Myeloproliferative Disorders

Limited effects on JAK2 mutational status after pegylated interferon α -2b therapy in polycythemia vera and essential thrombocythemia

Twenty-five patients with myeloproliferative diseases were treated with pegylated interferon α -2b. Prior to therapy, 15/25 patients had a $JAK2^{V617F}$ mutation. Eight JAK2-positive patients were on therapy in hematological complete remission at 24 months. Five of eight patients demonstrated a 1.2-3.6 fold reduction in the percentage of $JAK2^{V617F}$ cells.

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We have published clinical results of a 24-month phase II study of pegylated interferon α-2b (PegIntron[®]). Schering-Plough, PEG-IFN) therapy in polycythemia vera (PV) and essential thrombocythemia (ET). Briefly, patients were treated with PEG-IFN 0.5-1.0 µg/kg/ week. Twenty-nine of 42 patients (69%) achieved a complete platelet response, i.e a platelet count < 400×10⁹/L (symptomatic patients) or <600×10°/L (asymptomatic patients). Nineteen patients (45%) completed the 2-year treatment in complete remission.1 Similar results have been reported by others.^{2,3} We detected normalization of initially elevated polycythemia rubra vera-1 (PRV-1) expression in a subset of patients.1 Case reports have suggested that interferon can reverse chromosome abnormalities,4 restore polyclonal hematopoiesis and suppress erythropoietin-independent erythroid colony growth. 5 We therefore investigated the potential of PEG-IFN to suppress the malignant clone using the JAK2^{V617F} mutation as a disease marker 6

For this investigation, frozen samples were available from 25 patients, 14 with PV and 11 with ET. Sixteen were male and nine were female; their median age was 52 years (range 29-77), and the median duration of disease was 0.5 years (range 0.01-23.2). Prior cytoreductive treatment included anagrelide (n=4), hydroxyurea (n=2), busulfan (n=1) and radioactive phosphorus (n=1). Expression of PRV-1 mRNA was quantified in neutrophils as previously described.7 The allele ratio of mutant IAK2^{V617F} to total JAK2 was determined by a quantitative reverse transcriptase polymerase chain reaction (RT-PCR) assay of purified granulocyte RNA. Total JAK2 mRNA was determined with a forward primer 5' CAGCAAG-TATGATGAGCAAGCTTT-3', a reverse primer 5'-TGAACCAGAATATTCTCGTCTCCAC-3' and the MGB-Probe 5'-FAM-TCACAAGCATTTGGTTTT-MGB-3'. $JAK2^{V617F}$ was quantified using the same forward primer and probe but a reverse primer comprising the mutation and an additional mismatch at position 4 5'-CCAGAATATTCTCGTCTCCACTGAA-3'. The allele copy numbers were determined from a plasmid standard curve and the allele ratio was calculated. The level of *JAK2*-positivity is expressed as the percentage of mutant JAK2 compared to total JAK2. A percentage of <1% JAK2-positive cells was defined as JAK2-negative. This cut off was determined in a panel of 50 healthy controls (Goerttler and Pahl, unpublished observation).

Table 1 shows $JAK2^{v617F}$ and PRV-1 status prior to therapy. A good correlation between presence of the $JAK2^{v617F}$ mutation and PRV-1 overexpression was found in PV, as previously described.⁸ In the 15 JAK2-positive patients,

Table 1. PRV-1 and JAK2 status prior to therapy according to diagnosis.

Diagnosis	JAK2*/	JAK2*/	JAK2 ⁻ /	JAK2 ⁻ /
	PRV-1*	PRV-1 ⁻	PRV-1*	PRV ⁻ 1 ⁻
PV ET	10 3	1	2 2	1 5

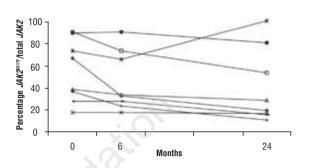


Figure 1. JAK2 mutational status in eight patients before therapy, and after 6 and 24 months of PEG-IFN therapy. No differences with regards to clinical features, white blood cell or platelet counts prior to or after therapy were noted between patients in whom JAK2 was downregulated and those in whom it was not (data not shown).

the percentage of mutant $JAK2^{V617F}$ ranged from 18-90% (mean 44%). The mean value in PV (49%) was somewhat higher than that in ET (32%).

Thirteen of 15 JAK2-positive and 8/10 JAK2-negative patients achieved complete remission with PEG-IFN. Although the numbers are small, it does not seem that *JAK2* status is related to response to PEG-IFN. Follow-up samples for molecular studies were only available in patients still on therapy, therefore clinical response data in this highly selected subset of patients are not presented in detail. After 6 months 12 IAK2-positive patients were on therapy in complete remission. Four of these patients had a reduction of JAK2-positivity by at least 10% in five patients JAK2-positivity remained unchanged, and three had an increase. The further evolution of JAK2 mutational status during therapy is shown in Figure 1. After 24 months, eight JAK2-positive patients (6 PV, 2 ET) were on therapy in hematologic remission: platelet count <400×109/L and in PV also a stable hematocrit <45 without phlebotomies. Five (3 PV, 2 ET) of eight patients had a 1.2-3.6-fold reduction of the percentage of JAK2^{V617F}, the percentages were unchanged in two and one had an increase.

Jones *et al.* reported that the median percentage of mutated JAK2 alleles was not different between PV patients treated with phlebotomy alone, hydroxyurea, anagrelide or imatinib. They found a significantly lower level of $JAK2^{V617F}$ in seven IFN-treated patients than in the other patient groups, and hypothesized that IFN therapy had reduced $JAK2^{V617F}$ levels. Our study clearly demonstrates that, in selected patients, PEG-IFN can reduce, to a limited extent, the level of JAK2-positivity. In this context, it should be stressed that the goal of our clinical trial was not to suppress $JAK2^{V617F}$ levels, but rather to maintain complete remission with the lowest PEG-IFN dose possible. This may have led to smaller effects on JAK2

due to a low mean dose, 0.3 µg/kg/week, at 24 months. In conclusion, PEG-IFN therapy can lower the percentage of circulating $IAK2^{V617F}$ mutant cells, but the effect is modest. Even in sustained hematologic remission under PEG-IFN treatment, the malignant myeloproliferative clone remained present. However, Kiladjian et al. very recently reported a molecular response to pegylated interferon α-2a in 24/27 PV patients with levels of the mutant cells decreasing from 49 to 27% (mean); furthermore in one patient mutant JAK2 was no longer detectable at 1 year. 10

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