et al. A comparison of the safety and efficacy of oral anticoagulation for the treatment of venous thromboembolic disease in patients with or without malignancy. Thromb Haemost 2000;84:805-10.

- Piovélla F, Crippa L, Barone M, Vigano D'Angelo S, Serafini S, Galli L, et al. Normalization rates of compression ultrasonography in patients with a first episode of deep vein thrombosis of the lower limbs: association with DVT recurrence and new thrombosis. Haematologica 2002; 87:515-22.
- thrombosis. Haematologica 2002; 87:515-22.
   Prandoni P, Lensing AW, Prins MH, Bernardi E, Marchiori A, Bagatella Pet al. Residual vein thrombosis as a predictive factor of recurrent venous thromboembolism. Ann Intern Med 2002;137:955-60.
- Palareti G, Legnani C, Cosmi B, Guazzaloca G, Pancani C, Coccheri S. Risk of venous thromboembolic recurrence: high negative predictive value of D-dimer performed after oral anticoagulation is stopped. Thromb Haemost 2002;87:7.
- Palareti G, Legnani C, Cosmi B, Valdre L, Lunghi B, Bernardi F, et al. Predictive value of D-dimer test for recurrent venous thromboembolism after anticoagulation withdrawal in subjects with a previous idiopathic event and in carriers of congenital thrombophilia. Circulation 2003;108:313-8.
   Eichinger S, Minar E, Bialonczyk C, Hirschl M, Ouehenberger
- Eichinger S, Minar E, Bialonczyk C, Hirschl M, Quehenberger P, Schneider B, et al. D-dimer levels and risk of recurrent venous thromboembolism. JAMA 2003;290:1071-4.
- 9. Prandoni P, Cogo A, Bernardi E, Villalta S, Polistena P, Simioni P, et al. A simple ultrasound approach for detection of recurrent proximal-vein thrombosis. Circulation 1993;88:1730-5.
- Fedullo PF, Tapson VF. The evaluation of suspected pulmonary embolism. N Engl J Med 2003;349:1247-56.

## Stem Cell Transplantation

A direct comparison of immunological characteristics of granulocyte colony-stimulating factor (G-CSF)-primed bone marrow grafts and G-CSF-mobilized peripheral blood grafts

Our preliminary results suggest the existence of quantitative and qualitative differences in immune cells and type1 and type2 cytokines between granulocyte colony-stimulating factor (G-CSF) primed bone marrow (G-BM) and G-CSF-mobilized peripheral blood grafts (G-PB). Our findings suggest that lower T-cell hyporesponsiveness and easier polarization of T cells from Th1 to Th2 are found in G-BM.

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haernatologica 2005; 90:715-716
http://www.haernatologica.org/journal/2005/5/715.html)
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Preliminary clinical trials have indicated that G-BM results in comparable engraftment, reduced severity of acute graft-versus-host disease (GVHD), and less subsequent chronic GVHD, as compared with G-PB.<sup>1,2</sup> Moreover, G-BM transplantations produce even less chronic GVHD than do steady-state bone marrow grafts.<sup>2</sup> In this study, we report on the immunological cells and the type1/type2 cytokine profile of lymphocytes present in G-PB and G-BM harvests.

The donors, consisting of eight men and seven women, provided informed consent and ranged in age from 13 to 65 years, with a median age of 40 years. Approval for this study was obtained from the Institutional Review Board and Ethical Committee of the Health Center at Peking University. Samples of G-BM and G-PB were obtained, isolated, and evaluated as described previously.<sup>34</sup> It was ensured that the G-BM and G-PB had the same cell concentration. Statistical comparisons were performed using t-tests for independent samples. The lymphocyte proliferation ability in G-PB (stimulation index: 1.13±0.24) was significantly higher than in G-BM (0.98±0.14, p=0.045; n=15 experiments). This finding suggests hyporespon-



Figure 1. The quantities of type 1 (IFN- $\gamma$ ) and type 2 (IL-4) cytokines secreted by lymphocytes per microliter between G-PB and G-BM, the formula for the calculation of the cytokines a follows: quantities of cytokines=secretion of cytokines (pg/10° MNCs)×cell counts (MNCs/ $\mu$ L). \**p*<0.001, independent t-tests was used (G-PB vs G-BM).

siveness of T cells in G-BM and is likely related to the lower incidence of GVHD observed in G-BM transplants.

The quantities of interferon- $\gamma$  (IFN- $\gamma$ ) (13.19±14.33 pg) and interleukin-4 (IL-4) (3.67±1.77 pg) secreted per microliter of G-PB mononuclear cells were, respectively, 8.5- and 4.5-fold higher than those of G-BM mononuclear cells (1.31±0.57 pg and 0.75±0.24 pg; p<0.001) (Figure 1). The ratio of IL-4/IFN-γ was significantly lower in G-PB than in G-BM (0.33±0.23 vs. 0.73±0.16, p<0.001). These results suggest that bone marrow T cells could be easily polarized from Th1 to Th2 and that patients transplanted with G-PB could accept more type1/type 2 cytokines than could patients transplanted with G-BM. The type-1 to type-2 immune deviation after in vivo application of G-CSF is associated with decreased acute GVHD or with the development of a chronic GVHD syndrome, characterized by decreased mortality and autoantibody formation.5

Krenger and Ferrara have proposed a model in which type 1 cytokines (IL-2, IFN- $\gamma$ ) are involved in the physiopathology of acute GVHD, and type 2 cytokines (IL-4, IL-10) play a crucial role in the physiopathology of chronic GVHD.<sup>5</sup> Fowler *et al.* found that type 1 and type 2 cells appear to play different roles in mediating GVHD and graft-versus-leukemia (GVL) effects.<sup>6,7</sup> Furthermore, type 2 T cells are more resistant to CD95 (Fas)-dependent activation-induced cell death than are type 1 T cells.<sup>8</sup> Therefore, the high quantities of type 1 and type 2 cytokines in G-PB may be related to the different outcomes of the GVL effect and GVHD after G-PB and G-BM transplantation.<sup>1,2</sup>

The quantities of nucleated cells and monocytes in G-PB were, respectively, 4- and 43-fold higher than in G-BM harvests (p<0.001), all lymphocyte subsets exhibited 26- to 46-fold higher cell counts (p<0.001), and the CD4/CD8 ratio was also significantly higher in G-PB than in G-BM (1.59±0.53 vs. 0.91±0.29, p<0.001). These findings indicate that patients transplanted with G-PB may accept more T cells and monocytes than patients transplanted with G-BM (*clinical data not shown*). The cell counts of dendritic cell (DC) 1 and DC2 subgroups in G-PB were, respectively, 11-and 7-fold higher than those in G-BM (p<0.001 and



Figure 2. Expression of CD28 co-stimulatory molecules on T cell subgroups (CD3, CD4, CD8 positive cells). Percentage (A), mean fluorescence intensity (MFI) (B) and overall expression (C) on T cells of G-PB (dark boxes) and G-BM (light boxes) treatment with rhG-CSF ( $5\mu g/kg/d$ ). \*p< 0.05, \*\*p< 0.01

p=0.001; respectively); however, the DC2:T lymphocyte ratio was significantly lower in G-PB (0.91±0.77%) than in G-BM (3.02±2.65%) (p=0.006). The percentage of CD28 expression in CD3<sup>+</sup>, CD4<sup>+</sup>, and CD8<sup>+</sup> T cells was lower in G-BM than in G-PB (p=0.005, 0.014, and 0.012, respectively) (Figure 2A), as was the overall expression of CD28 in CD4<sup>+</sup> and CD8<sup>+</sup> T cells (p=0.031 and 0.016, respectively) (Figure 2C). Some researchers have suggested that a modulation of T-cells from a Th1 to Th2 phenotype in G-CSF-treated bone marrow or peripheral blood harvests may be involved in the selective increase of type 2 DC and monocytes and the downregulation of CD28/B7 costimulatory signals.<sup>3,4,9</sup>

Hardling *et al.* have shown that T-cells anergy can be induced if the CD28/B7 co-stimulatory path is blocked;<sup>10</sup> therefore, it should be investigated whether the lower DC2:T lymphocyte ratio in G-PB and the lower expression of CD28 in G-BM CD4<sup>+</sup> and CD8<sup>+</sup> T cells contribute to the difference in polarization from Th1 to Th2 and T cell hyporesponsiveness between G-PB and G-BM.

In conclusion, we showed that there are both quantitative and qualitative differences of immunological cells and type 1 and type 2 cytokines between G-PB and G-BM. The correlation between our experimental results and the occurrence and severity of acute GVHD and chronic GVHD, relapse, or other transplant-related complications, are presently being elucidated.

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Funding: this work was supported by the National Natural Science Foundation of China (grant no. 30370591) and the Peking University "211" Foundation. We would like to thank Dr. Paul Kretchmer (kretchmer@sfedit.net) at San Francisco Edit for his assistance in editing this manuscript.

Key words: bone marrow grafis, G-CSF, peripheral blood stem cell grafis, T cells, dendritic cells, costimulatory molecules.

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## References

- Morton J, Hutchins H, Durrant S. Granulocyte-colony-stimulating factor (G-CSF)-primed allogeneic bone marrow:significiantly less graft-versus-host disease and comparable engraftment to G-CSF-mobilized peripheral blood stem cells. Blood 2001;98:3186-91.
- Elfenbein GJ, Sackatein R. Primed marrow for autologous and allogeneic transplantation: a review comparing primed marrow to mobilized blood and steady-state marrow. Bone Marrow Transplant 2004;32:327-39.
- Chen SH, Li X, Huang XJ. Effect of recombinant human granulocyte colony-stimulating factor on T-lymphocyte function and the mechanism of this effect. Int J Hematol 2004;79:178-84.
- Huang XJ, Chang YJ, Zhao XY. In vivo induction of T cell hyporesponsiveness and alteration of immunological cells of bone marrow grafts using a granulocyte colony-stimulating factor. Haematologica 2004;89:1517-24.
- Krenger W, Ferrara JLM. Graft-versus-host disease and the Th1/Th2 paradigm. Immuno Res 1996;15:50-73.
- Fowler DH, Breglio J, Nagel G, Hirose C, Gress RE. Allospecific CD4+, Th1/Th2 and CD8+,Tc1/Tc2 populations in murine GVL: type I cells generate GVL and type II abrogate GVL. Biol Blood Marrow Transplant 1996;3:118-25.
- Jung U, Foley JE, Erdmann AA, Eckhaus MA, Fowler DH. CD3/CD28-costimulated T1 and T2 subsets: differential in vivo allosensitization generates distinct GVL and GVHD effects. Blood 2003;102:3439-46.
- Ramsdell F, Seaman MS, Miller RE, Picha KS, Kennedy MK, Lynch DH. Differential ability of Th1 and Th2 cells to express Fas ligand and to undergo activation-induced cell death. Int Immunol 1994;6:1545-53.
- Klangsinsirikul P, Russell NH. Peripheral blood stem cell harvest from G-CSF-stimulated donors contain a skewed Th2 CD4 phenotype and a predominance of type 2 dendritic cells. Exp Hematol 2002;30:495-501.
- Harding FA, McArthur JG, Gross JA, Raulet DH, Alliaon JP. CD28-mediated signalling co-stimulates murine T cells and prevents induction of anergy in T-cell clones. Nature 1992; 356:607-9.