The molecular basis of von Willebrand disease: the under investigated, the unexpected and the overlooked

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on Willebrand disease (VWD) is a common autosomally inherited bleeding disorder resulting from quantitative and qualitative defects of plasma von Willebrand factor (VWF). VWD can be divided into three types: type 1 includes partial quantitative deficiencies of VWF, type 3 includes virtually complete quantitative deficiencies while type 2 encompasses four subtypes of qualitative deficiency in which specific VWF ligand-binding functions are affected.¹ In the last two decades there have been studies across the world attempting to elucidate the molecular basis of VWD in order to understand the mechanisms contributing to its pathogenesis more fully. Mutations have been identified throughout the VWF gene (VWF). Missense mutations resulting in alterations to protein structure or function predominate in types 1 and 2 VWD while nonsense mutations (resulting in premature protein termination), deletions or insertions, often resulting in frameshift plus splice-site mutations, predominate in type 3.

Although these common mutations have furthered our knowledge of the molecular pathogenesis of VWD, they fail to explain all cases of the disease. However, detailed analysis of individual patients or cohorts of patients with a defined phenotype is helping to enhance understanding of the disorder. Mutations associated with VWD may be missed by standard polymerase chain reaction (PCR) and DNA sequence analysis or may be identified and their potential effect misinterpreted. Recent advances in understanding of the mutation spectrum contributing to VWD along with mechanisms contributing to other inherited disorders that may play a role in VWD are described below.

Known causative mutation types requiring further investigation in the context of von Willebrand disease

Although reported as causing VWD, certain mutation types require further investigation before their contribution to disease pathogenesis can be fully ascertained.

Copy number variations comprising large deletions within *VWF* have been documented, but historically homozygous deletions were reported only in type 3 VWD, as heterozygous deletions (or duplications) were difficult to identify because of the presence of a normal copy of *VWF*. Patients homozygous for large *VWF* deletions, ranging in size from one exon to complete absence of *VWF*, have been identified, with mutations generally predicted to disrupt

Mutation type Effect on VWF Cause Large deletion Loss of one or more exons Reading frame shift leads to lack of protein expression in type 3 VWD Deletions characterized in types 1 and 2 VWD have been in-frame Large duplication¹ Addition of one or more exons May lead to frame-shift or to larger in-frame protein Transcriptional Disruption of transcription factor binding site Disruption or loss of normal mRNA synthesis or of micro RNA motif1 "Missense" Disruption/creation of exonic splice motif Disruption or loss of normal mRNA synthesis Synonymous exonic Disruption/creation of exonic splice motif Disruption or loss of normal mRNA synthesis Disruption to protein folding¹ Interferes with efficient protein translation Disruption or loss of normal mRNA synthesis e.g. creation of Deep intronic¹ Disruption of normal splicing of introns a pseudoexon from intronic sequence

Table 1. Additional molecular mechanisms identified in or potentially contributing to the pathogenesis of VWD.

¹Not identified in VWD to date.

protein translation and prevent VWF expression.¹ Rare occurrences of large heterozygous deletions causing both types 1 and 2 VWD have also been reported, identified through linkage analysis demonstrating unusual polymorphic marker inheritance^{2,5} or through detection of a previously characterized type 3 VWD deletion in the heterozygous form using a mutation-specific PCR assay.⁴ In cases in which detail has been reported, these deletions are inframe and function in a dominant-negative manner.^{4,5}

Standard PCR and DNA sequencing or mutation scanning techniques are incapable of detecting large heterozygous deletions. Significantly, techniques that can identify homozygous and heterozygous copy number variations, particularly multiplex ligation-dependent amplification (MLPA) have recently begun to identify additional deletions contributing to VWD in both heterozygous type 2 and in compound heterozygous type 3 patients.⁶ The role of copy number variations is, therefore, likely to be underestimated in VWD, as large deletions and exonic duplications are known to contribute to many other disorders, for example hemophilia A.⁷

Another region requiring further investigation is the VWF promoter. Following recent work investigating a Canadian patient with type 1 VWD, the first disease-causing promoter mutation, c.-1522_-1510del, was described.⁸ This mutation disrupts a transcription factor binding site consequently affecting the transcriptional expression of mutant *VWF*. Routine molecular analysis of *VWF* generally focuses on the coding exons and closely flanking intronic sequence.¹ Investigation of the proximal promoter may identify additional transcriptional mutations, especially in patients with mild type 1 VWD manifesting a partial quantitative deficiency of VWF.

Mutations resulting in an unexpected effect on the von Willebrand factor protein

The consequence of an identified mutation does not always result in the expected phenotype. This has been observed in VWD with apparent missense mutations which ordinarily would be expected to affect protein composition. The novel "missense" mutation p.Gly1180 Arg identified in type 2A VWD failed to mimic the disease phenotype when expressed *in vitro* despite segregating with VWD in the affected family.⁹ The reason for this became apparent following further investigation of the patient's RNA. The c.3538G>A nucleotide change that predicted a missense mutation was shown to disrupt a splicing motif causing in-frame deletions of exon 23 (p.Glu990_Lys1036del) and exon 26 (p.Pro1127_Gly1180delinsArg). Additionally, the p.Gly19Arg mutation previously identified in type 1 VWD also fails to influence VWF expression *in vitro* but is predicted *in silico* to influence splicing of exon 2.¹⁰

Novel mutations previously overlooked in von Willebrand disease

In this issue of the journal, Daidone *et al.*¹¹ highlight a mutation previously overlooked in VWF molecular analysis. The authors analyzed all 52 exons, flanking intronic sequence and the 5' and 3' untranslated regions of *VWF* in a family diagnosed with mild-moderate type 1 VWD. Having failed to identify a causative mutation, linkage between VWF and disease phenotype prompted the authors to analyze cDNA from the index case. The analysis identified a heterozygous c.7055_7081del mutation which mimicked disease phenotype when expressed in vitro. As a corresponding nucleotide deletion had not been observed in the genomic DNA (gDNA), it was postulated that the mutation must result from a splicing defect. Re-analysis of the gDNA highlighted the synonymous c.7056C>T (p.Gly2352Gly) alteration which was subsequently predicted, using in silico software, to create a novel splice donor site within exon 41, leading to loss of the remainder of the exon and an inframe deletion of nine amino acids. The mutation results in p.Gly2352_Cys2360del in the B2 domain.¹¹

This study highlights that mutations could be overlooked during standard *VWF* molecular analysis because they are perceived as being not disease-causative. Notably, the c.7056C>T mutation was previously observed in the Canadian type 1 VWD study, but was reported as a polymorphism.¹² Many silent alterations have been reported in *VWF*^{13,14} and may warrant *in silico* analysis to predict any potential influence on splicing along with cDNA analysis to further investigate any predicted effect.

Synonymous alterations have also been shown to affect protein function by interfering with protein folding, which subsequently disrupts protein translation. The synonymous c.459G>A (p.Val153Val) mutation in *F9* has recently been shown to cause mild hemophilia B in this manner.¹⁵

Further mutations may be under-investigated in VWF.

Both copy number variations and point mutations occurring deep within intronic sequences have been shown to interfere with normal splicing patterns and result in the formation of pseudoexons in other disorders.^{16,17} It is also feasible that deep intronic mutations could disrupt transcription factor binding site or micro RNA motifs, particularly if occurring within terminal introns.

Concluding remarks

Studies such as the one reported by Daidone et al.¹¹ emphasize that additional work is required in order to understand the molecular basis of VWD fully. New technologies such as next generation sequencing will highlight additional sequence variants that may contribute to VWD, but synonymous mutations are still likely to be overlooked. It is, therefore, beneficial to combine gDNA molecular analysis with information from several additional sources including disease and gene co-segregation within the family, any previous reports of the sequence variant, ^{13,14} in silico analysis to investigate predicted effects of sequence variants on the protein and on mRNA splicing plus analysis of cDNA when possible.¹⁸⁻²⁰ Mutation prediction packages [for example, Alamut® (Interactive Biosoftware, Rouen, France)] combine several tools and facilitate these analyses. Given the complexity of VWD, it is perhaps unsurprising that its molecular basis also appears complex. However, whether "overlooked" mutations are rare anomalies or significant contributors to the pathogenesis of VWD remains to be answered.

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References

- 1. Goodeve AC. The genetic basis of von Willebrand disease. Blood Rev. 2010;24(3):123-34.
- 2 Johansson AM, Lanke E, Säll T, Lethagen S, Halldén C. A large deletion identified in a Swedish family with type 1 VWD. Thromb Haemost.

2011:105(4):733-4.

- 3. Bernardi F, Marchetti G, Guerra S, Casonato A, Gemmati D, Patracchini P, et al. A de novo and heterozygous gene deletion causing a variant of von Willebrand disease. Blood. 1990;75(3):677-83.
- 4. Sutherland MS, Cumming AM, Bowman M, Bolton-Maggs PHB, Bowen DJ, Collins PW, et al. A novel deletion mutation is recurrent in von Willebrand disease types 1 and 3. Blood. 2009;114(5):1091-8.
- Casari C, Pinotti M, Lancellotti S, Adinolfi E, Casonato A, De Cristofaro R, et al. The dominant-negative von Willebrand factor gene deletion p.P1127_C1948delinsR: molecular mechanism and modulation. Blood. 2010;116(24):5371-6.
- Yadegari H, Driesen J, Hass M, Budde U, Pavlova A, Oldenburg J. Large deletions identified in patients with von Willebrand disease by MLPA. J Thromb Haemost. 2011;9(5):1083-6.
- 7. Zimmermann MA, Oldenburg J, Müller CR, Rost S. Characterization of duplication breakpoints in the factor VIII gene. J Thromb Haemost. 2010;8(12):2696-704.
- 8. Othman M, Chirinian Y, Brown C, Notley C, Hickson N, Hampshire D, et al. Functional characterization of a 13-bp deletion (c.-1522_-1510del13) in the promoter of the von Willebrand factor gene in type 1 von Willebrand disease. Blood. 2010;116(18):3645-52
- 9. James PD, O'Brien LA, Hegadorn CA, Notley CRP, Sinclair GD, Hough C, et al. A novel type 2A von Willebrand factor mutation located at the last nucleotide of exon 26 (3538G>A) causes skipping of 2 nonadjacent exons. Blood. 2004;104(9):2739-45.
- Eikenboom J, Hilbert L, Ribba AS, Hommais A, Habart D, Messenger S, et al. Expression of 14 von Willebrand factor mutations identified in patients with type 1 von Willebrand disease from the MCMDM-1VWD study. J Thromb Haemost. 2009;7(8):1304-12.
- 11. Daidone V, Gallinaro L, Cattini MG, Pontara E, Bertomoro A, Pagnan A, et al. An apparently silent nucleotide substitution (c.7056C>T) in the von Willebrand factor gene is responsible for type 1 von Willebrand disease. Haematologica. 2011;96(6):881-7.
- 12. James PD, Notley C, Hegadorn C, Leggo J, Tuttle A, Tinlin S, et al. The mutational spectrum of type 1 von Willebrand disease: results from a Canadian cohort study. Blod. 2007;109(1):145-54. 13. NCBI. db SNP. [cited 30/04/2011]; Available from: http://www.ncbi.
- nlm.nih.gov/projects/SNP/
- 14. VWFdb. International Society on Thrombosis and Haemostasis Scientific and Standardization Committee VWF Information Homepage. [cited 30/04/2011]; Available from: www.vwf.group.shef.ac.uk
- 15. Kimchi-Sarfaty C, Simhadri VL, Kopelman D, Friedman A, Edwards N, Javaid A, et al. The synonymous V107V mutation in factor IX is not so silent and may cause hemophilia B in patients. Blood. 2010;116 (21):abst 2197.
- 16. Khelifi MM, Ishmukhametova A, Khau Van Kien P, Thorel D, Méchin D, Perelman S, et al. Pure intronic rearrangements leading to aberrant pseudoexon inclusion in dystrophinopathy: a new class of mutations? Hum Mutat. 2011;32(4):467-75.
- 17. Castoldi E, Duckers Č, Radu C, Spiezia L, Rossetto V, Tagariello G, et al. Homozygous F5 deep-intronic splicing mutation resulting in severe factor V deficiency and undetectable thrombin generation in platelet rich plasma. J Thromb Haemost. 2011;9(5):9599-68.
- Bell J, Bodmer D, Sistermans E, Ramsden S. Practice guidelines for the 18. Interpretation and Reporting of Unclassified Variants (UVs) in Clinical [cited 30/04/2011]; Molecular Genetics. Available from: http://cmgsweb.shared.hosting.zen.co.uk/BPGs/Best_Practice_Guideli nes.htm
- 19. Houdayer C, Dehainault C, Mattler C, Michaux D, Caux-Moncoutier V, Pagès-Berhouet S, et al. Evaluation of in silico splice tools for decision-making in molecular diagnosis. Hum Mutat. 2008;29(7):975-82.
- 20. Keeney S, Bowen D, Cumming A, Enayat S, Goodeve A, Hill M. The molecular analysis of von Willebrand disease: a guideline from the UK Haemophilia Centre Doctors' Organisation Haemophilia Genetics Laboratory Network. Haemophilia. 2008;14(5):1099-111.