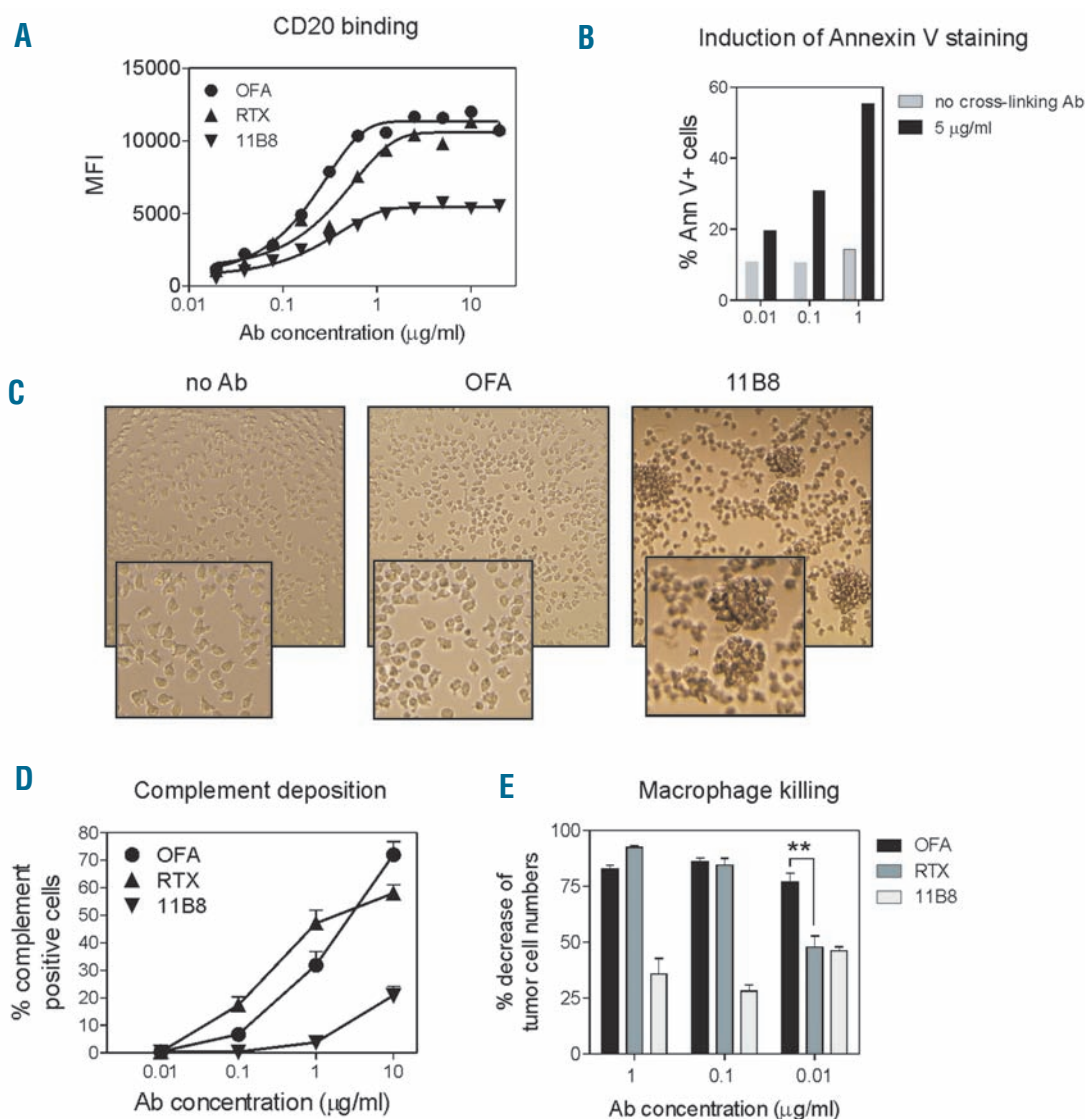


The *in vivo* mechanism of action of CD20 monoclonal antibodies depends on local tumor burden

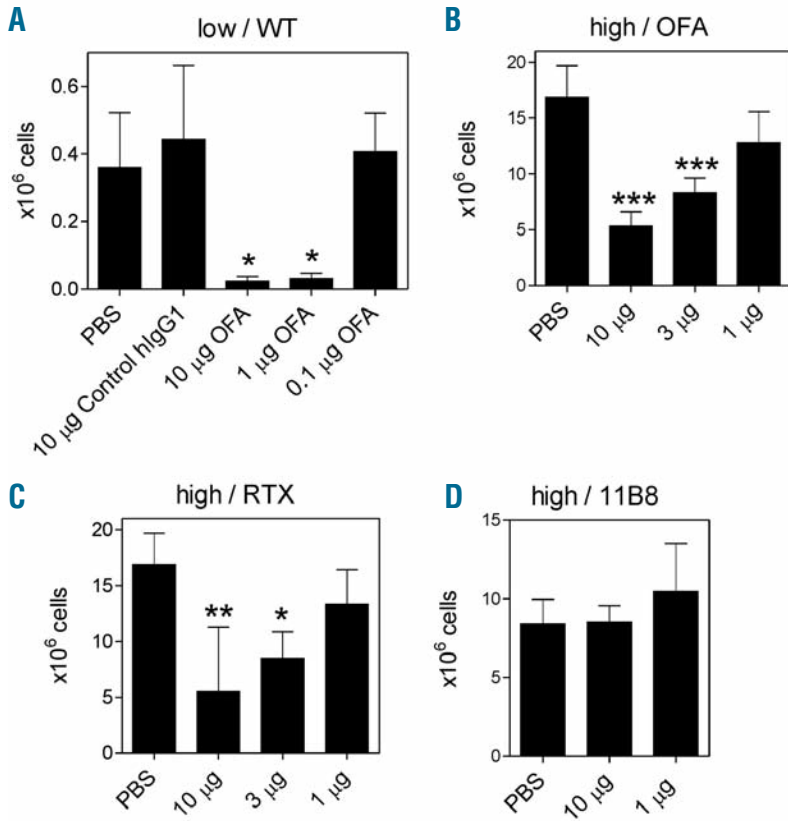
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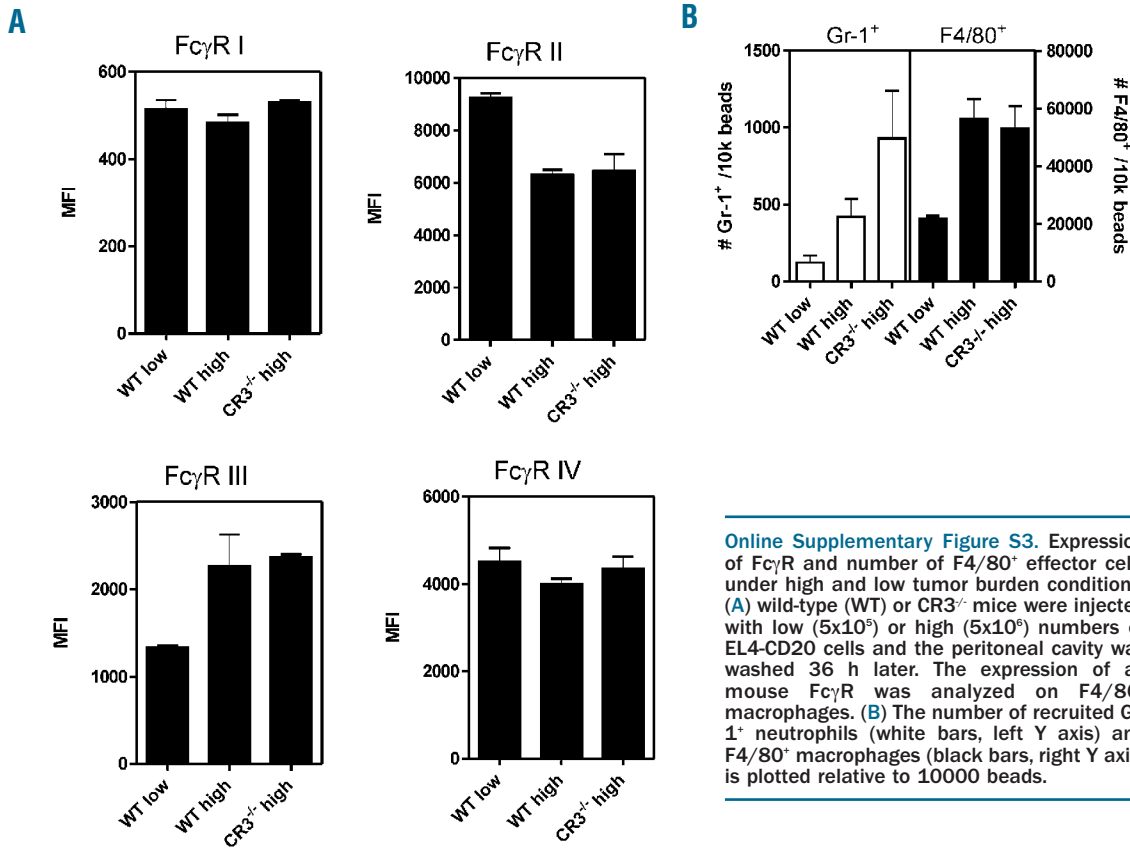
Citation: Boross P, Jansen JHM, de Haij S, Beurskens FJ, van der Poel CE, Bevaart L, Nederend M, Golay J, van de Winkel JGJ, Parren PWHL, and Leusen JHW. The *in vivo* mechanism of action of CD20 monoclonal antibodies depends on local tumor burden. *Haematologica* 2011; 96(12):1822-1830. doi:10.3324/haematol.2011.047159



Online Supplementary Figure S1. *In vitro* characterization of CD20 monoclonal antibody (mAb) using EL4-CD20 target cells. (A) CD20 expression on EL4-CD20 cells *in vitro*. Bound CD20 mAb was detected by FITC-labeled F(ab')₂ fragments of goat anti-human IgG antibody (Ab). Data are representative of two independent experiments. (B) Induction of annexin V positivity after *in vitro* cross-linking of ofatumumab (OFA). Bound OFA was cross-linked by 5 µg/mL rabbit F(ab')₂-anti-human IgG Ab for 24 h. The percentage of annexin V⁺ cells was assessed by cytofluorimetry. Data are representative of six independent experiments. (C) Induction of homotypic aggregates by CD20 mAb binding *in vitro*. EL4-CD20 cells were incubated for 24 h with 1 µg/mL OFA or 11B8 (20x magnification). (D) Complement deposition on EL4-CD20 cells *in vitro*. Pooled (n=3) normal mouse serum (0.5%) was incubated with CD20 mAb opsonized EL4-CD20 cells for 45 min at 37°C. Deposited complement fragments were detected using a mAb against C3b/iC3b/C3c. (E) *In vitro* macrophage killing assay. EL4-CD20 cells were labeled with CFSE and opsonized with CD20 mAb (1, 0.1, 0.01 µg/mL) for 30 min. Bone marrow derived macrophages (BMDM) were incubated with opsonized EL4-CD20 cells for 16 h at a E:T ratio of 10:1. The number of CFSE⁺ EL4-CD20 cells was determined by using a constant amount of beads. The condition without CD20 mAb was used to set the 0% level; the condition without tumor cells was used to set the 100% level and killing is expressed as percentage decrease in cell numbers compared to control (**P<0.01, ANOVA). Data are representative of five independent experiments.



Online Supplementary Figure S2. Titration of CD20 monoclonal antibody (mAb) in the EL4-CD20 model using high and low tumor burden conditions. EL4-CD20 intraperitoneal model: mice were injected i.p. with CFSE-labeled EL4-CD20 cells and 16 h later were given CD20 mAb or phosphate-buffered saline (PBS). After 24 h the number of tumor cells in a peritoneal wash was determined using TrueCount tubes (4-8 mice / group; *** $P < 0.001$, ** $P < 0.01$, * $P < 0.05$; ANOVA). (A) Wild-type (WT) mice were injected with 5×10^5 EL4-CD20 cells and 10 μg non-specific human IgG or various concentrations of ofatumumab (OFA). WT mice were injected with 5×10^6 EL4-CD20 cells and various amounts of OFA (B), rituximab (RTX) (C) or 11B8 (D).



Online Supplementary Figure S3. Expression of FcγR and number of F4/80⁺ effector cells under high and low tumor burden conditions. (A) wild-type (WT) or CR3^{-/-} mice were injected with low (5×10^5) or high (5×10^6) numbers of EL4-CD20 cells and the peritoneal cavity was washed 36 h later. The expression of all mouse FcγR was analyzed on F4/80⁺ macrophages. (B) The number of recruited Gr-1⁺ neutrophils (white bars, left Y axis) and F4/80⁺ macrophages (black bars, right Y axis) is plotted relative to 10000 beads.

Online Supplementary Table S1. Average total numbers of EL4-CD20 cells in all experiments recovered from the peritoneum 24 h after CD20 monoclonal antibody administration. Average fold reduction is calculated compared to numbers from mice treated with phosphate-buffered saline.

		Median	Minimum	Maximum	Average reduction
Low tumor burden (5×10^5)	PBS	3.9×10^5	1.2×10^5	14.2×10^5	
	OFA	0.17×10^5	0.01×10^5	0.46×10^5	95.63%
	RTX	0.18×10^5	0.02×10^5	0.56×10^5	95.37%
	11B8	1.16×10^5	0.001×10^5	3.5×10^5	70.23%
High tumor burden (5×10^6)	PBS	17.76×10^6	1.75×10^6	34.98×10^6	
	OFA	5.32×10^6	0.77×10^6	21.15×10^6	69.96%
	RTX	6.07×10^6	0.09×10^6	16.7×10^6	65.75%
	11B8	14.13×10^6	4.95×10^6	21.16×10^6	20%