## Seven novel mutations of the UGT1A1 gene in patients with unconjugated hyperbilirubinemia

The aim of this study was to identify new pathogenic variations of the UGT1A1 gene in 11 patients diagnosed with neonatal unconjugated hyperbilirubinemia. We describe two cases in which clinically unapparent heterozygotic mutations in the UGT1A1 gene may become evident in combination with certain environmental conditions or additional genetic defects.

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Genetic alterations of the *UGT1A1* gene result in Crigler-Najjar syndrome type I (CN1) and type II (CN2) and Gilbert's syndrome, autosomal recessive conditions that are characterized by non-hemolytic unconjugated hyperbilirubinemia.<sup>1-3</sup> Several other disorders associated with hemolysis caused by premature destruction of ery-throcytes are also characterized by hyperbiliribinemia.<sup>45</sup> Therefore, the expression of *UGT1A1* may be a major factor responsible for bilirubin variability (a modifier gene) in inherited hemolytic diseases.

The aim of this study was to analyze the mutation pattern causing unconjugated hyperbilirubinemia in 11 unrelated patients with neonatal hyperbilirubinemia. For the analysis of mutations of *UGT1A1* genes, the coding and promoter region and the splicing sites were analyzed by direct sequencing.<sup>6</sup>

Nine patients had a clinical diagnosis of CN1 (patients

Table 1. The patients' clinical features and mutations.

#1 to #4) and CN2 (patients #5 to #9). Two subjects (patients #10 and #11) presented with transient hyperbilirubinemia caused by hemolysis. Patient #10 had neonatal kernicterus associated with hyperbilirubinemia due to a skull hematoma. Phototherapy and phenobarbital were started in this neonate, whose bilirubin levels decreased shortly afterwards. The phototherapy was suspended when the bilirubin concentration was under 10 mg/dL following resorption of the hematoma. Patient #11, affected by glucose-6-phosphate dehydrogenase (G6PD) deficiency, showed increased bilirubin production due to hemolysis. Direct sequencing of G6PD revealed a change from cytosine to thymine at base position 563 (in exon 6) causing a change from serine to phenylalanine in amino acid position 188. Results of the analysis of UGT1A1 are summarized in Table 1.

We identified a total of 12 sequence variations, seven of which are described for the first time.

Two out of the four CN1 patients had a frameshift mutation, p.Q239fsX256, a recurrent change already found in a cohort of Italian patients.<sup>7</sup> Patients #3 and #4 showed novel variations. Patient #3 had a homozygous tri-nucleotide deletion c.513\_515 del CTT causing a deletion of leucine codon at position 172 and the substitution of a phenylalanine residue by a leucine at position 171 [p.I.172delF171L]. Although, this mutation eliminates a single amino acid (leucine), we believe that the substitution of a phenylalanine in a conserved diphenylalanine region of an aglycone binding domain may abolish the enzymatic activity of UGT1A1. Patient #4 was heterozygous for two novel mutations: a c.652insT insertion causing a frameshift and a C $\rightarrow$ T transition at nucleotide 847 in exon 1 that introduces a stop codon (p. Q283X).

Patient	Clinical manifestation	Biliribun levels (mg/dL)	Gestational age weeks	Birth weight (Kg)	Mutation	Mutant Protein	(TA) Polymorphism	References
#1	CN1	30	38	3,00[	c.717-718 del AG] [c.717-718 del AG]	[p.Q239fsX256]+ [p.Q239fsX256]	(TA) <sup>7</sup> (TA) <sup>7</sup>	Iolascon, 2000
#2	CN1	28,8	38	3,30	[c.717-718 del AG]+ [c.717-718 del AG]	[p.Q239fsX256]+ [p.Q239fsX256]	(TA) <sup>7</sup> (TA) <sup>7</sup>	Iolascon, 2000
#3	CN1	32,05	40	3,45	[c.513-515 del CTT]+ [c.513-515 del CTT]	[p.L172delF171L]+ [p.L172delF171L]	(TA) <sup>7</sup> (TA) <sup>7</sup>	Novel
# 4	CN1	31,2	37	3,05	[c.652insT]+[c.847C→T]	[p.S218Ffs257X]+ [p.Q283X]	(TA) <sup>6</sup> (TA) <sup>6</sup>	Novel+Novel
<b>#</b> 5	CN2	19	38	3,25	[c.835 A→T]	[p.N279Y]	(TA) <sup>6</sup> (TA) <sup>7</sup>	Elferink, 1994
#6	CN2	16,21	39	3,10	[c.1381 T→C]	[p.W461R]	(TA) <sup>6</sup> (TA) <sup>6</sup>	Maruo, 2003
# 7	CN2	17,9	40	3,60	[c.1223-1224 ins G]+ [c.1184G→T]	[p.A409SfsX422]+ [p.G395V]	(TA) <sup>6</sup> (TA) <sup>6</sup>	Labrune, 1994+ Servedio, 2005
#8	CN2	10	37	2,95	[c.1328 T→C]	[p.L443P]	(TA) <sup>6</sup> (TA) <sup>7</sup>	Novel
<b>#</b> 9	CN2	7,29	38	3,10	[c.1060 T→A]	[p.W354R]	(TA) <sup>6</sup> (TA) <sup>7</sup>	Novel
#10 h	nyperbil. due to hematoma	27,7	40	3,60	[c.1108 A→G]	[p.1370V]	(TA) <sup>6</sup> (TA) <sup>6</sup>	Novel
#11	G6PD	11,9	38	3,15	[c.210delC]	[p.D70QfsX76]	(TA) <sup>6</sup> (TA) <sup>6</sup>	Novel

GT1A1 cDNA sequence from GenBank accession number NM\_000463NM was used as the reference sequence: the A of the ATG translation initiation start site represents nucleotide +1. CN1: Crigler-Najjar type I; CN2: Crigler-Najjar type I; G6PD: glucose-6-phosphate dehydrogenase deficiency.

The milder phenotype of CN2 patients is usually associated with homozygosity or compound heterozygosity for missense mutations. In addition to known causative alterations (p.N279Y, p.W461R), two missense mutations were identified in heterozygous subjects affected by CN2. In patient # 8, a novel missense mutation c.1328  $T \rightarrow C$  was found in exon 5 which affected residue 443 (p. L443P) involving a polar amino acid substituition. In patient #9 a T $\rightarrow$ A heterozygous transition at nucleotide 1060 of the UGT1A1 gene resulted in the substitution of a tryptophan residue by arginine at position 354 (p.W354R) in the carboxy-terminal domain of the UGT1A1 protein. Recently, missense p.W354R was reported to be caused by substitution of T to C at nucleotide 1060.7 These results suggest that nucleotide 1060 in UGT1A1 may be a mutational hot spot. In our cohort, all the individuals affected by CN2, except patient #7, were heterozygous for one UGT1A1 coding region variation. However, the in trans presence of a c.-41\_-40dupTA polymorphism [(TA)<sup>7</sup>] can explain the CN2 phenotype. This polymorphism can play a role in enhancing the effect of the heterozygous coding mutation.

Here, we describe two cases in which clinically unapparent heterozygotic mutations in UGT1A1 may become evident when combined with certain environmental conditions or additional genetic defects. In patient #10 a novel missense mutation, c.1108 A $\rightarrow$ G, was detected in a heterozygous state. This change caused an amino acid substitution (p.I370V) in the catalytic core of the UGT1A1 protein. This patient's severe hyperbilirubinemia and kernicterus were consequences of the imbalance between bilirubin production and conjugation. In patient #11, affected by G6PD deficiency, a novel heterozygous small deletion of one nucleotide at position 210 (c.210delC) of the UGT1A1 gene was found in combination with the Mediterranean variant, c.563 C $\rightarrow$ T of the G6PD gene. This represents an example of co-inherited modifying gene causing clinical heterogeneity in monogenic disorders. The expression of UGT1A1 is a major modifying factor in inherited hemolytic diseases being responsible for a large proportion of the bilirubin variability in these conditions.

In conclusion, the identification of these novel mutations in the *UGT1A1* gene, increasing the mutational spectrum of *UGT1A1* allelic variants, contributes to a better understanding of the molecular pathology of disorders characterized by unconiugated hyperbilirubinemia.

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