A 4 base pair TGAT insertion at codon 116 of the beta globin gene caus¬es ,0 - Thal¬assaemia

A new β^0 thalassaemia allele caused by a TGAT insert in codon 116 of exon III was detected in a patient compound heterozygous for β^0 thalassaemia/Hb D Los Angeles and his father. The mutation unexpectedly causes a classical thalassaemic phenotype. The compound heterozygousity leads to mild microcytic anemia and no further clinical signs.

Haematologica 2005; 90:(6)e59-e60

Sir,

We report a new, thalassaemia mutation detected in a patient compound heterozygous for Hb D $_{\rm Los\ Angeles}$ and β^0 thalassaemia. A 3 - year old boy was referred to our hematology unit for evaluation of a microcytic anemia of unclear origin. Initial routine laboratory studies revealed a Hb of 96 g/L with a normal reticulocyte count and absence of red cell inclusions (i.e. Heinz bodies). On clinical examination no significant abnormalities were found except for a slight pallor despite the dark skin colorit. The hematological parameters of the propositus, his parents and one sister are reported in Table 1.

Table 1. Hematological parameters of propositus and his family (ND: not determined).

	Hb(g/L)	Ec(x1012/L) MCV(fL)		MCH(pg) Retics(×10º/L)	
Propositus	96	5.31	63	18	122
Father	121	5.76	68	21	ND
Mother	120	4.29	77	28	ND
Sister	125	4.31	81	29	ND

Poly CAT A cation exchange HPLC¹ showed only a peak in position of Hb D $_{\text{Los Angeles}}$ (96.5%), 3.0% Hb A² and 0.5% Hb F. No normal Hb A could be detected. Additionally, in isoelectric focussing² no normal Hb A was detectable (*data not shown*). The father of the propositus was a carrier of β thalassaemia (Hb A² 5.3% Hb F 0.4%) and the mother was heterozygous for Hb D $_{\text{Los Angeles}}$ (Hb A² 2.5%, Hb D 46.0%).

To determine the nature of the corresponding thalassaemia mutation and to confirm the nature of the structural variant we analyzed the β globin genomic DNA. DNA extraction and PCR were done using standard methods. A 527 bp long PCR product containing exon III of the , globin gene generated with primers 5'-GGGT-TAAGGČAATAĞCAĂT -3' (located at +1139 to 1159 relative to the Cap site) and 5'-CACTGAGACCTCCCA-CATTCCC-3' (position 1666 to 1647) was first analyzed by non-radioactive SSCP analysis.³ Strong heteroduplex and single strand signals indicated the presence of a deletion / insertion. This was confirmed by direct sequencing using an ABI310 Prism sequencer and Big Dye chemistry (Applied Biosystems, Foster City, CA, USA). The overlapping signal started at the second base of codon 116. The PCR product was electrophoresed on agarose and the corresponding band eluted using the QIAquick Gel Extraction Kit (Qiagen, Basle, Switzerland). The fragment was denatured for 5 min at 95°C and collected on ice. Ligation into pCR - TOPO vector and transformation were performed using the TOPO TA Cloning Kit (In vitro gen, Groningen, The Netherlands) according to the protocol of the manufacturer. Direct forward and reversed

Figure 1. Forward sequence of cloned PCR product using forward PCR primer as sequencing primer. Wildtyp sequence is shown as shaded area. The TGAT insert between nt 1379 and 1380 is shown on white background; The wild type positions are given relative to the cap site.

sequencing of cloned products show¬ed a TGAT insert between nt 1379 and 1380. The result of forward sequencing is shown in Figure 2. The Hb D $_{\text{Los Angeles}}$ mutation at codon 121, GAA→CAA, was confirmed by sequencing of the corresponding PCR product of the mother and by reversed sequencing of the double stranded PCR product of the propositus.

The 4 bp TGAT insert in Codon 116 leads to a premature stop at wild type codon 138/139. The father of the propositus shows the classical picture of a β^0 thalassaemia carrier with no signs of hemolysis or reticulocytosis, there was no sign of a mutant hemoglobin in peripheral blood. The frameshift does not produce, like many other frameshift or nonsense mutations in exon III, a more severe form of thalassaemia⁴ or even a dominant type.⁵ Whether this mild phenotype is caused by a transcript sensitive to nonsense mediated mRNA decay⁶ and/or yet unclear mechanisms as described for other exon III nonsense⁷ or frameshift⁸ mutations remains unclear.

On re-examination of the compound heterozygous patient at the age of 11 years he again showed a borderline anaemia with a Hb of 115–120 g/L and a pronounced microcytosis but no abnormal clinical findings. This confirms several previous reports about the relatively benign nature of the compound heterozygous state of Hb D_{Los} Angeles and classical β^0 thalassaemia.^{9,10} Unfortunately, since the family members are Tamil refugees residing in Switzerland, we were unable to study the patient's grandparents in order to determine whether this mutation is a *de novo* insertion or inherited.

Hannes Frischknecht,[†] Roland Kiewitz,³ Markus Schmugge² [†]Institute for medical & molecular Diagnostics, Rautistr. 13, 8047 Zürich, 2Division of Haematology and 3 Division of Clinical Chemistry, University Children's Hospital, 8032 Zürich, Switzerland

Acknowledgment: We thank Dr. H.G. Oprecht, MD, for co-operation and providing blood samples. Correspondence: Hannes Frischknecht, IMD Institute for Medical & Molecular Diagnostics Rautistr. 13, CH-8047 Zürich, Switzerland. Tel: *41 -1 - 497 30 60 Fax: *41-1-4973070 E-mail: hannes@imdlab.ch Key words: HbD / β° thalassaemia, 4 bp insert

References

- 1. Bissé E, and Wieland H. High-performance liquid chromatographic separation of human haemoglobins. Simultaneous quantitation of foetal and glycated haemoglobins. J Chromatog 1988; 434: 95-110.Basset P, Beuzard Y, Garel MC, and Rosa J. Isoelectric focusing
- of human hemoglobin: its application to screening, to the characterization of 70 variants, and to the study of modified fractions of normal hemoglobins. Blood 1978; 51: 971-82. Liechti-Gallati S, Schneider V, Neeser D, and Kraemer R. Two buffer PAGE system-based SSCP/HD analysis: a general proto-
- col for rapid and sensitive mutation screening in cystic fibrosis and any other human genetic disease. Eur J Hum Genet 1999; 7:590-98.
- Williamson D, Brown K, Langdown JV, Baglin TP. Mild thalas-saemia intermedia resulting from a new insertion / frameshift mutation in the beta gene. Hemoglobin 1997; 21:485-93. Thein SL. Dominant β thalassaemia: Molecular basis and 4.
- 5.

pathophysiology. Br J Haematol. 1992; 80:273-7.

- Danckwardt S, Neu-Yilik G, Thermann R, Frede U, Hentze MW, Kulozik AE. Abnormally spliced beta-globin mRNAs: a 6 single point mutation generates transcripts sensitive and insensitive to nonsense-mediated mRNA decay. 2002;99:1811-6. Blood
- 7. Divoky V, Gu LH, Indrak K, Mocikova K, Zarnovicanova M, HuismanTHJ. A new beta zero-thalassaemia nonsense muta-tion (codon 112, $T \rightarrow A$) not associated with a dominant type of thalassaemia in the heterozygote. Br J Haematol 1993;83:523-4.
- Hopmeier P, Krugluger W, Gu LH, Smetanina NS, Huisman THJ. A newly discovered frameshift at codons 120-121 (+A) of 8 the beta gene is not associated with a dominant form of beta-
- 9.
- the beta gene is not associated with a dominant form of beta-thalassemia. Blood 1996;87:5393-4. Dawood ST, Abulaban M, Kamel K, HuismanTHJ. Hemoglobin D β^0 thalassemia: A case report and family study. Am J Pediatr. Hematol Oncol 1988;10: 316-8. Adekile AD, Kazanetz EG, Leonova JY, Marouf R, Khmis A, Huisman THJ. Co-inheritance of Hb D Punjab and β^0 Thalassemia (IVS-II-1, G \rightarrow A). Am J Pediatr. Hematol Oncol 1996:18:151-3. 10 1996;18:151-3.