

using the HbA₂ value of 3.4% as cut off.⁸

Using the value of 30 µg/L for serum ferritin as cut off, 861 samples showed iron deficiency (ID) (group A) and 271 were without ID (group B) (Figure 1). The mean HbA₂ value was 2.8%±0.79 in group A and 3.50%±1.23 in group B, with a significant difference ($P=0.00001$) among between the two groups.

The distribution of the 253 samples with molecular analysis between groups A and B showed that 170 samples were part of group A and 83 of group B (Figure 1).

From 170 samples of group A, 21 resulted positive for a β -globin gene mutation with a mean of HbA₂ value of 4.90%±1.29. From 83 samples of group B, 29 resulted positive for a β -mutation (mean of HbA₂ value: 5.37%±0.87).

The comparison between the HbA₂ mean value of β -thalassemia carriers with (group A) or without (group B) ID, using the value of 30 µg/L as serum ferritin cut off, does not show a significant difference ($P=0.060$) and in both groups HbA₂ levels are more than 3.4%. Reduction of HbA₂ has been reported to be linked to the severity of anemia⁹ so that it is possible that the value of 30 µg/L for serum ferritin defines an ID not sufficiently severe or not sufficiently prolonged to significantly reduce the level of HbA₂.

The ROC analysis, performed with samples of group A, at the 3.4% HbA₂ cut-off value, showed sensitivity and specificity of 74.19% and 95.8%, respectively. The false negative samples were 8 of 30 (26,6%): 3 presented Hb β variants, co-eluting with HbA, (Hb Valletta, Hb Ern and Hb City of Hope) that do not require prenatal diagnosis, one showed an undefined single nucleotide polymorphism (SNP) in the β -globin gene of ambiguous diagnostic significance,⁸ 2 samples presented, respectively, the α -globin gene triplication ($\alpha\alpha\alpha$ ^{amb3,7}), and the β -globin gene promoter mutation, -101 (HBB c.-151C→T), and finally, 2 samples showed co-heredity of β^+ and δ^+ mutation.

The samples with $\alpha\alpha\alpha$ ^{amb3,7} and the -101 beta mutation showed, respectively, HbA₂ values of 3.0% and 3.2% with a lower value of Hb (<12 g/dL) and MCV (<75 fL). The contemporary analysis of hematologic and hemoglobin data enable us to identify these subjects as samples that must be submitted to molecular analysis if their partners are carriers of β thalassemia.⁶ While, in other cases, it is recommended to remeasure HbA₂ after ID treatment.

In the 2 samples showing co-heredity of β^+ and δ^+ mutation, the large reduction is principally associated with the presence of delta mutation rather than serum ferritin value.

Our results show that the presence of iron deficiency did not preclude the detection of classical β carrier in our population. There could be some problems in the presence of silent β mutation or α gene triplication with ID, because HbA₂ shows almost normal levels. However, the reduction in total Hb and MCV, and possible persistently low MCV and MCH after iron supplementation, should suggest greater attention is needed and molecular analysis exploring both the α and the β genes should be carried out, especially if the subject is a partner of a classical β thalassemia carrier.

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References

1. Giambona A, Passarello C, Renda D, Maggio A. The significance of the hemoglobin A(2) value in screening for hemoglobinopathies. Clin Biochem. 2009;42(18):1786-96.
2. Giambona A, Lo Gioco P, Marino M, Abate I, Di Marzo R, Renda M, et al. The great heterogeneity of thalassemia molecular defects in Sicily. Hum Genet. 1995;95(5):526-30.
3. Steinberg MH, Adams JG. Hemoglobin A2: Origin, evolution and aftermath. Blood. 1991;78(9):2165-77.
4. Harthoorn-Lasthuizen EJ, Lindemans J, Langenhuijsen MM. Influence of iron deficiency anaemia on haemoglobin A2 levels: possible consequences for beta-thalassaemia screening. Scand J Clin Lab Invest. 1999;59(1):65-70.
5. Madan N, Sikka M, Sharma S, Rusia U. Phenotypic expression of hemoglobin A2 in beta-thalassemia trait with iron deficiency. Ann Hematol. 1998;77(3):93-6.
6. Giambona A, Passarello C, Ruggeri G, Renda D, Teresi P, Anzà M, et al. Analysis of δ -globin gene alleles in the Sicilian population: identification of five new mutations. Haematologica. 2006;91(12):1684-7.
7. Mast AE, Blinderb MA, Gronowsky AM, Chumley C, Scott MG. Clinical utility of the soluble transferrin receptor and comparison with serum ferritin in several populations. Clin Chem. 1998;44(1):45-51.
8. Giambona A, Passarello C, Vinciguerra M, Li Muli R, Teresi P, Anzà M, et al. Significance of borderline hemoglobin A2 values in an Italian population with a high prevalence of β -thalassemia. Haematologica. 2008;93(9):1380-4.
9. Alperin JB, Dow BS, Petteway BS. Hemoglobin A2 levels in health and various hematologic disorders. Am J Clin Pathol. 1977;67(3):219-26.

Efficacy of combining dasatinib and FLAG-IDA for patients with chronic myeloid leukemia in blastic transformation

The prognosis of patients with chronic myeloid leukemia (CML) has improved considerably over the last ten years with the introduction of ABL tyrosine kinase inhibitors (TKI) into clinical practice. TKIs induce complete cytogenetic remissions (CCyR) in 10-45% of patients who are treated with these drugs in advanced phase with minimal toxicity; unfortunately, these remissions are typically short lasting.^{1,2} Dasatinib alone induces CCyR in 20-40% of patients.^{2,3} However, the majority relapse within one year and the median survival is eight months.² Conventional chemotherapy regimens, such as FLAG-IDA, can induce CCyR in 30-40% of patients who have progressed to blastic phase, but again most patients relapse within six months and survival is poor.⁴ A logical approach might be to combine both strategies in order to improve the outcome. The proposed schedule of combination TKI and chemotherapy is supported by two clinical

Table 1. Patients' characteristics and responses.

Patient ID	Patients' characteristics					Response to the #1dasatinib-FLAG-IDA			Response to #2dasatinib-FLAG-IDA		
	Age/sex	% blasts (BM)	Cytogenetics	BCR-ABL/ABL (%)	Phenotype	% blasts (BM)	Cytogenetics	BCR-ABL/ABL (%)	% blasts (BM)	Cytogenetics	BCR-ABL/ABL (%)
1	65/F	45	46,XX,(9;22)(q34;q11)[1]/46,XY[9]	95.6	lymphoid	2	46,XX[20]	0.037	0	46 XX[30]	0.08
2*	44/F	67	G-banding failed	139.7	biphenotypic	1	46,XX[30]	0.015	2	46 XX[30]	0.70
3#	41/M	33	G-banding failed	24.8	myeloid	3	46,XY[30]	0.019	1	46 XY[30]	0.09
4	55/M	73	46,XY,(9;22)(q34;q11)[28]/46,XY[2]	115.7	myeloid	5	46,XY,(9;22)(q34;q11)[1]/46,XY[29]	1.2	1	46 XY[30]	0.09

*Patient 2 harbored the ABL kinase domain mutation E459K at the time of starting chemotherapy. Dasatinib was permanently discontinued on Day +16 of the first course of chemotherapy due to HRCT chest abnormalities later attributed to a fungal infection. # Dasatinib was interrupted during the second cycle of chemotherapy due to the development of pleural effusions but was unevenly reintroduced at a reduced dose of 70mg after three weeks.

observations: i) a number of chemotherapeutic agents commonly used in the management of blastic phase CML have been shown to have synergism with a TKI (e.g. cytarabine);^{5,8} and ii) TKIs have been used successfully in combination with conventional chemotherapy for the treatment of Ph⁺ ALL.⁹ We report our experience of dasatinib/FLAG-IDA chemotherapy in 4 CML patients who progressed to blastic phase while on imatinib.

The 4 patients received first-line imatinib for 11, 7, 26 and 10 months, respectively, before progressing to blastic phase. Table 1 shows patients' characteristics. Patients received two courses of FLAG-IDA (G-CSF 300 µg/day sc Days 0 to 6, fludarabine 30mg/m² iv Days 1 to 5, cytarabine 2g/m² iv Days 1 to 5, and idarubicin 12 mg/m² iv Days 1 to 3) together with dasatinib 100 mg daily. Dasatinib was administered continuously from Day 0 of the first course of chemotherapy. The combination was fairly well tolerated (Table 1) and all patients recovered a neutrophil count over 1×10⁹/L within 30 days of the start of combination therapy.

After the first course of chemotherapy, all patients achieved morphological remission (95CI 40-100%); one achieved major cytogenetic response and the remaining 3 achieved both CCyR and major molecular response (MMR). The patients who failed to achieve CCyR after the first course of chemotherapy achieved CCyR and MMR after the second cycle. Currently all 4 patients are alive; Patients 1, 2 and 3 have undergone allogeneic stem cell transplantation (continuing in remission 3, 12 and 13 months post transplant) and patient 4 is scheduled to do so shortly. Dasatinib was reintroduced on Day +30 after transplant in all 3 patients with good tolerance. We have shown that dasatinib can be safely combined with conventional chemotherapy, and although this approach should be tested in a larger number of patients, the combination seems to induce deep remissions in patients with CML in blastic phase, allowing for further therapeutic strategies to enable a continuing response.

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References

1. Druker BJ, Sawyers CL, Kantarjian H, Resta DJ, Reese SF, Ford JM, et al. Activity of a specific inhibitor of the BCR-ABL tyrosine kinase in the blast crisis of chronic myeloid leukemia and acute lymphoblastic leukemia with the Philadelphia chromosome. *N Engl J Med.* 2001;344(14):1038-42.
2. Saglio G, Hochhaus A, Goh YT, Masszi T, Pasquini R, Maloisel F, et al. Dasatinib in imatinib-resistant or imatinib-intolerant chronic myeloid leukemia in blast phase after 2 years of follow-up in a phase 3 study: efficacy and tolerability of 140 milligrams once daily and 70 milligrams twice daily. *Cancer.* 2010;116(16):3852-61.
3. Cortes J, Rousselot P, Kim DW, Ritchie E, Hamerschlag N, Coutre S, et al. Dasatinib induces complete hematologic and cytogenetic responses in patients with imatinib-resistant or -intolerant chronic myeloid leukemia in blast crisis. *Blood.* 2007;109(8):3207-13.
4. Wadhwa J, Szydlo RM, Apperley JF, Chase A, Bua M, Marin D, et al. Factors affecting duration of survival after onset of blastic transformation of chronic myeloid leukemia. *Blood.* 2002;99(7):2304-9.
5. Tipping AJ, Mahon FX, Zafirides G, Lagarde V, Goldman JM, Melo JV. Drug responses of imatinib mesylate-resistant cells: synergism of imatinib with other chemotherapeutic drugs. *Leukemia.* 2002;16(12):2349-57.
6. Kano Y, Akutsu M, Tsunoda S, Mano H, Sato Y, Honma Y, et al. In vitro cytotoxic effects of a tyrosine kinase inhibitor STI571 in combination with commonly used antileukemic agents. *Blood.* 2001;97(7):1999-2007.
7. Scappini B, Onida F, Kantarjian HM, Dong L, Verstovsek S, Keating MJ, et al. In vitro effects of STI 571-containing drug combinations on the growth of Philadelphia-positive chronic myelogenous leukemia cells. *Cancer.* 2002;94(10):2653-62.
8. Deau B, Nicolini FE, Guilhot J, Huguet F, Guerci A, Legros L, et al. The addition of daunorubicin to imatinib mesylate in combination with cytarabine improves the response rate and the survival of patients with myeloid blast crisis chronic myelogenous leukemia (AFR01 study). *Leuk Res.* 2010;35(6):777-82.
9. Rea D, Legros L, Raffoux E, Thomas X, Turlure P, Maury S, et al. High-dose imatinib mesylate combined with vincristine and dexamethasone (DIV regimen) as induction therapy in patients with resistant Philadelphia-positive acute lymphoblastic leukemia and lymphoid blast crisis of chronic myeloid leukemia. *Leukemia.* 2006;20(3):400-3.