

family history needs to be done more than once. In addition to anticipation, the cause of death is more often attributed to CLL rather than other associated medical conditions of the aged. Prolymphoid transformation and the occurrence of second primary neoplasms are thought to be more frequent in familial CLL.¹⁴ Recently several groups have detected a B cell monoclonal lymphocytosis (BCML) or expansion in environmental studies, blood bank donors and aging individuals.¹⁷⁻²⁰ and BCML as a precursor state can be used as a surrogate marker in first degree relatives in familial CLL.^{21,22} Although there are some ethical considerations about what to tell patients who have BCML, this can be problematic when it occurs in an unaffected HLA matched sibling. Regardless, BCML in the setting of familial MCL, CLL or LPD offers not only the potential for early detection but also the opportunity to study early molecular events in the pathogenesis of these disorders.

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References

1. Tort F, Camacho E, Bosch F, Harris NL, Montserrat E, Campo E. Familial lymphoid neoplasms in patients with mantle cell lymphoma. *Haematologica* 2004;89:314-9.
2. Ardashnikov, SN, Genetics of leukemia in man. *J Hyg* 1937;37:286.
3. Videbaek, A. Familial Leukemia. A preliminary report. *Acta Medica Scandinavica* 1947;127:26-52.
4. Gunz, FW, Gunz, JP, Veal, AMO, Chapman, CJ, Houston, IB, Familial leukaemia: a study of 909 families. *Scand J Haematol* 1975; 15:117-31.
5. Fraumeni JF Jr, Vogel CL, DeVita VT. Familial chronic lymphocytic leukemia. *Ann Intern Med* 1969;71:279-84.
6. Fraumeni, JF, Wertelecki W, Blattner WA, Jensen RD, Leventhal BG. Varied manifestations of a familial lymphoproliferative disorder. *Am J Med* 1975;59:145-51.
7. Blattner, WA, Dean, JH, Fraumeni JFJ. Familial lymphoproliferative malignancy: clinical and laboratory follow-up. *Ann Intern Med* 1979;90:943-4.
8. Blattner WA, Strober W, Muchmore AV, Blaese RM, Broder S, Fraumeni JF Jr. Familial chronic lymphocytic leukemia. Immunologic and cellular characterization. *Ann Intern Med* 1987;84:554-7.
9. Cuttner J. Increased incidence of hematologic malignancies in first-degree relatives of patients with chronic lymphocytic leukemia. *Cancer Invest* 1992;10:103-9.
10. Goldgar DE, Easton DF, Cannon-Albright LA, Skolnick MH. Systematic population based assessment of cancer risk in first-degree relatives of cancer probands. *J Natl Cancer Inst* 1994;86:1600-8.
11. Linet MS, Van Natta ML, Brookmeyer R, Khoury MJ, McCaffrey LE, Humphrey RL, Szklo M. Familial cancer history and chronic lymphocytic leukemia: a case control study. *Am J Epidemiol* 1989; 130:655-64.
12. Cartwright RA, Bernard SM, Bird CC, Darwin CM, O'Brien C, Richards IDG, et al. Chronic lymphocytic leukaemia: case control epidemiological study in Yorkshire Br *J Cancer* 1987;56:79-82.
13. Goldin LR, Sgambati M, Marti GE, Fontaine L, Ishibe N, Caporaso N. Anticipation in familial chronic lymphocytic leukemia. *Am J Hum Genet* 1999;65:265-9.
14. Ishibe N, Sgambati MT, Fontaine L, Goldin LR, Jain N, Weissman N, et al. Clinical characteristics of familial B-CLL in the National Cancer Institute Familial Registry. *Leu Lymphoma* 2001;42:99-108.
15. Sakai A, Marti GE, Caporaso N, Pittaluga S, Touchman JW, Fend F,

et al. Analysis of expressed immunoglobulin heavy chain genes in familial B-CLL. *Blood* 2000;95:1413-9.

16. Ishibe N, Goldin LR, Caporaso NE, Sgambati MT, Dean M, Albitar M, et al. ATM mutations and protein expression are not associated with familial B-CLL cases. *Leuk Res* 2003;27:973-5.
17. Rawstron AC, Green MJ, Kuzmicki A, Kennedy B, Fenton JA, Evans PA, et al. Monoclonal B lymphocytes with the characteristics of "indolent" chronic lymphocytic leukemia are present in 3.5% of adults with normal blood counts. *Blood* 2002;100:635-9.
18. Ghia P, Prato G, Scielzo C, Stella S, Geuna M, Guida G, et al. Monoclonal CD5⁺ and CD5⁻ B lymphocyte expansions are frequent in the peripheral blood of the elderly. *Blood* 2003 (in press).
19. Determining the role of environmental exposures as risk factors for B-cell lymphoproliferative disorders. Marti GE, Vogt RF, Zenger VE, editors. Proceedings of a US Public Health Service Workshop; 1995 Jun 14-15; Atlanta. Washington DC: US Government Printing Office; 1997b. p. 173-80.
20. Rachel JM, Zucker ML, Plapp FV, Fox CM, Marti GE, Abbasi F, et al. B cell monoclonal lymphocytosis in blood donors. *Blood* 2002; 100:590a[abstract].
21. Rawstron AC, Yuille MR, Fuller J, Cullen M, Kennedy B, Richards SJ, et al. Inherited predisposition to CLL is detectable as sub-clinical monoclonal B-lymphocyte expansion. *Blood* 2002;100:2289-91.
22. Marti GE, Carter P, Abbasi F, Washington GC, Jain N, Zenger VE, et al. B-cell monoclonal lymphocytosis and B-cell abnormalities in the setting of familial B-cell chronic lymphocytic leukemia. *Cytometry* 2003;52B:1-12.

Expansion of cord blood hematopoietic stem cells for clinical application

Umbilical cord blood is a realistic alternative to mobilized peripheral blood or bone marrow transplantation in children.¹ The use of umbilical cord blood for allogeneic transplantation has been hindered in adults by the concern that a single collection may contain insufficient numbers of hematopoietic stem cells. Several papers on cord blood hematopoietic stem cells have appeared in this journal in the last two years.²⁻¹⁰ In this issue Zhai and co-workers¹¹ describe a short-term *ex vivo* expansion protocol that sustains the homing-related properties of umbilical cord blood hematopoietic stem and progenitor cells. In *This Month in Haematologica* section (see page 257), Carmela Calés illustrates the interest of this Chinese study.

References

1. Dini G, Cancedda R, Giorgiani G, Porta F, Messina C, Uderzo C, et al. Unrelated donor marrow transplantation in childhood: a report from the Associazione Italiana Ematologia e Oncologia Pediatrica (AIEOP) and the Gruppo Italiano per il Trapianto Midollo Osseo (GITMO). *Haematologica*. 2002;Suppl 8:51-7.
2. Gaipa G, Coustan-Smith E, Todisco E, Maglia A, Campana D. Characterization of CD34⁺, CD13⁺, CD33⁻ cells, a rare subset of immature human hematopoietic cells. *Haematologica* 2002;87: 347-56.
3. Laurenti L, Perrone MP, Bafti MS, Ferrari F, Screnci M, Pasqua I, Girelli G. HLA typing strategies in a cord blood bank. *Haematologica* 2002;87:851-4.
4. Jung YJ, Woo SY, Ryu KH, Chung WS, Kie JH, Seoh JY. Functional maturation of myeloid cells during in vitro differentiation from human cord blood CD34⁺ cells. *Haematologica* 2002;87:1222-3.
5. Gonelli A, Mirandola P, Grill V, Secchiero P, Zauli G. Human herpesvirus 7 infection impairs the survival/differentiation of megakaryocytic cells. *Haematologica* 2002;87:1223-5.
6. Timeus F, Crescenzo N, Saracco P, Doria A, Fazio L, Albani R, Cordero Di Montezemolo L, Perugini L, Incarbone E. Recovery of

- cord blood hematopoietic progenitors after successive freezing and thawing procedures. *Haematologica* 2003;88:74-9.
7. Bruno S, Gunetti M, Gammaitoni L, Dane A, Cavalloni G, Sanavio F, et al. In vitro and in vivo megakaryocyte differentiation of fresh and ex-vivo expanded cord blood cells: rapid and transient megakaryocyte reconstitution. *Haematologica* 2003;88:379-87.
 8. Encabo A, Mateu E, Carbonell-Uberos F, Minana MD. IL-6 precludes the differentiation induced by IL-3 on expansion of CD34+ cells from cord blood. *Haematologica* 2003;88:388-95.
 9. Sun R, Wei H, Zhang J, Tian Z. The effects of natural killer-cell depletion on ex vivo expansion of hematopoietic progenitor cells from umbilical cord blood. *Haematologica* 2003;88:561-9.
 10. Cacoullos NT, Gritzapis AD, Tsitsilonis AE, Tsiatas ML, Baxevanis CN, Papamichail M. Efficacy of novel culture environments on the ex vivo expansion kinetics of cord blood progenitor cells. *Haematologica* 2002;87:320-1.
 11. Zhai QL, Qiu LG, Li Q, Meng HX, Han JL, Herzig RH, et al. Short-term ex vivo expansion sustains the homing-related properties of umbilical cord blood hematopoietic stem and progenitor cells. *Haematologica* 2004;89:265-272.

International Committee of Medical Journal Editors (ICMJE): Uniform Requirements for Manuscripts Submitted to Biomedical Journals: writing and editing for biomedical publication

A small group of editors of general medical journals met informally in Vancouver, British Columbia, in 1978 to establish guidelines for the format of manuscripts submitted to their journals. The group became known as the Vancouver Group. Its requirements for manuscripts, including formats for bibliographic references developed by the National Library of Medicine, were

first published in 1979. The Vancouver Group expanded and evolved into the International Committee of Medical Journal Editors (ICMJE), which meets annually. The ICMJE gradually has broadened its concerns to include ethical principles related to publication in biomedical journals.

The ICMJE has produced multiple editions of the Uniform Requirements for Manuscripts Submitted to Biomedical Journals. Over the years, issues have arisen that go beyond manuscript preparation, resulting in the development of a number of Separate Statements on editorial policy. The entire Uniform Requirements document was revised in 1997; sections were updated in May 1999 and May 2000. In May 2001, the ICMJE revised the sections related to potential conflict of interest. For the last revision (2003), the committee revised and reorganized the entire document and incorporated the Separate Statements into the text.

The Editorial Board of *Haematologica* invites all authors to visit the ICMJE web site (<http://www.icmje.org/>) and carefully read the following sections before submitting a manuscript:

- a) ethical considerations in the conduct and reporting of research;
- b) publishing and editorial issues related to publication in biomedical journals;
- c) manuscript preparation and submission.