Acute Lymphoblastic Leukemia

MLL/GAS7 fusion in a pediatric case of t(11;17)(q23;p13)-positive precursor B-cell acute lymphoblastic leukemia

MLL/GAS7, resulting from t(11;17)(q23;p13), has been reported in one case of treatment-related acute myeloid leukemia (AML). We present a de novo case of t(11;17)-positive pediatric acute lymphoblastic leukemia. Fluorescent in situ hybridization and reverse transcriptase polymerase chain reaction analyses revealed an MLL/GAS7 chimera identical to the one previously described in AML. The molecular genetic features of MLL/GAS7 and the clinical impact of t(11;17) are discussed.

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To date, the 5' part of MLL has been shown to fuse with the 3' part of approximately 50 different genes in acute myeloid leukemia (AML), chronic myeloid leukemia,

myelodysplastic syndromes, acute lymphoblastic leukemia (ALL), and lymphomas. Some of these fusions are quite common, such as MLL/AFF1(AF4) [t(4;11)(q21;q23)] in ALL and *MLL/MLLT3(AF9)* [t(9;11)(p21;q23)] in AML, whereas many of them have only been reported in single cases, such as the MLL/GAS7 [t(11;17)(q23;p13)] in therapy-related AML.12 Although the prognostic impact of the frequent MLL fusions is well known, 3,4 the clinical ramifications of the infrequent chimeras are unknown. Considering that most treatment protocols are based on the presence of certain genetic changes in AML and ALL, it is important to describe such rare MLL fusions, with the aim that other groups will report additional cases, thus increasing the clinical information available for the medical community, as recently exemplified by the rare MLL/ARHGAP26(GRAF) fusion, generated by a t(5;11)(q31;q23), in pediatric AML.5 For this reason, we present here the molecular genetic and clinical features of the first t(11;17)(q23;p13)-positive ALL shown to harbor the MLL/GAS7 chimera.

A previously healthy 15-month old girl was admitted to hospital in April 2005 because of fever and abnormal blood values (hemoglobin 62 g/L, platelet count  $68\times10^{\circ}/L$ , and white blood cell count  $3.9\times10^{\circ}/L$  with 28% blasts). The clinical examination revealed hepatomegaly; there was no splenomegaly or mediastinal mass. The bone marrow was

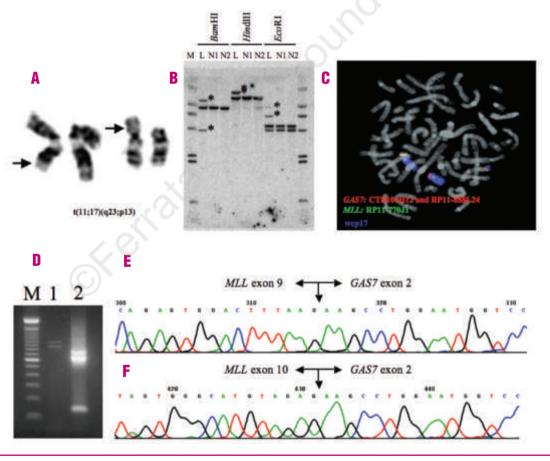


Figure 1. A. Partial karyogram showing the t(11;17)(q23;p13). The derivative chromosomes 11 and 17 and the chromosomal breakpoints are indicated by arrows. B. Southern blot analysis of BamHI-, HindIII- and EcoRI-digested DNA from the t(11;17)-positive ALL (L) and from two healthy individuals (N1 and N2) using the MLL probe B859. Aberrant fragments are indicated by asterisks. M is  $\lambda$  DNA/HindIII fragments. C. FISH using RP11-770J1 for the MLL gene (green signal), CTB-104H12 and RP11-888L24 for the GAS7 gene (red signal), and wcp17 (blue signal). Fusion signals of MLL and GAS7 are seen on both the der(11) and the der(17) of the t(11;17). D. Amplification of a chimeric MLL/GAS7 transcript. Total RNA was reverse-transcribed, and cDNA was used as a template in PCR amplification using MLL-3735F (CCCATCAGCAAGAGAGGAGCATCCTGC) and GAS7-700R (CATCGTGTCTGG GTGAGGAACG) primers (Lane 1). Two microliters of the first PCR product were used in a second PCR with MLL-3878F (AGTCAAGCAAGCAGGTCTCCCAGC) and GAS7-634R (CTGCTTTTTGCTTGGCGATGAGG) (Lane 2). M: 100 bp DNA ladder. E and F. Partial sequence chromatogram showing that nt 4241 (NM\_005933.2; exon 9) of MLL was fused in-frame with nt 345 (NM\_201433.1; exon 2) of GAS7 (E) and that nt 4355 (exon 10) of MLL was also fused in-frame with nt 345 of GAS7 (F).

Table 1. Clinical, immunophenotypic, and genetic features of t(11;17)(q23;p13)-positive acute leukemias reported in the literature.

Reference	Diagnosis	Sex/age	CNS	WBC (×10°/L)	Immunophenotype	Cytogenetics	Molecular genetics	Survival (mo)
10	AL bi	M/5 mo	Yes	400	HLA-DR°, CD19°, CD33°, CD13°, TdT°, CD10°, CD20°, sig°, CD14°	t(11;17) (Dx) t(11;17),+8 (Rel)	NR	<1
2	t-AML M4	M/14 yrs	NR	NR	NR	t(11;17)	MLL/GAS7	4
9	ALL	M/9 mo	NR	900	pro-B	t(11;17),+22	MLL R	59+
Present case	ALL	F/15 mo	No	3.9	CD34°, CD19°, CD22°, CD24°, CD15°, CD123°, TdT°, cCD79°, CD10°	t(11;17)	MLL/GAS7	14+

AL bi: acute biphenotypic leukemia (FAB type M1/L2); ALL: acute lymphoblastic leukemia; CNS: central nervous system involvement; Dx: diagnosis; F: female; M: male; mo: months; NR: not reported; R: rearrangement; Rel: relapse; t-AML M4: treatment-related acute myeloid leukemia (FAB type M4) occurring chemotherapy (cyclophosphamide, doxorubicin, vincristine, cisplatin, and etoposide), radiotherapy, and surgery for neuroblastoma; WBC: white blood cell count; yrs: years; +: alive at the time of reporting.

hypercellular, containing mainly blasts. Flow cytometric four-color analysis showed that 95% of the bone marrow cells were positive for CD34, CD19, CD22, CD24, CD15. CD123, TdT, and cCD79a but negative for CD10. A diagnosis of precursor B-cell ALL was made. Genetic analyses of a diagnostic bone marrow sample revealed the karyotype 46,XX,t(11;17)(q23;p13)[25] and an MLL rearrangement (Figure 1A and B). Further molecular studies were negative for BCR/ABL1, ETV6/RUNX1, TCF3/PBX1, and FLT3 mutations. A monoclonal IgH rearrangement was identified, characterized and used for evaluating minimal residual disease by real-time polymerase chain reaction (PCR). Based on the finding of an MLL rearrangement, the girl continued treatment according to the intensive arm of the Nordic NOPHO-ALL 2000 protocol. Complete cytogenetic, fluorescent in situ hybridization, morphologic, flow cytometric, and PCR remission was achieved on day 29. At present, 14 months after diagnosis, she remains in remission. Southern blot analysis, using the MLL probe B8596, yielded two extra bands in the BamHI, HindIII, and EcoRI digestions (Figure 1B). The t(11;17)(q23;p13) was then further characterized by FISH using wcp11, wcp17, the bacterial artificial chromosome (BAC) RP11-770J1 (covering MLL), and BAC CTB-104H12 and RP11-888L24 for the GAS7 locus. The FISH analysis revealed fusion signals of MLL and GAS7 on both derivative chromosomes 11 and 17 (Figure 1C). Five micrograms of total RNA were reverse-transcribed, PCR-amplified, and sequenced as described previously.6 The reverse transcription (RT)-PCR with MLL-3735F and GAS7-700R, nested PCR with MLL-3878F and GAS7-634R, and sequence analyses of the two amplified cDNA fragments revealed an in-frame fusion of exon 9 of MLL with exon 2 of GAS7 and, in the smaller fragment, of exon 10 of MLL with exon 2 of GAS7 (Figure 1D-F), both identical to the previously reported MLL/GAS7 transcripts in AML.2

As a consequence, the translocation places the GAS7 gene under the control of the MLL promoter. The putative MLL/GAS fusion protein would retain the AT-hook DNA binding domain, the DNA methyl transferase motif, and the transcription repression domain of MLL and all the functional domains of the GAS7 protein.2 MLL/GAS7 has been shown to transform multipotent hematopoietic progenitors and to induce mixed lineage leukemia in mice.8 The present case shows that the MLL/GAS7 fusion may result in ALL as well as in AML, akin to what has been reported for other MLL chimeras.

Only four acute leukemias with t(11;17)(q23;p13), two of which with confirmed MLL/GAS7 chimeras, have been reported to date (Table 1). Thus, it is difficult to draw any firm conclusions as regards its clinical impact. However, all patients have been children and pronounced leukocytosis

was reported in two of them. Furthermore, it may be noteworthy that both patients with ALL, i.e., the present case and one previously published, 9 responded well to treatment. On the other hand, the two patients with AML both succumbed to their disease within a few months. Thus, the prognostic impact of MLL/GAS7 may differ between AML and ALL.

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