



## Teclistamab in relapsed/refractory Waldenström macroglobulinemia

by Michal Kascak, Ludmila Muronova, Ondrej Venglar, Gabriela Kascakova, Eva Radova, Lucie Broskevicova, Tereza Popkova, Katerina Karasova, Tereza Novakova, Jana Mihályová, Juraj Duras, Pavlina Kusnierova, Martin Havel, Tereza Sevcikova, David Zihala, Roman Hajek and Tomas Jelinek

Received: April 28, 2026.

Accepted: July 3, 2026.

Citation: Michal Kascak, Ludmila Muronova, Ondrej Venglar, Gabriela Kascakova, Eva Radova, Lucie Broskevicova, Tereza Popkova, Katerina Karasova, Tereza Novakova, Jana Mihályová, Juraj Duras, Pavlina Kusnierova, Martin Havel, Tereza Sevcikova, David Zihala, Roman Hajek and Tomas Jelinek. Teclistamab in relapsed/refractory Waldenström macroglobulinemia.

Haematologica. 2026 July 16. doi: 10.3324/haematol.2026.301159 [Epub ahead of print]

### *Publisher's Disclaimer.*

*E-publishing ahead of print is increasingly important for the rapid dissemination of science.*

*Haematologica is, therefore, E-publishing PDF files of an early version of manuscripts that have completed a regular peer review and have been accepted for publication.*

*E-publishing of this PDF file has been approved by the authors.*

*After having E-published Ahead of Print, manuscripts will then undergo technical and English editing, typesetting, proof correction and be presented for the authors' final approval, the final version of the manuscript will then appear in a regular issue of the journal.*

*All legal disclaimers that apply to the journal also pertain to this production process.*

# Teclistamab in relapsed/refractory Waldenström macroglobulinemia

## Authors

Michal Kascak <sup>1,2#</sup>, Ludmila Muronova <sup>1, 2#</sup>, Ondrej Venglar <sup>1,2</sup>, Gabriela Kascakova <sup>3</sup>, Eva Radova <sup>1,2</sup>, Lucie Broskevicova <sup>1,2</sup>, Tereza Popkova <sup>1,2</sup>, Katerina Karasova <sup>1,2</sup>, Tereza Novakova <sup>1,2</sup>, Jana Mihályová <sup>1,2</sup>, Juraj Duras <sup>1,2</sup>, Pavlina Kusnierova <sup>4,5</sup>, Martin Havel <sup>3</sup>, Tereza Sevcikova <sup>1,6</sup>, David Zihala <sup>1,2,6</sup>, Roman Hajek <sup>1,2</sup>, Tomas Jelinek <sup>1,2\*</sup>

# MK and LM contributed equally as co-first authors

## Affiliations

<sup>1</sup>Department of Hematooncology, University Hospital Ostrava, Ostrava, Czech Republic;

<sup>2</sup>Department of Hematooncology, Faculty of Medicine, University of Ostrava, Ostrava, Czech Republic

<sup>3</sup>Department of Nuclear Medicine, University Hospital Ostrava, Ostrava, Czech Republic.

<sup>4</sup>Institute of Laboratory Medicine, Faculty of Medicine, University of Ostrava, Ostrava, Czech Republic.

<sup>5</sup> Institute of Laboratory Medicine, Department of Clinical Biochemistry, University Hospital Ostrava, Ostrava, Czech Republic.

<sup>6</sup>Department of Biology and Ecology, Faculty of Science, University of Ostrava, Ostrava, Czech Republic.

\*Corresponding author:

Assoc. Prof. Tomas Jelinek, MD, PhD

Department of Hematooncology

University Hospital Ostrava

17. listopadu 1790/5, 708 52 Ostrava, Czech Republic

Email: tomas.jelinek@fno.cz

#### Author Contribution:

Kascak M, Muronova L: Conceptualization, Data curation, Writing – original draft, Writing – review & editing.

Venglar O: Flow cytometric analysis, Data curation, Creating figures, Writing – review & editing.

Kascakova G, Havel M: Radiologic assessment of imaging studies, Writing – review & editing.

Kusnierova P: Biochemical analyses, Writing – review & editing.

Jelinek T: Conceptualization, Data curation, Writing – review & editing.

Radova E, Broskevicova L, Popkova T, Karasova K, Novakova T, Mihalyova J, Duras J, Sevcikova T, Zihala D, Hajek R: Writing – review & editing.

#### Conflict of Interest:

M.K. has received honoraria for participation in advisory boards and satellite symposia from Gilead, Takeda, BMS, Roche, Swixx and AstraZeneca.

J.M. has received conference sponsorship from Abbvie and Johnson&Johnson, and honoraria for advisory boards from Gilead, Abbvie, AstraZeneca, Johnson&Johnson.

T.P. has received honoraria from Johnson&Johnson, Pfizer and Sanofi.

J.D. has received honoraria from Abbvie, Roche. Consulting or Advisory Role: Company: Abbvie, Bristol Myers Squibb, Eli Lilly, Johnson and Johnson, Roche, Swixx, Takeda.

R.H. reports grants or contracts from Amgen, BMS, Celgene, Janssen, Novartis, and Takeda; consulting fees from AbbVie, Amgen, BMS, Celgene, Janssen, Novartis, PharmaMar, and Takeda; honoraria for lectures, presentations, and speaker bureaus from Amgen, BMS, Celgene, Janssen, PharmaMar, and Takeda; support for attending meetings and/or travel from Amgen, Celgene, Janssen, and Takeda; and participation on a data safety monitoring board or advisory board for Amgen, BMS, GSK, Janssen, Oncopeptides, Sanofi, and Takeda.

T.J. has received research funding from Johnson&Johnson and Sanofi, honoraria and consultancy/advisory role for Bristol Myers Squibb, GlaxoSmithKline, Johnson&Johnson, Pfizer, and Sanofi.

L.M., O.V., G.K., E.R., K.K., T.N., T.S., and D.Z. disclose no conflict of interest.

#### Acknowledgment:

This work was supported by OPJAK SALVAGE project (No. CZ.02.01.01/00/22\_008/0004644) with co-financing from EU and the State Budget; by the European Union project LERCO (No. CZ.10.03.01/00/22\_003/0000003); by the Czech Health Research Council (NU23-03-00374); by Institutional support (MH CZ— DRO— FNOs/2022, MH CZ—DRO— FNOs/2023, MH CZ—DRO— FNOs/2024); by the Ministry of

Education, Youth and Sports of the Czech Republic through the e-INFRA CZ (ID:90254). TJ was supported by EHA Physician Scientist Research Grant RG-202409-06772.

Data-sharing statement: The data that support the findings of this case report are available on the request from the corresponding author.

Waldenström macroglobulinemia (WM) is a rare indolent B lymphoma with an annual incidence of 3-4 cases per million.<sup>1</sup> The disease is characterized by bone marrow (BM) infiltration by malignant cells with lymphoplasmacytic differentiation, a detectable serum monoclonal IgM immunoglobulin (mIgM), and the recurrent MYD88 L265P somatic mutation in the majority of cases (>90 %).<sup>2</sup> Time-limited rituximab-based chemoimmunotherapy, such as bendamustine/rituximab, and continuous therapy with covalent Bruton tyrosine kinase inhibitors (BTKi) (ibrutinib, zanubrutinib) are key treatment options in the management of WM.<sup>3</sup> While current therapies provide high overall response rates with durable control, complete remissions (CR) with eradication of clonal cells remain difficult to achieve, and the disease is still considered incurable.<sup>3</sup> Relapsed disease, particularly after BTK inhibitor failure, is a therapeutic challenge and is associated with an adverse prognosis.<sup>4</sup>

Bispecific antibodies (bsAbs) redirect T cells toward malignant cells by simultaneous binding of a tumor-associated antigen and CD3 on T cells. They are highly active agents in relapsed or refractory multiple myeloma (targeting BCMA, GPRC5D) and B-cell lymphomas (CD20).<sup>5</sup> Recent data suggest that anti-BCMA bsAbs effectively eliminate both tumor and normal plasma cells (PCs) as well as B cells.<sup>6</sup> Thus, we hypothesized that teclistamab, a BCMA targeted bsAb, might be a novel and effective treatment approach in WM. Here we present, to our knowledge, the first case report of a relapsed/refractory WM patient successfully treated with an anti-BCMA bsAb. This case report was conducted in accordance with the Declaration of Helsinki. Written informed consent was obtained from the patient, and all samples were processed according to standard operating procedures approved by the Institutional Ethics Committee under approval number 413/2012.

A 56-year-old non-Hispanic White male with otherwise unremarkable medical history was diagnosed with symptomatic WM in 2014, preceded by IgM kappa monoclonal gammopathy of undetermined significance (MGUS) by 9 years. At diagnosis, BM infiltration was 70% by lymphoplasmacytic lymphoma. Next-generation sequencing (NGS) on CD19-sorted BM cells identified MYD88 L265P and a CXCR4 C-terminal truncating frameshift mutation (p.T318Nfs\*26). Disease was low risk by International Prognostic Scoring System for WM (IPSSWM) and intermediate risk according to the modified staging system (MSS-WM). In 2023, WM progression was accompanied by the

development of hepatosplenomegaly. The diagnosis was subsequently reconfirmed by BM evaluation in 2018, 2023, and 2025.

Prior to teclistamab, the patient had received seven lines of therapy: rituximab, cyclophosphamide, dexamethasone (2014); achieving stable disease (SD), rituximab, cyclophosphamide, doxorubicin, vincristine, prednisone (2014); achieving partial remission (PR), bendamustine, rituximab (2018; PR); bortezomib, dexamethasone, rituximab (2023; SD followed by extramedullary progressive disease (PD); ibrutinib (2023-2024; 6 months, SD); zanubrutinib (2024-2025; 12 months); achieving a minimal response (MR) followed by extramedullary progressive disease (PD); venetoclax (2025; ramp-up to 800mg/day, 5 months) achieving SD as the best response followed by PD. Two stem cell collections yielded suboptimal CD34+ counts ( $2.6 \times 10^6$  and  $1.8 \times 10^6$  cells/kg) and the products were not released due to failed quality-control parameters. Mobilization failure was attributed to extensive prior cytotoxic therapy and late collection.

By October 2025, venetoclax was considered ineffective because of worsening edema, ascites, hepatosplenomegaly and absent hematologic response. The liver and spleen measured 23.2 cm and 16.8 cm in the midclavicular line, respectively. Liver or spleen biopsy was not performed. Concomitant AL amyloidosis was not demonstrated on abdominal fat pad biopsy. BM aspirate analyzed by flow cytometry, performed according to EuroFlow protocols, demonstrated 0.15% PCs (100% clonal) with CD19+ CD38+ CD45+ CD138+ cytKappa+ immunophenotype and 0.1% B-lymphocytes (100% clonal) with a CD5-, CD11c-, CD19+, CD20+, CD22+, CD25low, CD27+, CD38low, CD79b+, CD81+, CD305-, sIgM+, cytKAPPA+ immunophenotype. The trephine biopsy was not performed. Fluorescence in situ hybridization was negative for del(17p), and TP53 mutation was not detected by NGS. Spectral cytometry was also performed for detailed lymphoid cell and PC phenotyping with use of anti-BCMA/PE conjugate,<sup>7</sup> detecting 0.1% PCs, 90% of which were BCMA-positive. No considerable BCMA expression was detected on mature B cells.

Teclistamab monotherapy was initiated in December 2025 after off-label reimbursement approval based on highly refractory symptomatic WM, absence of standard alternatives, and the biological rationale for BCMA targeting. At treatment initiation, laboratory workup showed anemia (Hb 112 g/L; 135–175), elevated  $\beta$ 2-microglobulin (6.67 mg/L; 0.8–2.34), IgM 25 g/L (0.4–2.3), m-IgM 13.5 g/L, FLC kappa 357 mg/L (3.3–19.4), and FLC ratio 87, with suppressed IgG 4.4 g/L (7–16) and IgA <0.22 g/L (0.88–4.1); albumin, LDH, and CRP were within normal range, and soluble BCMA was

102.9 µg/L. Teclistamab was administered subcutaneously according to the standard step-up schedule: 0.06 mg/kg, 0.3 mg/kg, then 1.5 mg/kg weekly. Per local institutional protocol, tocilizumab 8 mg/kg was given before the first step-up dose to reduce the risk and severity of cytokine release syndrome (CRS). No CRS or neurotoxicity occurred. Grade 3 neutropenia developed on day 8 and resolved after one dose of granulocyte colony-stimulating factor. From day 12, the patient developed renal impairment (peak creatinine 164µmol/L; 60–105, GFR 0,66ml/s; 1–2.35) of unclear etiology despite comprehensive workup, without oliguria, proteinuria, hematuria, obstruction, infection, thrombotic microangiopathy, or metabolic disturbance. Renal function stabilized after cycle 2 and normalized by cycle 4 without intervention. Immunoglobulin replacement, Pneumocystis prophylaxis, and varicella-zoster prophylaxis were administered. No infectious complications occurred during follow-up to date.

For response assessment, 11th International Workshop on Waldenstrom's macroglobulinemia response criteria were used. A rapid hematologic response was observed: PR after cycle 1, very good partial response after cycle 2, and CR after cycle 4, maintained through cycle 5, with early normalization of free light chains (FLC) (Figure 1, Figure 2C). Flow cytometry of the BM aspirate after the third cycle showed complete eradication of clonal PCs and clonal B cells (Figure 2A). A radiologic response was also documented, with partial regression of hepatomegaly and resolution of splenomegaly; liver and spleen diameters decreased to 19.0 cm and 11.5 cm, respectively (Figure 2B). The patient also experienced rapid clinical improvement during cycle 1, with disappearance of lower-extremity edema and subsequent regression of ascites. At 5 months, the hematologic CR and clinical benefit were ongoing, with a CR duration of 4 weeks at manuscript submission. Teclistamab is planned for 12 months, with interval extension to every 2 weeks from month 6 and every 4 weeks from month 9 if disease control and tolerability persist.

Malignant cells in WM most likely originate from an IgM+ memory B cell with partial plasma-cell differentiation into a lymphoplasmacytic/plasma-cell clone that secretes IgM.<sup>8</sup> Flow cytometry studies in WM have clearly demonstrated the co-existence of clonal B-cells and an identical light-chain-isotype clonal PC population with an immunophenotypic profile resembling normal PCs rather than myeloma PCs.<sup>9</sup> This biological overlap supports the investigation of B-cell and plasma-cell-associated targets as potential therapeutic strategies in WM. Daratumumab monotherapy produced

suboptimal activity in WM, with an overall response rate of 23% and median progression-free survival of 2 months, underscoring the need for alternative targets and strategies.<sup>10</sup> Direct data on BCMA expression in WM are limited; however, broader analyses suggest BCMA expression in subsets of non-Hodgkin lymphomas, including WM, generally at lower levels than in multiple myeloma.<sup>11</sup> Elsayah et al. found BCMA expression by flow cytometry in 8/12 (67%) WM patients, with mostly low and occasional intermediate expression.<sup>12</sup>

There are conflicting reports on BCMA expression on precursor, naïve, and memory B cells.<sup>13</sup> Despite limited expression of BCMA on B-cells by flow cytometry, anti-BCMA bsAbs are able to effectively target and eliminate all B-cell subsets from the small pre-B stage onward. Single-cell RNA sequencing demonstrated low but detectable BCMA mRNA expression across mature B cells and B-cell precursors, including immature B cells and both small and large pre-B cells, with the highest expression observed in the small pre-B subset.<sup>6</sup>

BsAbs are among the most active agents currently available for relapsed or refractory multiple myeloma and B-cell lymphomas, producing high response rates even in heavily pretreated patients.<sup>5</sup> Off-label use of teclistamab has recently been reported in three patients with refractory BCMA-positive plasmablastic lymphoma.<sup>14</sup> Evidence of bsAbs activity in WM is very limited. A 2022 case report described plamotamab, a CD20×CD3 bsAb, in ibrutinib-refractory, CXCR4-mutated extramedullary WM, inducing minor hematologic response and near-complete extramedullary disease resolution before relapse with CD20-negative disease.<sup>15</sup> A phase II trial of epcoritamab (CD20×CD3) in previously treated WM (NCT06510491) is currently recruiting; however, no preliminary results have been reported to date. Etentamig (ABBV-383), a BCMA×CD3 bsAB, is being evaluated in phase I/II trial for RRWM (NCT07420959), led by Mayo Clinic.

In this heavily pretreated relapsed/refractory WM patient, teclistamab monotherapy led to a rapid and deep hematologic response (CR) with complete eradication of clonal PCs and B cells in the BM. The patient had received multiple prior lines of therapy – including standard chemoimmunotherapy, a proteasome inhibitor, first and second generation BTKi, and venetoclax administered over several years, none of which induced a major response. In contrast, teclistamab proved to be highly effective and very well tolerated with no serious adverse events. Although limited to a single-patient observation, this case provides initial clinical evidence supporting BCMA-directed bsAbs

as a promising therapeutic approach in WM by targeting both malignant plasma-cell and B-cells.

## References

1. McMaster ML. The epidemiology of Waldenström macroglobulinemia. *Semin Hematol.* 2023;60(2):65-72.
2. Alaggio R, Amador C, Anagnostopoulos I, et al. The 5th edition of the World Health Organization classification of haematolymphoid tumours: lymphoid neoplasms. *Leukemia.* 2022;36(7):1720-1748.
3. Gertz MA. Waldenström macroglobulinemia: 2025 update on diagnosis, risk stratification, and management. *Am J Hematol.* 2025;100(6):1061-1073.
4. Frustaci AM, Zappaterra A, Galitzia A, et al. Salvage treatment after covalent BTKi failure: an unmet need in clinical practice in Waldenström macroglobulinemia. *Hemasphere.* 2025;9(2):e70094.
5. Braun A, Gouni S, Pulles A, Strati P, Minnema MC, Budde LE. Bispecific antibody use in patients with lymphoma and multiple myeloma. *Am Soc Clin Oncol Educ Book.* 2024;44(3):e433516.
6. Jelinek T, Zihala D, Zabaleta A, et al. Selective B-cell subset depletion underlies increased infection risk in patients with MM treated with anti-BCMA vs anti-GPRC5D bsAbs. *Blood.* 2026;147(10):1070-1082.
7. Venglar O, Radova E, Broskevicova L, Hajek R, Jelinek T. OMIP-117: 40-parameter/37-color spectral cytometry panel for robust immunoprofiling of human lymphoid subsets in cancer patients. *Cytometry A.* 2025;107(10):641-648.
8. García-Sanz R, Jiménez C, Puig N, et al. Origin of Waldenström's macroglobulinaemia. *Best Pract Res Clin Haematol.* 2016;29(2):136-147.
9. Jelinek T, Bezdekova R, Zatopkova M, et al. Current applications of multiparameter flow cytometry in plasma cell disorders. *Blood Cancer J.* 2017;7(10):e617.
10. Castillo JJ, Libby EN, Ansell SM, et al. Multicenter phase 2 study of daratumumab monotherapy in patients with previously treated Waldenström macroglobulinemia. *Blood Adv.* 2020;4(20):5089-5092.
11. Dogan A, Siegel D, Tran N, et al. B-cell maturation antigen expression across hematologic cancers: a systematic literature review. *Blood Cancer J.* 2020;10(6):73.
12. Elsawa SF, Novak AJ, Grote DM, et al. B-lymphocyte stimulator (BLyS) stimulates immunoglobulin production and malignant B-cell growth in Waldenström macroglobulinemia. *Blood.* 2006;107(7):2882-2888.
13. Carpenter RO, Evbuomwan MO, Pittaluga S, et al. B-cell maturation antigen is a promising target for the immunotherapy of multiple myeloma. *Clin Cancer Res.* 2013;19(8):2048-2060.
14. Cannova JM, DuVall AS, Kline J, Feinberg N, Wool GD, Riedell PA. Targeting BCMA in plasmablastic lymphoma with teclistamab: a case study of three patients. *J Natl Compr Canc Netw.* 2026;24(3):128-131.

15. Parrondo RD, Paulus A, Alegria V, et al. Plamotamab (XmAb 13676) for ibrutinib-refractory CXCR4-mutated extramedullary Waldenström macroglobulinemia. *Leuk Lymphoma*. 2022;63(3):738-742.

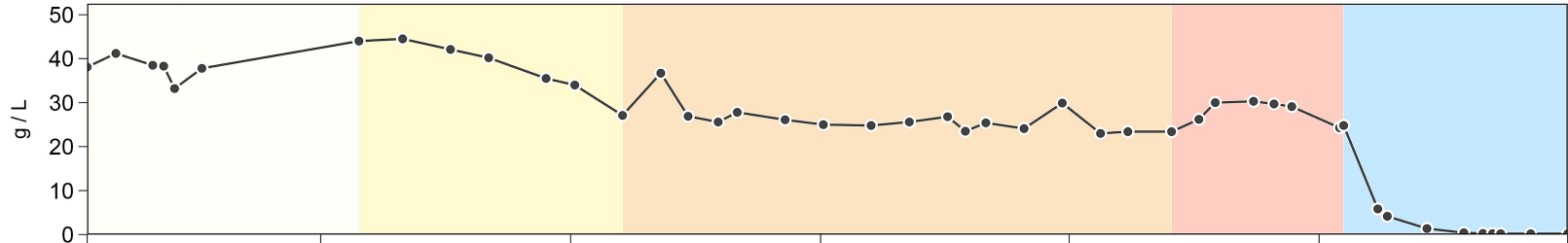
**FIGURE 1.** Longitudinal monitoring of serum total IgM, involved free light chain kappa concentrations during treatment course. Borte-Dexa-Ritu: bortezomib-dexamethasone-rituximab; CT: computed tomography; HSM: hepatosplenomegaly; TCP: thrombocytopenia.

**FIGURE 2. Multimodal Assessment of Response to Teclistamab Therapy (A)** Flow cytometry of the bone marrow aspirate before teclistamab administration and after 3 months of therapy, showing complete eradication of both - clonal PCs and B cells. **(B)** Computed tomography demonstrating hepatosplenomegaly before teclistamab initiation and partial regression of hepatomegaly and normalization of spleen size at month 4 of therapy. **(C)** Serum protein electrophoresis and immunofixation at baseline and at month 4 of therapy.

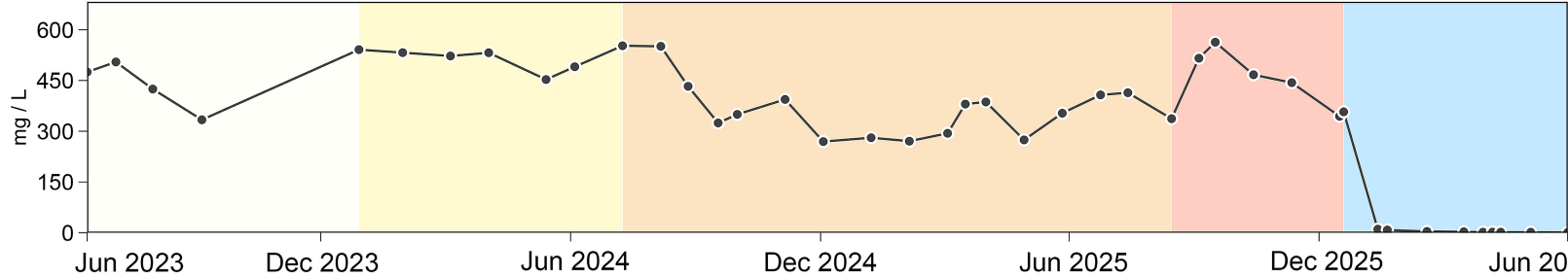
Regimen



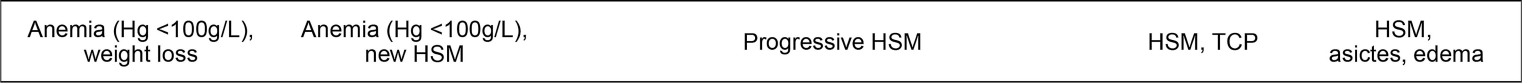
Total IgM



FLC Kappa



Indication for Treatment



Hematological Response



Extramedullary Response (by CT)



