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editorials and views

Development and overcoming of ATRA resistance in acute promyelocytic leukemia

Despite the fact that virtually 100% of patients with genetically proven, PML/RAR α -positive acute promyelocytic leukemia (APL) are responsive to all-trans retinoic acid (ATRA), this agent is per se unable to eradicate the leukemic clone, such that a status of minimal residual disease remains detectable during remission by conventional RT-PCR assays, and clinical relapse almost invariably occurs in all patients receiving ATRA monotherapy.¹ This evidence provided the rationale for modern treatment approaches with combined ATRA and chemotherapy which currently result in long-lasting molecular remission and potential cure in the majority of patients.²

The phenomenon of ATRA resistance in APL has interested numerous investigators and several mechanisms have been proposed to explain its development, such as: i) reduced plasma concentrations following prolonged administration; ii) enhanced intracellular sequestration by the cellular retinoic acid binding protein (CRABP); iii) absence of ATRAinduced PML/RARa degradation, and iv) emergence of leukemic cells with functional defects of the PML/RARα protein including mutations in the ligand binding domain (LBD).²⁻⁸ Initially described in a cell line subclone,⁵ such mutations, which would abrogate or impair ligand binding to the target receptor, were subsequently identified in fresh blasts collected at the time of clinical relapse from a few ATRA-resistant APL patients.6,7

In this issue of Haematologica, Marasca et al.9 describe two patients with acquired ATRA resistance who, at relapse, showed mutations in the LBD of PML/RAR α , leading in both cases to amino acid substitutions. In one patient, the authors report a previously unrecognized point mutation which would also predict for alteration in the ligand binding capacity, thereby providing a suggestive molecular explanation for the clinical resistance to ATRA. Most importantly, Marasca et al.9 report for the first time the overcoming of ATRA resistance in patients with mutated LBD of PML/RAR α by use of arsenic trioxide. This latter agent was found to induce apoptosis rather than differentiatiation by selectively targeting the same PML/RAR α fusion protein *in vitro* in ATRA-resistant patients' blasts.8 The achievement of remission in these two cases provides further compelling evidence of non-cross resistant effectiveness of this agent, with respect to ATRA, in the treatment of APL.

A number of clinical and biologic issues remain to be addressed by future investigations on ATRA resistance in APL. First, the incidence of PML/RAR α LBD mutation in relapsing patients is presently unknown and too few cases have been characterized to date. Second, although preliminary in vitro studies using site-directed mutagenesis indicate a pathogenetic link between PML/RAR α LBD mutations and altered binding of the ligand,¹⁰ it is presently unclear to what extent these molecular changes would determine clinical resistance to ATRA in vivo. In a substantial proportion of APL patients, disease relapse is in fact still responsive to ATRA and long-term remission is achieved after ATRA reinduction followed by intensive consolidation.² Finally, clinical observations clearly indicate that the likelihood of response to salvage treatment with ATRA is inversely proportional to the duration of first remission and in particular to the time period during which patients have been off ATRA.² This would rather favor the pharmacokinetic theory,³ according to which adequate plasma concentrations might be reinstored after prolonged discontinuation of the drug. Do patients with late relapse harbor PML/RARα LBD mutaions as well? What is the effect of anthracyclines on mutated subclones? Are they as effective as arsenic trioxide? At the biologic level, one intriguing question is whether these alterations are already present at the time of diagnosis or whether, alternatively, they are events acquired during the clinical evolution of the disease. In the former instance, subclonal mutations might simply be undetectable at presentation due to the limited sensitivity of our PCR assays and they would become identifiable subsequently following clonal selection favored by a proliferative advantage of mutated cells. Mutations acquired after initial diagnosis might be the consequence of genomic instability, cytotoxic treatment or both.

Combined with clinical investigation, more extensive biochemical and molecular studies on APL relapses should allow better understanting of ATRA resistance development and might provide important therapeutic indications for overcoming it.

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References

- Miller WH Jr., Levine K, De Blasio A, Frankel SR, Dmitrovsky E, Warrell RP Jr. Detection of minimal residual disease in acute promyelocytic leukemia by a reverse transcription polymerase chain reaction. Blood 1993; 82:1689-94.
- Lo Coco F, Nervi C, Avvisati G, Mandelli F. Acute promyelocytic leukemia: a curable disease. Leukemia 1998; 12:1866-80.
- 3. Muindi J, Frankel S, Miller WH, et al. Continuous treatment with all-trans retinoic acid causes a progressive reduction in plasma drug concentrations: implications for relapse and retinoid "resistance" in patients with acute promyelocytic leukemia. Blood 1992; 79:299-307.
- Delva L, Cornic M, Balitrand N, et al. Resistance to alltrans retinoic acid (ATRA) therapy in relapsing acute promyelocytic leukemia: study of in vitro sensitivity and cellular retinoid binding protein levels in leukemic cells. Blood 1993; 82:2175-81.
- Shao W, Benedetti L, Lamph WW, Nervi C, Miller WH. A retinoid resistant acute promyelocytic subclone expresses a dominant negative PML/RARα mutation. Blood 1997; 89:4282-9.
- Imazumi M, Suzuki H, Sato A, et al. Mutations in the E-domain of RARα portion of the PML/RARα chimeric gene may confer clinical resistance to alltrans retinoic acid in acute promyelocytic leukemia. Blood 1998; 92:374-82.
- 7. Ding W, Nobile LM, Grills G, et al. Leukemic cellular retinoid acid resistance and missense mutations in the PML/RAR α fusion gene after relapse of acute promyelocytic leukemia with all-trans retinoic acid and intensive chemotherapy. Blood 1998; 92:1172-83.
- Shao W, Fanell M, Ferrara FF, et al. Arsenic trioxide as an inducer of apoptosis and loss od PML/RARα protein in acute promyelocytic leukemia. J Natl Cancer Inst 1998: 90:124-33.
- Inst 1998; 90:124-33.
 Marasca R, Zucchini P, Galimberti S, et al. Missense mutations in the PML/RARα ligand binding domain in ATRA-resistant As₂O₃ sensitive relapsed acute promyelocytic leukemia. Haematologica 1999; 84: 963-8.
- Lamour F, Lardelli P, Apfel C. Analysis of the ligandbinding domain of human retinoic acid receptor a by site-directed mutagenesis. Mol Cell Biol 1996; 16: 5386-92.

Treatment of myelodysplastic syndromes

Recent articles in this journal have analyzed etiology, pathogenesis, diagnosis and treatment of myelodysplastic syndromes (MDSs).¹⁻¹¹ A metaanalysis of available studies has showed that the only two treatments that can prolong survival are allogeneic stem cell transplantation (SCT) and intensive chemotherapy, although only a minority of MDS patients can really benefit from them.¹⁰

In this issue Santini and Giles¹² analyze the potential of amifostine in the treatment of MDSs. There are great expectations about this drug, but – as the authors underline – prospective randomized clinical trials are needed to establish the real usefulness of amifostine.

References

- 1. Gallagher A, Darley RL, Padua R. The molecular basis of myelodysplastic syndromes. Haematologica 1997; 82:191-204.
- Aul C, Bowen DT, Yoshida Y. Pathogenesis, etiology and epidemiology of myelodysplastic syndromes. Haematologica 1998; 83:71-86.
- Vallespi T, Imbert M, Mecucci C, Preudhomme C, Fenaux P. Diagnosis, classification, and cytogenetics of myelodysplastic syndromes. Haematologica 1998; 83:258-75.
- Rigolin GM, Cuneo A, Roberti MG, Bardi A, Castoldi G. Myelodysplastic syndromes with monocytic component: hematologic and cytogenetic characterization. Haematologica 1997; 82:25-30.
- Del Cañizo MC, Brufau A, Mota A, et al. The value of cell cultures for the diagnosis of mixed myelodysplastic/myeloproliferative disorders. Haematologica 1998; 83:3-7.
- Elghetany MT. Surface marker abnormalities in myelodysplastic syndromes. Haematologica 1998; 83:1104-15.
- Invernizzi R, Pecci A, Rossi G, et al. Idarubicin and cytosine arabinoside in the induction and maintenance therapy of high-risk myelodysplastic syndromes. Idarubicin and cytosine arabinoside in the induction and maintenance therapy of high-risk myelodysplastic syndromes. Haematologica 1997; 82:660-3.
- Estey EH. Prognosis and therapy of secondary myelodysplastic syndromes. Haematologica 1998; 83: 543-9.
- Sanz GF, Sanz MA, Greenberg PL. Prognostic factors and scoring systems in myelodysplastic syndromes. Haematologica 1998; 83:358-68.
 Cazzola M, Anderson JE, Ganser A, Hellström-Lind-
- Cazzola M, Anderson JE, Ganser A, Hellström-Lindberg E. A patient-oriented approach to treatment of myelodysplastic syndromes. Haematologica 1998; 83: 910-35.
- Mandelli F, Petti MC, Lo Coco F. Therapy of acute myeloid leukemia: towards a patient-oriented, riskadapted approach. Haematologica 1998; 83:1015-23.
- 12. Santini V, Giles FJ. The potential of amifostine: from cytoprotectant to therapeutic agent. Haematologica 1999; 84:1035-42.