	0.954	0.800 0.771	0.800 0.551	0.759	
A20 mutation	Yes (n=1) No (n=24)	1 0.958	1 0.846	0.686	1 0.797	1 0.607	0.541	
CARD11 mutation	Yes (n=3) No (n=18)	1 0.944	1 0.872	0.571	1 0.807	1 0.588	0.471	
MYD88 mutation	Yes (n=5) No (n=20)	1 0.950	0.800 0.871	0.713	0.8 0.793	0.533 0.634	0.514	
A20 or CARD11 or MYD88 mutation	Yes (n=7) No (n=14)	1 0.929	1 0.825	0.281	1 0.734	0.667 0.554	0.675	

2 yrs: 2-year cumulative probability of survival 5 yrs: 5-year cumulative probability of survival

P value by log rank (Mantel-Cox)