

Correspondence: Gerhard Heil, MD, Ph D, Dept. Hematology and Oncology, Hannover Medical School, Carl-Neuberg-Str. 1 D-30625 Hannover, Germany. Phone: international +49.511.5323610/3720. Fax: international +49.511.5323611. E-mail: heil.gerhard@mh-hannover.de

Manuscript processing

This manuscript was peer-reviewed by two external referees and by Professor Mario Cazzola, Editor-in-Chief. The final decision to accept this paper for publication was taken jointly by Professor Cazzola and the Editors. Manuscript received May 8, 2003; accepted June 30, 2003.

References

1. Downing JR. The AML1-ETO chimaeric transcription factor in acute myeloid leukaemia: biology and clinical significance. *Br J Haematol* 1999;106:296-308.
2. Ogawa E, Inuzuka M, Maruyama M, Satake M, Naito Fujimoto M, Ito Y, et al. Molecular cloning and characterization of PEBP2 β , the heterodimeric partner of a novel *Drosophila* runt-related DNA binding protein PEBP2 alpha. *Virology* 1993;194:314-31.
3. Wang J, Hoshino T, Redner RL, Kajigaya S, Liu JM. ETO, fusion partner in t(8;21) acute myeloid leukemia, represses transcription by interaction with the human N-CoR/mSin3/HDAC1 complex. *Proc Natl Acad Sci USA* 1998;95:10860-5.
4. Lutterbach B, Westendorf JJ, Linggi B, Patten A, Moniwa M, Davie JR, et al. ETO, a target of t(8;21) in acute leukemia, interacts with the N-CoR and mSin3 corepressors. *Mol Cell Biol* 1998;18:7176-84.
5. Zhang DE, Zhang P, Wang ND, Hetherington CJ, Darlington GJ, Tenen DG. Absence of granulocyte colony-stimulating factor signaling and neutrophil development in CCAAT enhancer binding protein α -deficient mice. *Proc Natl Acad Sci USA* 1997;94:569-74.
6. Noti JD. Sp3 mediates transcriptional activation of the leukocyte integrin genes CD11C and CD11B and cooperates with c-Jun to activate CD11C. *J Biol Chem* 1997;272:24038-45.
7. Lopez-Rodriguez C, Corbi AL. PU.1 negatively regulates the CD11c integrin gene promoter through recognition of the major transcriptional start site. *Eur J Immunol* 1997;27:1843-7.
8. Westendorf JJ, Yamamoto CM, Lenny N, Downing JR, Selsted ME, Hiebert SW. The t(8;21) fusion product, AML1-ETO, associates with C/EBP- α , inhibits C/EBP- α -dependent transcription, and blocks granulocytic differentiation. *Mol Cell Biol* 1998;18:322-33.
9. Pabst T, Mueller BU, Harakawa N, Schoch C, Haferlach T, Behre G, et al. AML1-ETO downregulates the granulocytic differentiation factor C/EBP α in t(8;21) myeloid leukemia. *Nat Med* 2001;7:444-51.
10. Krauter J, Wattjes MP, Nagel S, Heidenreich O, Krug U, Kafert S, et al. Real-time RT-PCR for the detection and quantification of AML1/MTG8 fusion transcripts in t(8;21)-positive AML patients. *Br J Haematol* 1999;107:80-5.

Early changes in bone marrow morphology induced by thalidomide in patients with refractory myeloma

Bone marrow morphology and the number of CD34⁺ cells were evaluated in 17 patients with refractory multiple myeloma at the start of therapy with low-dose thalidomide and after 3 months. All responding patients showed an evident increase of cellularity, reappearance of erythroblasts and myeloid precursors in various phases of differentiation, and an increase of megakaryocytes. Nine of the ten responders also had increase of bone marrow CD34⁺ cells.

haematologica 2003; 88:958-960

(http://www.haematologica.org/2003_08/958.htm)

Although high-dose therapy produces high response rates and overall survival in multiple myeloma (MM), recurrence of disease usually develops and options of salvage therapy are limited.^{1,2} Thalidomide has proven to be very effective in MM^{3,4} through several biological pathways.^{5,6} We have previously reported that clinical and hematologic recovery can be observed even in patients who obtain less response.⁷ Moreover, we described the early morphologic changes in two refractory MM patients treated with thalidomide.⁸

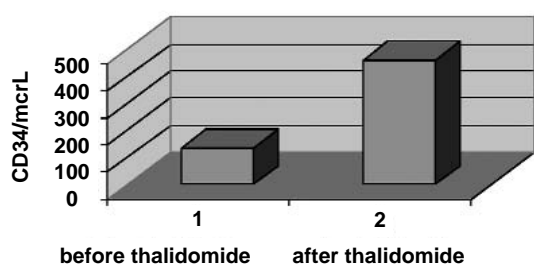
Based on these observations, we performed this study on a cohort of patients with refractory MM treated with thalidomide in order to evaluate the morphologic bone marrow changes and to estimate the variation of bone marrow CD34⁺ cells.

The 17 patients included in the study were refractory to at least 2 previous lines of therapy, which in 6 patients had been one or two autologous bone marrow transplantations. Thalidomide was given at a dose of 100 mg/day and escalated to 200 mg/day in case of resistance. Four patients, because of severe disease-related symptoms, also received dexamethasone 20 mg/day for two days every two weeks for the first two months. Response to therapy was assessed according to the criteria of the European Bone Marrow Transplantation Group.⁹ A minimum of three months of uninterrupted therapy was necessary to evaluate the response. The clinical and laboratory characteristics of the 17 evaluable patients recorded at the start of thalidomide were: median age 55 (37-67) years; 13 IgG, 2 IgA, 2 Bence Jones; 1 stage I, 4 stage II, 12 stage III. The evaluation of the bone marrow morphology was performed on May-Grünwald Giemsa smears before and after three months of thalidomide. Bone marrow smears were evaluated by three independent investigators, and by an external morphologist. Percentages of erythroblasts, myeloid precursors, and plasma cells were calculated on a minimum of 500 cells. Cytometric evaluation of bone marrow CD34⁺ cells was performed as previously reported.¹⁰

Thalidomide was well tolerated and side effects were always mild. After three months of thalidomide, 10 out of 17 patients (59%) had achieved a response (6 partial, 1 minor, 3 stable disease), and 7 (41%) had progressed. Table 1 describes the morphologic changes in detail. As shown, an increase of bone marrow cellularity was evident in 8 of the 10 responders. A striking increase in megakaryocyte number accompanied by reappearance of myeloid precursors, very rare in pre-therapy smears, at various stages of differentiation was observed in all responding patients. The percentage of erythroblasts was significantly higher after therapy, increasing from a mean value of 3.21% to 10.14% ($p < 0.0001$). An increase of the bone marrow eosinophils was also noted. The bone marrow CD34⁺ cells, evaluated as absolute numbers and percentages, showed a parallel increase in 9 of 10 responding patients (Figure 1), from a mean of $134.4 \pm 194/\text{mcrL}$ (0.6%) before starting therapy to $462 \pm 335/\text{mcrL}$ (1.8%) after thalidomide ($p = 0.02$ and $p = 0.007$, respectively). By contrast, in non-responding patients bone marrow cellularity very often appeared reduced. Similarly, in these

Table 1. Bone marrow morphology and CD34⁺ count before and after three months of thalidomide in 10 responding and 7 non-responding (*) MM patients.

Patient	Months	Cellularity	Erythroblasts	Megakaryocyte	BMPC	Lymphocyte	CD34/ μ L	CD34%
MC	0	low	6.0	rare	66.5	4.5	16	0.4
	3	normal	23.5	normal	17.5	9.5	540	1.0
LF	0	normal	11.5	rare	51.0	4.5	189	1.0
	3	low	55.0	increased	3.5	8.5	61	0.6
RF	0	scarce	15.0	rare	10.0	14.0	75	0.5
	3	normal	46.5	increased	10.0	6.0	83	3.3
AR	0	low	30.0	normal	28.5	14.0	12	2.0
	3	normal	49.0	increased	5.0	16.5	450	2.5
BG	0	scarce	1.0	rare	72.0	14.0	45	0.6
	3	high	24.5	normal	9.5	4.0	201	1.0
GA	0	normal	12.0	rare	66.5	4.5	480	0.8
	3	high	55.0	increased	11.5	7.5	598	2.3
LA	0	low	22.0	normal	23.5	9.0	496	0.6
	3	normal	57.0	normal	12.5	5.5	735	1.0
PP	0	high	7.0	rare	58.5	15.5	18	0.3
	3	normal	51.0	decreased	7.0	12.5	450	3.0
TD	0	scarce	3.0	rare	22.0	38.0	10	0.1
	3	normal	38.5	increased	3.0	1.0	1185	1.5
TM	0	low	6.0	decreased	29.0	18.0	3	0.1
	3	normal	46.5	normal	7.5	13.5	320	2.0
*VP	0	high	63.0	decreased	19.0	4.0	137	0.7
	3	normal	41.0	rare	36.0	17.0	108	1.2
*NE	0	scarce	24.0	rare	45.5	5.5	78	0.1
	3	high	6.0	rare	87.0	1.0	96	0.4
*CM	0	normal	31.5	normal	39.0	6.5	290	1.2
	3	scarce	7.0	rare	37.0	27.0	74	0.8
*LC	0	normal	32.5	normal	15.5	1.5	357	0.8
	3	low	40.5	rare	12.0	8.0	325	5.5
*FG	0	low	22.0	rare	27.5	10.0	162	4.0
	3	scarce	23.0	rare	21.0	30.0	5	0.2
*MF	0	high	9.0	normal	74.5	5.5	72	0.6
	3	normal	2.0	rare	62.0	12.0	13	0.2
*GR	0	low	2.0	rare	63.0	15.0	33	0.3
	3	high	2.0	rare	92.0	3.0	21	0.3

**Figure 1. Mean values of bone marrow CD34⁺ cells before and after thalidomide treatment in 10 responding patients.**

patients the number of bone marrow CD34⁺ cells appeared to decrease, although not significantly, from a mean value of 161±120/ μ L (0,75%) before thalidomide treatment to 97±123/ μ L (0,40%) after (p =NS).

We report the first study on the bone marrow morphologic changes in patients with refractory MM treated with thalidomide. Patients with refractory myeloma are difficult to treat because of the resistance to therapy acquired by plasma cells and the fragility of the patients. The massive bone marrow plasmacytosis, often present in this phase, affects the normal hematopoietic compartment causing peripheral blood cytopenias. Conventional chemotherapy can reduce the percentage of plasma cells but also damages the normal hematopoietic compartment.

In this study, before starting thalidomide all patients showed different grades of hypocellularity with a prevalence of plasma cells and plasmablasts, and a marked reduction or absence of

megakaryocytes. After three months of thalidomide therapy, an increase in bone marrow cellularity was found in all responding patients, with reappearance of three-lineage precursors in different phases of maturation. A significant increase of bone marrow CD34⁺ cells was recorded. These changes produced an amelioration of the peripheral cytopenias and in four patients eliminated the need for transfusions. In contrast, non-responding patients did not show any morphologic changes.

In conclusion, in myeloma patients, response to thalidomide is characterized by an early increase of bone marrow cellularity with an expansion of hematopoietic stem cells. This could be related to an inhibition of the neoplastic clone and, with respect to chemotherapy, to less damage to normal hematopoietic stem cells.

Alessandro Corso, Angela Lorenzi, Patrizia Zappasodi, Rosangela Invernizzi, Laura Vanelli, Mario Lazzarino
 Division of Hematology, *Internal Medicine and Medical Oncology, IRCCS Policlinico S. Matteo, University of Pavia, Pavia, Italy

Key words: thalidomide, multiple myeloma, refractory disease, morphology, CD34⁺ cells.

*Correspondence: Alessandro Corso, MD, Division of Hematology, Policlinico San Matteo, 27100, Pavia, Italy.
 Fax: international +39.0382.502250.
 E-mail: a.corso@smatteo.pv.it*

Manuscript processing

This manuscript was peer-reviewed by two external referees and by Dr. Juan Bladé, who acted as an Associate Editor. The final decision to accept this paper for publication was taken jointly by Dr. Bladé and the Editors. Manuscript received April 29, 2003; accepted June 16, 2003.

References

1. Attal M, Harousseau JL, Stoppa AM, Sotto JJ, Fuzibet JG, Rossi JF, et al. A prospective, randomized trial of autologous bone marrow transplantation and chemotherapy in multiple myeloma. Intergroup Français du Myelome. *N Engl J Med* 1996;335:91-7.
2. Barlogie B, Jagannath S, Desikan KR, Mattox S, Vesole D, Siegel D, et al. Total therapy with tandem transplants for newly diagnosed multiple myeloma. *Blood* 1999;93:55-65.
3. Singhal S, Mehta J, Desikan R. Antitumor activity of thalidomide in refractory multiple myeloma. *N Engl J Med* 1999;341:1565-71.
4. Agha IY, Moreau P, Leyvraz S, Berthou C, Payen C, Dumontet C, et al. Thalidomide in patients with advanced multiple myeloma. *Hematology J* 2000;1:186-9.
5. Hideshima T, Chauhan D, Shima Y, Raje N, Davies FE, Tai YT, et al. Thalidomide and its analogs overcome drug resistance of human multiple myeloma cells to conventional therapy. *Blood* 2000;96:2943-50.
6. Neben K, Moehler T, Kraemer A, Benner A, Egere G, Ho AD, et al. Response to thalidomide in progressive multiple myeloma is not mediated by inhibition of angiogenic cytokine secretion. *Br J Haematol* 2001;115:605-8.
7. Corso A, Lorenzi A, Orlandi E, Astori C, Mangiacavalli S, Lazzarino M. Advantage of using thalidomide for the management of patients with refractory myeloma. *Haematologica* 2002;87:327-8.
8. Zappasodi P, Lorenzi A, Corso C. Thalidomide in refractory myeloma patients: early changes in bone marrow cellularity. *Haematologica* 200;86:448.
9. Bladé J, Samson D, Reece D, Apperley J, Björkstrand B, Gahrton C, et al. Criteria for evaluation disease response and progression in patients with multiple myeloma treated by high dose therapy and haemopoietic stem cell transplantation. *Br J Haematol* 1998;102:1115-23.
10. Corso A, Arcaini L, Caberlon S, Zappasodi P, Mangiacavalli S, Lorenzi A, et al. A combination of dexamethasone, cyclophosphamide, etoposide, and cisplatin (DCEP) is less toxic and more effective than high-dose cyclophosphamide for peripheral stem cell transplantation in multiple myeloma. *Haematologica* 2002;87:1041-5.