

# Regulation of PTEN by CK2 and Notch1 in primary T-cell acute lymphoblastic leukemia: rationale for combined use of CK2- and $\gamma$ -secretase inhibitors

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## Supplementary information

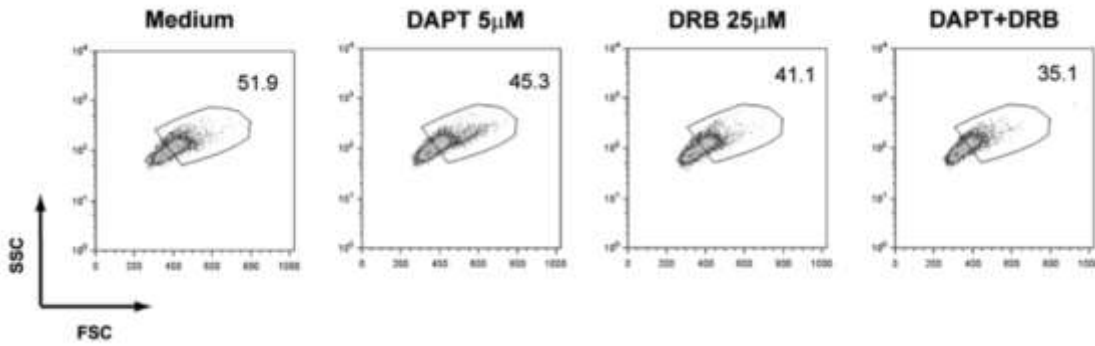
The cycling protocol for *NOTCH1* amplifications consisted of 1 cycle at 94°C for 3 min, followed by 10 cycles of annealing at decreasing temperatures from 61°C to 57°C (1°C decrease every two cycles) for 30 s, extension at 72°C for 1:45 min and denaturation at 95°C for 30 s. Subsequently, 30 cycles were performed with annealing at 56°C for 30 s. A final extension step for 5 min at 72°C was included. PCR reactions contained 50ng of DNA or 0.5  $\mu$ l cDNA, 10 pmol of each primer, 2 mM MgCl<sub>2</sub>, 5% DMSO, 1X PCR Buffer (Biotools), and 1.5 U of Taq DNA Polymerase (Biotools). The PCR cycling protocol for *PTEN*, *HES1*, and *ABL* consisted of 10 min at 95°C, followed by 40 cycles of 58°C for 30 s, 72°C for 40 s, 95°C for 15 s, and a final extension step for 5 min at 72°C. For *MYC* a two-step cycling protocol was used consisting of 10 min at 95°C, followed by 40 cycles of annealing/extension at 65°C for 1 min and 95°C for 15 s, and a final extension step for 5 min at 72°C. A standard melting-curve cycle was used to check the quality of amplification, such as no primer dimer being formed during PCR. Fluorescence was measured at 72°C.

Online Supplementary Table S1. Primers used for *NOTCH1* sequencing and RQ-PCR analysis of *HES1*, *MYC*, *PTEN*, and *ABL*.

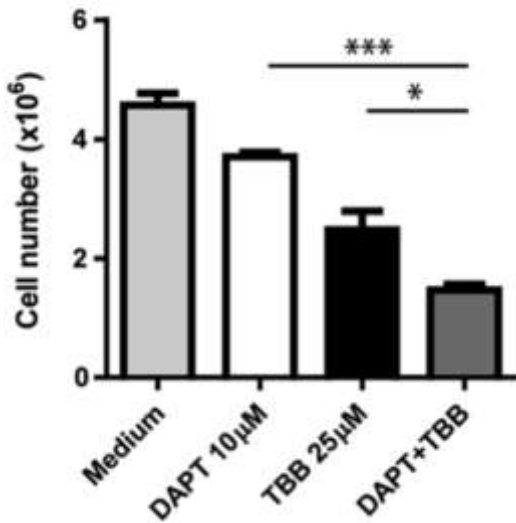
Primer	Primer sequence (5'→3')	Reference
<b>Primers for PCR and sequencing of <i>NOTCH1</i></b>		
Notch2627F	GGACTGTGCGGAGCATGTA	1
Notch2627R	CAGCCCACGAAGAACAGAA	1
Notch26F	CAGTGGGCTGGAGGCA	1
Notch26R	GGGGATTGACCGTGGG	1
Notch27F	GGTGGGTATCTGGGATGAGC	1
Ex27R743	GAAAAGCCCTACCCCAACTC	2
Notch34TF	TTTGAATGGTCAATCGGAGTG	1
Notch34PR	AAAAGGCTCCTCTGGTCG	1
Ex34PestF	CCTCACCTGGTGCAGACC	2
Ex34PestR	AAATAAAATCCTCGTTCCTTTTGG	2
Notch34PF	AGCCAGCAAACATCCAGC	1
<b>Primers for RQ-PCR</b>		
HES1-F	TGGAATGACAGTGAAGCACCT	3
HES1-R	GTTTCATGCACTCGCTGAAGC	3
MYC-F	CTTCTCTCCGTCCTCGGATTCT	4
MYC-R	GAAGGTGATCCAGACTCTGACCTT	4
PTEN-F	GCTACCTGTTAAAGAATCATCTGG	1
PTEN-R	CATGAACCTGTCTTCCCGT	5
ABL-F	TGGAGATAACACTCTAAGCATAACTAAAGGT	6
ABL-R	GATGTAGTTGCTTGGGACCCA	6

## References

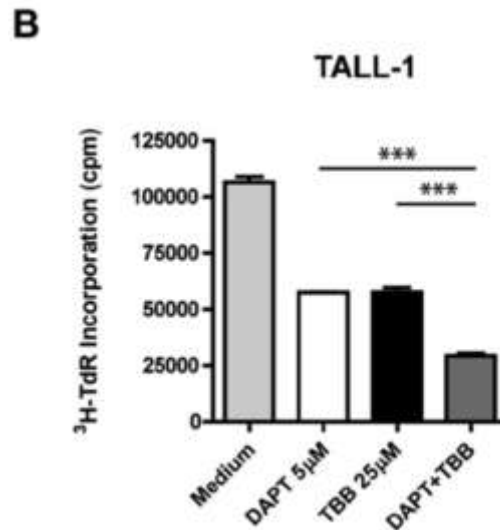
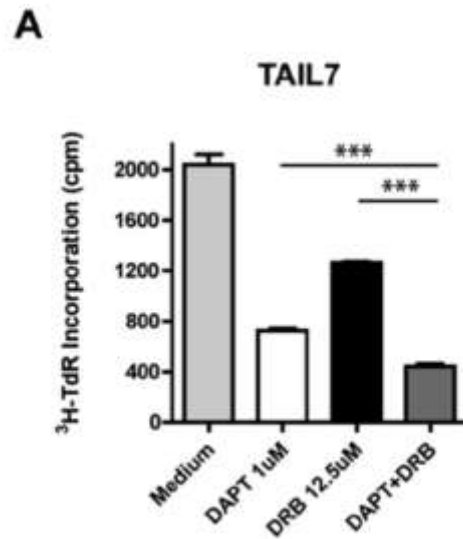
1. This work. PTEN forward primers were designed to provide the most possible difference to the PTEN pseudogene transcript.
2. Kindly provided by Dr Adolfo Ferrando, Columbia University, NY.
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Online Supplementary Figure S1. Cooperative effects of GS- and CK2 inhibitors in decreasing the size of TAIL7 cells. TAIL7 cells were cultured in the indicated conditions for 72h and cell size was determined by FSC x SSC discrimination by flow cytometry analysis.



Online Supplementary Figure S2. Cooperative effects of GS- and CK2 inhibitors in decreasing HPB-ALL cell numbers. HPB-ALL cells were cultured in triplicates for four days in the indicated conditions and number of viable cells was assessed by trypan blue exclusion using a hemocytometer. \* $P < 0.05$ ; \*\*\* $P < 0.001$ .



Online Supplementary Figure S3. Cooperative effects of GS- and CK2 inhibitors in decreasing TAIL7 and TALL-1 cell proliferation. TAIL7 (A) and TALL-1 (B) cells were cultured in triplicates for seven days in the indicated conditions and proliferation was evaluated by assessing DNA synthesis by  $^3\text{H}$ -thymidine incorporation. \*\*\* $P < 0.001$ .