cations due to iron overload and on desferrioxamine therapy was not increased among those with *TFR2* polymorphisms. Our results show that the *TFR2* polymorphisms, I238M and IVS16 +251 CA deletion, while prevalent in Chinese patients, do not influence the degree of iron loading in transfusion-independent β -thalassemia intermedia. These *TFR2* polymorphisms are therefore not useful in explaining the severe iron overload that may be encountered in our patients. This agrees with findings on I238M polymorphism in normal Asian subjects.¹⁰ Furthermore, detection of common *HFE* polymorphisms is also not expected to be fruitful, given the low prevalence of these in our area. Nevertheless, the presence of other, hither-to unidentified genetic determinant(s) of iron overload in the Chinese population cannot be excluded and may need to be unraveled in the future.

Edmond S. K. Ma,* Karen K. Y. Lam,* Amy Y. Y. Chan,* Shau-Yin Ha,° Wing-Yan Au,# Li-Chong Chan*

Departments of Pathology, Pediatrics and Medicine, The University of Hong Kong, Queen Mary Hospital, Pokfulam Road, Hong Kong

Correspondence: Dr. Edmond S. K. Ma, Division of Hematology, Department of Pathology, Queen Mary Hospital, 102 Pokfulam Road, Hong Kong. Phone: international +852.285 4570. Fax: international +852. 28177565. E-mail: eskma@hkucc.hku.hk

Funding: this study was supported by the Children's Thalassemia Foundation and Research Grants Council (HKU 7323/02M). The authors thank Dr. Richard Pang, Division of Clinical Biochemistry, Department of Pathology, Queen Mary Hospital, for performing the iron profile assays.

Key words: TFR2 polymorphism, iron overload, β -thalassemia intermedia, genetic hemochromatosis.

Manuscript processing

This manuscript was peer-reviewed by two external referees and by Professor Mario Cazzola, Editor-in-Chief. The final decision to accept this paper for publication was taken jointly by Professor Cazzola and the Editors. Manuscript received November 20, 2002; accepted February 6, 2003.

References

- Piperno A, Mariani R, Arosio C, Vergani A, Bosio S, Fargion S, et al. Haemochromatosis in patients with β-thalassaemia trait. Br J Haematol 2000;111:908–14.
- Melis MA, Cau M, Deidda F, Barella S, Cao A, Galanello R. H63D mutation in the HFE gene increases iron overload in β-thalassemia carriers. Haematologica 2002;87: 242–5.
- Rees DC, Luo LY, Thein SL, Singh BM, Wickramasinghe S. Nontransfusional iron overload in thalassemia: association with hereditary hemochromatosis. Blood 1998;90: 3234–6.
- Cappellini MD, Fargion SR, Sampietro M, Graziadei G, Fiorelli G. Nontransfusional iron overload in thalassemia intermedia: role of the hemochromatosis allele. Blood 1998;92:4479-89.
- Borgna-Pignatti C, Solinas A, Bombieri C, Micciolo R, Gamberini MR, De Stefano P, et al. The haemochromatosis mutations do not modify the clinical picture of thalassaemia major in patients regularly transfused and chelated. Br J Haematol 1998;103:813–6.
- Merryweather-Clarke AT, Pointon JJ, Shearman JD, Robson KJH. Global prevalence of putative haemochromatosis mutations. J Med Genet 1997;34:275–8.
- Mura C, Raguenes O, Férec C. HFE mutations analysis in 711 hemochromatosis probands: evidence for S65C implication in mild form of hemochromatosis. Blood 1999;93:2502–5.
- 8. Kawabata H, Yang R, Hirama T, Vuong PT, Kawano S,

Gombart AF, et al. Molecular cloning of transferrin receptor 2 – a new member of the transferrin receptor-like family. J Biol Chem 1999;274:20826–32.

- Camaschella C, Roetto A, Cali A, De Gobbi M, Garozzo G, Carella M, et al. The gene TFR2 is mutated in a new type of haemochromatosis mapping to 7q22. Nat Genet 2000;25:14–5.
- Lee PL, Halloran C, West C, Beutler E. Mutational analysis of the transferrin receptor-2 gene in patients with iron overload. Blood Cells Mol Dis 2001;27:285–9.

Molecular characterization of glucose-6-phosphate dehydrogenase deficiency in the Fars province of Iran

We investigated 78 glucose-6-phosphate dehydrogenase (G6PD)-deficient alleles from the Fars province of Iran by polymerase chain reaction-single strand conformation polymorphism (PCR-SSCP) and direct sequencing. The frequency of G6PD Mediterranean in Fars was 84.6%, G6PD Chatham was found to be highly polymorphic and two other sporadic variants (G6PD A- and G6PD Canton) were detected in single cases.

Haematologica 2003;88:346-347 (http://www.haematologica.org/2003_3/88346.htm)

Glucose-6-phosphate dehydrogenase (G6PD) deficiency is the most common enzyme disorder in humans and is characterized by considerable biochemical and molecular heterogeneity.1 The prevalence of G6PD deficiency in the Middle East varies greatly, ranging from 1% among Egyptians to 11.55% among Iranians.^{2,3} G6PD Mediterranean (563 $\dot{C} \rightarrow T$) mutation is probably the most common G6PD variant in the world; it has been widely reported in Europe but also in the Middle-East and in neighboring countries not bordering the Mediterranean sea.4.5 Among the known variants, the relative frequency of this mutation ranges from 70% among Egyptians to 97% for Kurdish Jews.⁶ A recent study carried out on the population of the Mazandaran state of North Iran near the Caspian sea showed a frequency for the G6PD Mediterranean mutation of 66.2% and the presence of two oth-er polymorphic mutations: G6PD Chatham^{1003A} (27%) and G6PD Cosenza^{1376C} (6.7%).⁷ We report here a study performed on 78 G6PD-deficient alleles from a different population of Iran, originating from the Fars province, located in the South of the country. The incidence of G6PD deficiency in this area is estimated to be about 12% in males and 0.9% in females.3

The study was carried out on 74 unrelated G6PD deficient subjects (66 males, 8 females) aged between 10 days to 20 years (mean 8 ± 5 years) all originating from the Fars province of Iran. The subjects were recruited from neonatal and school screening. The diagnosis of G6PD deficiency was based on the fluorescent spot test. Clinical data were recorded considering neonatal jaundice, favism or drug-related hemolysis.

As preliminary screening, the following polymorphic G6PD molecular variants were tested by polymerase chain reactionrestriction fragment length polymorphism (PCR-RFLP): G6PD Mediterranean⁵⁶³¹, G6PD A_^{3766/202A}, G6PD Seattle^{844C}, G6PD Aures^{143C} and G6PD Santamaria^{3766/542T}.

The G6PD Mediterranean mutation was detected in 62/74 (83.8%) samples. Four females were homozygous for this mutation, leading to an overall allele frequency of 84.6%. Among the other variants screened for, we identified one subject with G6PD A- (1.3%) whereas G6PD Seattle, G6PD Aures and G6PD Santamaria were absent from all our samples. The 11 negative samples were submitted to SSCP analysis of the entire G6PD coding region that allowed us to identify two different abnormal patterns in exon 9 and 12, respectively. Nucleotide sequencing of exon 9 revealed a G to A substitution at nt 1003 responsible for
 Table 1. Frequencies of G6PD mutations in the Fars

 province of Iran.

Molecular variant	Cases		Alleles	
	Ν.	(%)	Ν.	(%)
Mediterranean (563 C \rightarrow T)	62	83.78	66	84.62
Chatham (1003 G \rightarrow A)	10	13.51	10	12.82
A- (202 G→A/376 A→G) Canton (1376 G→T)	1 1	1.35 1.35	1 1	1.28 1.28
Total	74	100	78	100

Table 2. Clinical data of the 74 G6PD-deficient subjects.

Clinical manifestation	Total	Med	Chatham	A-	Canton
Favism	33	28/33 (84.9%)	4/33 (12.1%)	1/33 (3.0%)	0/33
Acute hemolytic anemia	35	29/35 (82.9%)	5/35 (14.3%)	1/35 (2.8%)	0/33
Neonatal jaundice	24	18/24 (75.0%)	5/24 (20.8%)	0/24	1/24 (4.2%)
Hyperbilirubinemia	41	35/41 (85.4%)	5/41 (12.2%)	1/41 (2.4%)	0/41

G6PD Chatham in 10/74 samples (13.5%). Nucleotide sequencing of exon 12 revealed the substitution C \rightarrow T at position 1376 which is responsible for G6PD Canton variant (allele frequency: 1.3%). All mutations and frequencies are summarized in Table 1. G6PD Chatham was reported for the first time in an Indian boy but it is now recognized as one of the most common variants worldwide, being present in several populations.7-9 The medical records of all the G6PD-deficient subjects showed that neonatal jaundice occurred in 32.4% of the cases (24/74), favism in 44.6% (33/74) and moderate to severe hyperbilirubinemia in 55.4% (41/74). The incidence of clinical manifestations related to the different molecular variants are reported in Table 2. This is the first report on the molecular basis of G6PD deficiency in the Fars province of Iran showing that the allele frequency of G6PD Mediterranean mutation in Fars (84.6%) is similar to that described for other neighboring countries9,10 and for the Mediterranean region, but higher than that observed in Mazandaran (66.2%), located on the South coast of the Caspian sea, in the North of Iran.⁷ Furthermore, the absence of mutations that are polymorphic in North Africa and the Arabian peninsula (G6PD Aures, Seattle and Santamaria)^{8,10} suggests a different origin and spread of G6PD variants in the South of Iran.

The G6PD Chatham mutation (1003 G \rightarrow A) is highly polymorphic in the Fars region (13%), although its frequency does not reach the values recently observed in the Mazandaran province (27%). However, G6PD Mediterranean and Chatham variants are by far the most commonly observed in both areas. It is noteworthy that G6PD Cosenza and Canton, found in two different regions of Iran, affect the same nucleotide position in the G6PD gene, but the origin of the two mutations is likely to be different. In fact, G6PD Cosenza, identified for the first time in Italy, is a known variant already described in the Middle-East and in the Mediterranean probably migrating from Western countries to Iran. By contrast, G6PD Canton is one of the most common variants in the South East of Asia⁹ and is likely to have been spread during the agricultural migration from China.

Mehran Karimi,* Franco Martinez di Montemuros,° Maria Gabriella Danielli,° Shirin Farjadian,^ Abdolreza Afrasiabi,* Gemino Fiorelli,° Maria Domenica Cappellini°

*Hematology Research Center, Department of Pediatrics, Nemazee Hospital, Shiraz, Iran; [^]Cooley's Center, Shiraz University of Medical Sciences, Shiraz, Iran; [°]Centro Anemie Congenite, Ospedale Maggiore Policlinico IRCCS, Dipartimento di Medicina Interna, University of Milan, Italy

Key words: G6PD deficiency, G6PD mutations, Iran, molecular characterization.

Correspondence: Maria Domenica Cappellini, MD, Centro Anemie Congenite, Dipartimento di Medicina Interna, Università di Milano, Ospedale Maggiore Policlinico IRCCS via F. Sforza 35, 20122 Milan, Italy. Phone: international +39.02.5465571. Fax: international +39.02.50320291. E-mail: maria.cappellini@unimi.it

Manuscript processing

This manuscript was peer-reviewed by two external referees and by Professor Mario Cazzola, Editor-in-Chief. The final decision to accept this paper for publication was taken jointly by Professor Cazzola and the Editors. Manuscript received October 28, 2002; accepted January 23, 2003.

References

- Luzzatto L, Metha A, Glucose-6-phosphate dehydrogenase deficiency. In: Scriver CR, Beaudet AL, Sly WS,Valle D, Editors. The metabolic basis of inherited disease, 5th ed. New York. McGraw-Hill, 1995. p. 3367-98.
- Usanga EA, Ameen R. Glucose-6-phosphate dehydrogenase deficiency in Kuwait, Syria, Egypt, Iran, Jordan and Lebanon. Hum Hered 2000;50:158-61.
- Pishva N, Amoozgar H. Hyperbilirubinemia following exchange transfusion with G-6-PD deficient donor blood. Irn J Med Sci 2001;26:143-5.
- Kurdi-Haidar B, Mason PJ, Berrebi A, Ankra-Badu G, al-Ali A, Oppenheim A et al. Origin and spread of the glucose-6phosphate dehydrogenase variant (G6PD-Mediterranean) in the Middle East. Am J Hum Genet 1990;47:1013-9.
- Al-Ali AK, Al-Mustafa ZH, Al-Madan M, Qaw F, Al-Ateeq S. Molecular characterization of glucose-6-phosphate dehydrogenase deficiency in the Eastern province of Saudi Arabia. Clin Chem Lab Med 2002;40:814-6.
- Oppenheim A, Jury CL, Rund D, Vulliamy TJ, Luzzatto L. G6PD Mediterranean accounts for the high prevalence of G6PD deficiency in Kurdish Jews. Hum Genet 1993;91:293-
- Mesbah-Namin SA, Sanati MH, Mowjoodi A, Mason PJ, Vulliamy TJ, Noori-Daloii MR. Three major glucose-6-phosphate dehydrogenase-deficient polymorphic variants identified in Mazandaran state of Iran. Br J Haematol 2002; 117:763-4.
- Daar S, Vulliamy TJ, Kaeda J, Mason PJ, Luzzatto L. Molecular characterization of G6PD deficiency in Oman. Hum Hered 1996;46:172-6.
- Iwai K, Hirono A, Matsuoka H, Kawamoto F, Horie T, Lin K, et al. Distribution of glucose-6-phosphate dehydrogenase mutations in Southeast Asia. Hum Genet 2001;108:445-9.
- Bayoumi RA, Nur E, Kamal MS, Tadayyon M, Mohamed KK, Mahboob BH, et al. Molecular characterization of erythrocyte glucose-6-phosphate dehydrogenase deficiency in Al-Ain District, United Arab Emirates. Hum Hered 1996;46: 136-41.