

Integrating killer cell immunoglobulin-like receptor high-resolution genotyping for predicting transplant outcomes in allogeneic hematopoietic stem cell transplantation

Antonia Schäfer,¹ Stéphane Buhler,¹ Ticiana D.J. Farias,² Katherine M. Kichula,² Helen Baldomero,³ Zuleika Calderin Sollet,¹ Sylvie Ferrari-Lacraz,¹ Baptiste Micheli,⁴ Stavroula Masouridi-Levrat,⁵ Vanessa Mesquita,⁶ Oliver Kürsteiner,⁷ Gayathri Nair,⁷ Jörg Halter,³ Tayfun Güngör,⁸ Dominik Schneidawind,⁶ Yves Chalandon,⁵ Jakob R. Passweg,³ Paul J. Norman² and Jean Villard¹ on behalf of the Swiss Blood Stem Cell Transplantation Group

¹Transplantation Immunology Unit and National Reference Laboratory for Histocompatibility, Department of Diagnostics, Geneva University Hospitals, Geneva, Switzerland; ²Department of Biomedical Informatics, and Department of Immunology and Microbiology, University of Colorado School of Medicine, Aurora, CO, USA; ³Division of Hematology, Basel University Hospital, Basel, Switzerland; ⁴Genetic Medicine Division, Department of Diagnostics, Geneva University Hospitals, Geneva, Switzerland; ⁵Division of Hematology, Department of Oncology, Geneva University Hospitals, Geneva, Switzerland; ⁶Department of Medical Oncology and Hematology, University Hospital Zurich, Zurich, Switzerland; ⁷Swiss Blood Stem Cells Registry, Blutspende SRK Schweiz, Bern, Switzerland and ⁸Department of Stem Cell Transplantation, University Children's Hospital Zurich, CIC 334, Zurich, Switzerland

Correspondence: J. Villard
jean.villard@hcuge.ch

Received: November 28, 2024.
Accepted: June 26, 2025.
Early view: July 10, 2025.

<https://doi.org/10.3324/haematol.2024.287061>

©2026 Ferrata Storti Foundation

Published under a CC BY-NC license



Supplementary material

Antonia Schäfer¹, Stéphane Buhler¹, Ticiana D. J. Farias², Katherine M. Kichula², Helen Baldomero³, Zuleika Calderin Sollet¹, Sylvie Ferrari-Lacraz¹, Baptiste Micheli⁴, Stavroula Masouridi-Levrat⁵, Vanessa Mesquita⁶, Oliver Kürsteiner⁷, Gayathri Nair⁷, Jörg Halter³, Tayfun Güngör⁸, Dominik Schneidawind⁶, Yves Chalandon⁵, Jakob R. Passweg³, Paul J. Norman², Jean Villard¹; on behalf of the Swiss Blood Stem Cell Transplantation Group.

¹*Transplantation Immunology Unit and National Reference Laboratory for Histocompatibility, Department of Diagnostic, Geneva University Hospitals, Geneva, Switzerland*

²*Department of Biomedical Informatics, and Department of Immunology and Microbiology, University of Colorado School of Medicine, Aurora, CO, USA*

³*Division of Hematology, Basel University Hospital, Basel, Switzerland*

⁴*Genetic Medicine Division, Department of Diagnostic, Geneva University Hospitals, Geneva, Switzerland*

⁵*Division of Hematology, Department of Oncology, Geneva University Hospitals, Geneva, Switzerland*

⁶*Department of Medical Oncology and Hematology, University Hospital Zurich, Zurich, Switzerland;*

⁷*Swiss Blood Stem Cells Registry, Blutspende SRK Schweiz, Bern, Switzerland*

⁸*Department of Stem Cell Transplantation, University Children's Hospital Zurich, CIC 334, Zurich, Switzerland*

Corresponding author:

*Jean Villard, Transplantation Immunology Unit and National Reference Laboratory for Histocompatibility, Geneva University Hospitals, Gabrielle-Perret-Gentil 4, 1211 Geneva 4, +41 22 372 93 94/ +41 79 553 34 09, jean.villard@hcuge.ch

Table of contents

Supplementary methods

Table S1. KIR alleles characteristics of activating KIRs in the cohort

Table S2. KIR alleles characteristics of inhibitory KIRs in the cohort

Table S3. Frequency of *KIR2DS4*, *KIR2DL1*, *KIR2DL2/L3* and *KIR3DL1* allotypes in the cohort according to their functional aspects

Table S4. Stratification of *KIR2DS4*, *KIR2DL1*, *KIR2DL2/L3* and *KIR3DL1* alleles in the cohort according to their functional aspects

Table S5. Multivariable Cox regression analysis of KIR variables on transplant outcomes in the entire cohort

Table S6. Multivariable Cox regression analysis of KIR variables on transplant outcomes in AML recipients

Supplementary methods

1. Study cohort

The post-transplant immunosuppression protocol for GvHD prophylaxis consisted of either a calcineurin inhibitor with methotrexate or a calcineurin inhibitor with mycophenolate mofetil. Anti-thymocyte globulin (ATG) was used in 10/10 matched unrelated transplants and in 9/10 matched unrelated transplants until 2015. Post-transplant cyclophosphamide (PTCy) was used in 9/10 matched unrelated transplants from 2016 onward.

2. High-resolution *KIR* genotyping

Genomic DNA was directly extracted from whole blood samples and purified using the QIAGEN Blood and Tissue Kit according to the manufacturer's instructions. DNA purity and concentration were assessed by Qubit fluorometer. All samples were stored at 4°C until use. For library preparation, 500 ng of genomic DNA was first fragmented by digestive enzymes (New England Biolabs, Boston, MA, USA), followed by barcode ligation with unique adaptors (IDT, Coralville, Iowa, USA). After post-ligation cleanup, dual size selection was performed with AMPure magnetic beads (Beckman Coulter, Brea, California, USA) to acquire fragment sizes of 800 to 1200 bp length. In a second step, a pool of oligonucleotide probes specific for the KIR and HLA genomic regions was used for the targeted capture (1). Final enriched libraries were normalized to a concentration of 12 pmol/l. Paired-end sequencing was performed using a NovaSeq instrument with a sequencing length of 2 x 250 bp (Illumina, San Diego, CA, USA).

3. High-resolution *HLA* genotyping

High-resolution *HLA* genotyping was performed on all recipients and donors using reverse PCR-sequence-specific oligonucleotide microbead arrays, high-throughput sequencing (One Lambda, Canoga Park, CA, USA) or PCR-sequence-specific primers (Genovision, Milan Analytika AG, Switzerland).

4. *KIR* haplotype assignment

The centromeric portion is defined as *KIR* genes present in between the framework genes *KIR3DL3* and *KIR3DP1*, while the telomeric part encompasses genes from *KIR2DL4* to *KIR3DL2*. The presence of one or more of the following *KIRs* – *2DL2*, *2DL5*, *3DS1*, *2DS1*, *2DS2*, *2DS3* and *2DS5* – defines Bx haplotypes, whereas the presence of *2DL1*, *2DL3*, *3DL1* and *2DS4* marks A haplotypes.

5. *KIR* allotype assignment

KIR allotype refers to a distinct amino acid sequence and *KIR* alleles were grouped according to published known *KIR* allotypes: *KIR3DL1* expression levels were classified into high, low or null as previously described (2, 3). *KIR2DS4* alleles were classified into the full-length version or the truncated variant (4). *KIR2DL1* alleles were segregated into strongly and weakly inhibiting based on the presence of an arginine or a cysteine group at position 245, respectively (5). Functionally stronger *KIR2DL2* and *KIR2DL3* alleles were defined as alleles having a glutamic acid at position 35, whereas weak alleles were defined by the presence of a glutamine at position 35 (6) (Supplemental Tables S3 and S4).

6. *KIR* and HLA interactions

HLA-A, -B and -C alleles were categorized according to their relevant epitopes following known classifications using the IPD database (7). *KIR*-HLA pairs were then summed up for each individual, with homozygous *KIR* or HLA alleles counted twice: HLA-C (C1 epitope) and HLA-B*46/B*73 with *KIR2DL2* and *KIR2DL3* (8), HLA-C (C2 epitope) with *KIR2DL1* (8), HLA-C (C2 epitope) with *KIR2DS1*, HLA-C2 (*02:02, *04:01, *05:01), HLA-C1 (*01:02, *14:02, *16:01) and HLA-A*11 with *KIR2DS4**001 (8-10), HLA-C1 (*16, *01:02) and HLA-A*11:01 with *KIR2DS2* (10-12), HLA-A (Bw4 epitope) and HLA-B (Bw4 epitope) with *KIR3DL1* (13, 14). HLA-A25 and HLA-A23 molecules were not considered ligands for *KIR3DL1* as they do not educate *KIR3DL1*⁺ NK cells (15, 16). *KIR2DS5**003, *004, *005, *006, *007, *008 with HLA-C2 (17). The interaction between *KIR3DS1* and HLA-B was assumed based on its 97% sequence homology in the extracellular domain with the *KIR3DL1* receptor, despite the lack of *in vitro* demonstration (18).

7. Statistical endpoints and analysis

The following covariates were tested by forward selection in univariable analysis and by backward selection to eliminate non-significant variables in multiple regression models: recipient and donor age, disease type, HLA matching, Karnofsky performance status, EBMT risk score, disease status, conditioning regimen, graft source, *in vitro* T-cell depletion, total body irradiation, comorbid conditions, donor/recipient cytomegalovirus matching, gender matching and transplant center.

Descriptive results are presented as medians and interquartile ranges (IQRs) for continuous variables and counts and percentages for categorical variables.

Table S1. *KIR* alleles characteristics of activating KIRs in the cohort ($n = 1247$).

<i>KIR2DS1</i>	<i>k</i>	%	% (Amorim <i>et al.</i>) *	<i>KIR2DS4</i>	<i>k</i>	%	% (Amorim <i>et al.</i>)
*null	1951	78.23	77.63	*null	551	22.1	22.65
*001	1	0.04	-	*00101	479	19.2	27.82
*00201	462	18.52	17.42	*00104	2	0.008	1.92
*00502	21	0.84	0.02347	*00301	576	23.1	14.62
*006	31	1.24	0.28	*00401	71	2.85	3.57
*008	1	0.04	-	*00601	426	17.08	12.04
*011	2	0.08	-	*010	359	14.4	14.34
*unresolved	25	1.0	4.58	*016	1	0.004	-
<i>KIR2DS2</i>	<i>k</i>	%	% (Amorim <i>et al.</i>)	*022	2	0.008	-
*null	1738	69.68	70.63	*unresolved	27	1.08	2.89
*00101	701	28.11	24.11	<i>KIR3DS1</i>	<i>k</i>	%	% (Amorim <i>et al.</i>)
*00102	2	0.08	-	*null	1950	77.97	-
*00104	1	0.04	-	*01301	525	21	18.87
*00106	2	0.08	-	*014	2	0.008	-
*00107	1	0.04	-	*049N	19	0.76	-
*002	34	1.36	0.89	*1107	1	0.004	-
*02001	6	0.24	-	*unresolved	4	0.16	-
*005	1	0.04	-				
*007	1	0.04	-				
*008	1	0.04	0.0235				
*unresolved	6	0.24	4.30				

*Comparative allele frequencies from the publication by Amorim *et al.* are shown (19).

Table S2. *KIR* alleles characteristics of inhibitory *KIRs* in the cohort ($n = 1247$).

<i>KIR2DL1</i>	<i>k</i>	%	% (Amorim <i>et al.</i>) *	<i>KIR2DL2/L3</i>	<i>k</i>	%	% (Amorim <i>et al.</i>)
*00101	116	4.63	5.35	3*00101	957	38.7	38.36
*00201	583	23.25	24.29	3*00110	14	0.56	1.10
*00302	914	36.46	38.12	3*00201	591	23.6	25.21
*00303	1	0.004	-	3*003	12	0.48	0.68
*00401	281	11.21	11.48	3*00501	116	4.65	4.30
*00402	12	0.48	0.26	3*006	1	0.04	0.07
*007	34	1.36	1.17	3*009	3	0.12	0.07
*008	4	0.16	0.40	3*015	2	0.08	0.12
*010	1	0.04	-	3*030	1	0.04	0.0469
*01201	1	0.04	-	*unresolved	45	1.8	2.42
*01202	2	0.08	-				
*014	1	0.04	0.02347				
*020	4	0.16	0.33				
*029	1	0.04	-				
*3201	8	0.32	0.40				
*03701	9	0.36	-				
*040	1	0.04	-				
*04301	5	0.2	-				
*044	4	0.16	-				
*049	1	0.04	-				
*05401	1	0.04	-				
*057	5	0.2	-				
*063	1	0.04	-				
*null	425	16.95	16.76				
*unresolved	92	3.67	1.03				
<i>KIR2DL2/L3</i>	<i>k</i>	%	% (Amorim <i>et al.</i>)				
2*00101	402	16.1	14.81				
2*00301	322	12.9	11.55				
2*00303	2	0.08	-				
2*012	3	0.12	0.05				
2*unresolved	23	0.92	-				

<i>KIR3DL1</i>	<i>k</i>	%	% (Amorim <i>et al.</i>) *
*00101	385	15.4	16.38
*00103	1	0.04	-
*00201	286	11.43	9.46
*00401	360	14.4	13.85
*00402	30	1.2	1.57
*00501	344	13.75	12.18
*00701	64	2.56	2.68
*00801	149	5.96	4.81
*00901	18	0.72	0.99
*01501	5	0.2	0.19
*01502	149	5.96	6.24
*01702	1	0.04	0.05
*019	17	0.68	0.82
*02001	59	2.36	-
*021	1	0.04	0.05
*02901	3	0.12	0.05
*03101	2	0.08	-
*033	3	0.12	0.05
*039	1	0.04	0.16
*043	2	0.08	0.02
*052	1	0.04	0.05
*053	14	0.56	-
*072	1	0.04	0.02
*089	1	0.04	-
*110	1	0.04	-
* <i>null</i>	43	1.72	-
* <i>unresolved</i>	50	2	-

*Comparative allele frequencies from the publication by Amorim *et al.* are shown (19).

Table S3. Frequency of KIR2DS4, KIR2DL1, KIR2DL2/L3 and KIR3DL1 allotypes in the cohort according to their functional aspects ($n = 1247$).

KIR2DS4	<i>n</i>	%
Expressed/expressed	60	4.81
Expressed	361	28.9
Not expressed	808	64.8
Unresolved	18	1.44
KIR2DL1	<i>n</i>	%
Strong/strong or strong	854	68.48
Strong/weak	221	17.7
Weak/weak or weak	82	6.57
Unresolved	51	4.1
KIR2DL2/L3	<i>n</i>	%
E ³⁵ homozygous	157	12.6
E ³⁵ / Q ³⁵ heterozygous	545	43.7
Q ³⁵ homozygous	519	41.6
Unresolved	26	2.1
KIR3DL1	<i>n</i>	%
High/high or high	644	51.64
High/low	167	13.4
Low/low or low	215	17.24
Not expressed	130	10.4
Unresolved	30	2.41

E³⁵; glutamic acid at position 35, Q³⁵; glutamine at the position 35

Table S4. Stratification of *KIR2DS4*, *KIR2DL1*, *KIR2DL2/L3* and *KIR3DL1* alleles in the cohort according to their functional aspects ($n = 1247$).

KIR2DL1	Alleles	Reference
Strong	<i>*00101, *00201, *00301, *00302, *00303, *00501, *008, *01201, *01202, *014, *010, *020, *040, *044, *049, *057, *063, *4301</i>	(5)
Weak	<i>*00401, *00402, *00701, *007, *05401, *029</i>	
Not expressed	<i>*03201</i>	
KIR2DL2/L3		
2DL2 E ³⁵	<i>*00101, *00301, *00303, *012</i>	(6)
2DL3 E ³⁵	<i>*00501, *015</i>	
2DL3 Q ³⁵	<i>*00101, *01101, *00201, *003, *00501, *006, *00701, *009, *015, *030</i>	
KIR3DL1		
High	<i>*00101, *00103, *00201, *00801, *00901, *01501, *01502, *01702, *02001, *02901, *043, *052, *089</i>	(2, 3)
Low	<i>*00501, *00701, *033, *03101, *053</i>	
Not expressed	<i>*00401, *00402, *039, *019, *072</i>	
KIR2DS4		
Expressed	<i>*00101, *00104</i>	(4)
Not expressed	<i>*00301, *00401, *00601, *010, *016, *022</i>	

E³⁵; glutamic acid at position 35, Q³⁵; glutamine at the position 35

Table S5. Multivariable Cox regression analysis of the KIR variables tested on six different transplant outcomes ($n = 1247$).

		OS		TRM	
		<i>Adjusted for Karnofsky score, disease state, disease type, presence of comorbidities</i>		<i>Adjusted for Karnofsky score, disease type, transplant center, source of transplant</i>	
Variable	<i>n</i>	HR (95%-CI)	<i>p</i>	HR (95%-CI)	<i>p</i>
<i>KIR genotype</i>					
AA	406	1		1	
Bx	841	0.71 (0.59 – 0.87)	< 0.001	0.73 (0.56 – 0.96)	0.026
<i>Cen AA</i>	607	1		1	
<i>Cen Bx</i>	640	0.76 (0.63 – 0.92)	0.004	0.76 (0.58 – 1.0)	0.05
<i>Te/ AA</i>	742	1		1	
<i>Te/ Bx</i>	505	0.96 (0.79 – 1.16)	0.69	0.93 (0.71 – 1.23)	0.63
<i>Centromeric genotype</i>					
AA	587	1		1	
cA01/cB01	212	0.82 (0.63 - 1.07)	0.16	0.79 (0.54 – 1.17)	0.24
cA01/cB02	276	0.72 (0.56 – 0.92)	0.009	0.82 (0.58 – 1.16)	0.26
cB01/x	61	0.8 (0.51 – 1.26)	0.34	0.47 (0.2 – 1.06)	0.07
cB02/cB02	40	0.78 (0.44 – 1.38)	0.39	1.1 (0.53 – 2.27)	0.8
<i>B content score</i>					
Neutral	860	1		1	
Better	267	1.01 (0.8 – 1.27)	0.93	1.12 (0.81 – 1.54)	0.49
Best	120	0.83 (0.59 – 1.17)	0.29	0.7 (0.44 – 1.24)	0.25
<i>KIR2DS1 – HLA-C2</i>					
Absence	947	1		1	
Presence	300	0.95 (0.76 – 1.18)	0.65	1.03 (0.76 – 1.41)	0.85
<i>KIR2DS1 – HLA-C2</i>					
Absence	947	1		1	
2DS1 – C1/x	233	0.93 (0.73 – 1.2)	0.59	0.98 (0.69 – 1.39)	0.9
2DS1 – C2/C2	67	1.00 (0.67 – 1.49)	1.0	1.21 (0.7 – 2.1)	0.49
<i>KIR2DS2*00101 – HLA-C*16,C*01:02, A*11:01</i>					

Absence	1035	1		1	
Presence	212	1.15 (0.9 – 1.46)	0.25	0.97 (0.68 – 1.39)	0.89
KIR2DS4*00101 – HLA-C1/C2/A11					
Absence	1000	1		1	
Presence	229	1.26 (1 – 1.59)	0.047	1.65 (1.2 – 2.27)	0.002
KIR3DS1					
Absence	779	1		1	
Presence	468	0.96 (0.79 – 1.17)	0.72	1.0 (0.76 – 1.31)	0.98
KIR3DS1 – Bw4 (HLA-B)					
Absence	946	1		1	
Presence	301	0.88 (0.71 – 1.1)	0.27	1.02 (0.75 – 1.4)	0.89
KIR2DL1 – HLA-C2					
Strong	671	1		1	
Weak	52	0.74 (0.43 – 1.28)	0.28	0.38 (0.14 – 1.03)	0.06
Missing ligand	429	1.06 (0.87 – 1.3)	0.56	0.83 (0.62 – 1.11)	0.2
KIR2DL2/L3 – HLA-C1					
≥ 1 KIR2DL2 – C1	549	1		1	
KIR2DL3 – C1	512	1.38 (1.13 – 1.69)	0.002	1.29 (0.96 – 1.73)	0.09
Missing ligand	186	1.0 (0.75 – 1.34)	1.0	1.2 (0.81 – 1.78)	0.36
KIR2DL2/L3 – HLA-C1					
Weak/weak	437	1		1	
Weak/strong	253	0.83 (0.65 – 1.07)	0.16	0.89 (0.62 – 1.29)	0.55
Strong/strong	342	0.72 (0.56 – 0.91)	0.007	0.68 (0.48 – 0.97)	0.033
Missing ligand	186	0.73 (0.55 – 0.99)	0.041	0.92 (0.62 – 0.97)	0.67
KIR2DL2/L3 – HLA-C1					
No strong 2DL3	949	1		1	
Strong 2DL3	88	0.95 (0.67 – 1.35)	0.77	0.97 (0.57 – 1.66)	0.92
Missing ligand	186	0.85 (0.65 – 1.12)	0.26	1.06 (0.73 – 1.52)	0.77
KIR3DL1 – Bw4 (HLA-B)					

Strong inhibiting	375	1		1	
Weak inhibiting	307	1.23 (0.95 – 1.59)	0.11	1.06 (0.75 – 1.51)	0.73
Non-inhibiting	410	1.05 (0.82 – 1.35)	0.69	0.83 (0.59 – 1.17)	0.3
KIR3DL1 – Bw4 (HLA-A and -B)					
Strong inhibiting	375	1		1	
Weak inhibiting	307	1.23 (0.96 – 1.59)	0.11	1.06 (0.74 – 1.51)	0.75
Non-inhibiting	434	1.06 (0.83 – 1.35)	0.65	0.83 (0.59 – 1.16)	0.27
HLA-A*24 and A*32	100	1.27 (0.88 – 1.83)	0.21	1.1 (0.67 – 1.8)	0.7
KIR3DL1 and KIR2DS1					
Strong + 2DS1	85	1		1	
Weak + 2DS1	75	1.0 (0.59 – 1.69)	1.0	1.74 (0.85 – 3.57)	0.13
None + 2DS1	112	0.86 (0.51 – 1.43)	0.56	1.08 (0.52 – 2.26)	0.84
Strong without 2DS1	290	0.92 (0.6 – 1.42)	0.72	1.35 (0.74 – 2.48)	0.33
Weak without 2DS1	232	1.22 (0.6 – 1.42)	0.37	1.21 (0.64 – 2.29)	0.56
None without 2DS1	298	1.04 (0.68 – 1.58)	0.87	1.05 (0.56 – 1.95)	0.88

		Relapse/progression		PFS	
		<i>Adjusted for age of recipient, disease type, disease state, risk score, source of transplant, transplant center</i>		<i>Adjusted for Karnofsky score, disease state, disease type, presence of comorbidities, transplant center</i>	
Variable	<i>n</i>	HR (95%-CI)	<i>p</i>	HR (95%-CI)	<i>p</i>
KIR genotype					
AA	406	1		1	
Bx	841	1.04 (0.84 – 1.27)	0.73	0.82 (0.69 – 0.98)	0.032
<i>Cen</i> AA	607	1		1	
<i>Cen</i> Bx	640	0.98 (0.81 – 1.19)	0.83	0.88 (0.74 – 1.04)	0.13
<i>Tel</i> AA	742	1		1	
<i>Tel</i> Bx	505	1.02 (0.84 – 1.24)	0.83	0.96 (0.8 – 1.14)	0.63
Centromeric genotype					
AA	587	1		1	
cA01/cB01	212	1.23 (0.95 – 1.6)	0.12	1.01 (0.8 – 1.29)	0.87
cA01/cB02	276	0.88 (0.69 – 1.13)	0.32	0.83 (0.67 – 1.03)	0.09
cB01/x	61	0.95 (0.61 – 1.49)	0.83	0.72 (0.47 – 1.1)	0.13
cB02/cB02	40	1.0 (0.56 – 1.81)	0.99	1.02 (0.61 – 1.7)	0.95
B content score					
Neutral	860	1		1	
Better	267	0.99 (0.78 – 1.26)	0.95	1.09 (0.89 – 1.34)	0.41
Best	120	0.87 (0.62 – 1.23)	0.43	0.78 (0.57 – 1.06)	0.11
KIR2DS1 – HLA-C2					
Absence	947	1		1	
Presence	300	0.99 (0.79 – 1.23)	0.91	0.95 (0.78 – 1.16)	0.6
KIR2DS1 – HLA-C2					
Absence	947	1		1	
2DS1 – C1/x	233	0.97 (0.75 – 1.24)	0.78	0.94 (0.76 – 1.18)	0.62
2DS1 – C2/C2	67	1.06 (0.7 – 1.62)	0.77	0.96 (0.67 – 1.39)	0.83

KIR2DS2*00101 – HLA-C*16,C*01:02,A*11					
Absence	1035	1		1	
Presence	212	1.17 (0.92 – 1.49)	0.19	1.15 (0.92 – 1.43)	0.22
KIR2DS4*00101 – C1/C2/A11					
Absence	1000	1		1	
Presence	229	1.25 (0.98 – 1.59)	0.07	1.39 (1.12 – 1.71)	0.002
KIR3DS1					
Absence	779	1		1	
Presence	468	1.04 (0.86 – 1.27)	0.67	0.99 (0.83 – 1.18)	0.88
KIR3DS1 – Bw4 (HLA-B)					
Absence	946	1		1	
Presence	301	1.0 (0.8 – 1.25)	0.97	0.97 (0.8 – 1.19)	0.8
KIR2DL1 – HLA-C2					
Strong	671	1		1	
Weak	52	0.78 (0.46 – 1.32)	0.36	0.67 (0.41 – 1.09)	0.11
Missing ligand	429	1.11 (0.91 – 1.37)	0.3	1.04 (0.86 – 1.25)	0.68
KIR2DL2/L3 – HLA-C1					
≥ 1 KIR2DL2 – C1	549	1		1	
KIR2DL3 – C1	512	1.08 (0.88 – 1.33)	0.46	1.19 (0.99 – 1.44)	0.06
Missing ligand	186	0.97 (0.72 – 1.3)	0.84	0.95 (0.73 – 1.23)	0.64
KIR2DL2/L3 – HLA-C1					
Weak/weak	437	1		1	
Weak/strong	253	0.97 (0.75 – 1.26)	0.81	0.91 (0.72 – 1.14)	0.41
Strong/strong	342	0.91 (0.71 – 1.16)	0.43	0.81 (0.65 – 1.0)	0.05
Missing ligand	186	0.89 (0.66 – 1.21)	0.46	0.79 (0.6 – 1.03)	0.08
KIR2DL2/L3 – HLA-C1					
No strong 2DL3	949	1		1	
Strong 2DL3	88	0.97 (0.67 – 1.42)	0.89	0.92 (0.66 – 1.27)	0.6
Missing ligand	186	0.93 (0.7 – 1.23)	0.61	0.86 (0.67 – 1.2)	0.23
KIR3DL1 – Bw4 (HLA-B)					

Strong inhibiting	375	1		1	
Weak inhibiting	307	1.7 (1.3 – 2.21)	< 0.001	1.44 (1.14 – 1.81)	0.002
Non-inhibiting	410	1.4 (1.09 – 1.81)	0.009	1.12 (0.89 – 1.4)	0.33
KIR3DL1 – Bw4 (HLA-A and -B)					
Strong inhibiting	375	1		1	
Weak inhibiting	307	1.69 (1.3 – 2.2)	< 0.001	1.44 (1.14 – 1.81)	0.002
Non-inhibiting	434	1.39 (1.08 – 1.78)	0.011	1.11 (0.89 – 1.38)	0.37
HLA-A*24 and A*32	100	0.88 (0.56 – 1.37)	0.56	1.08 (0.77 – 1.52)	0.65
KIR3DL1 and KIR2DS1					
Strong + 2DS1	85	1		1	
Weak + 2DS1	75	1.26 (0.73 – 2.19)	0.41	1.25 (0.77 – 2.03)	0.36
None + 2DS1	112	1.37 (0.83 – 2.27)	0.22	1.07 (0.67 – 1.7)	0.78
Strong without 2DS1	290	0.98 (0.62 – 1.55)	0.94	1.04 (0.7 – 1.56)	0.84
Weak without 2DS1	232	1.82 (1.18 – 2.82)	0.007	1.58 (1.06 – 2.35)	0.025
None without 2DS1	298	1.04 (0.9 – 2.15)	0.14	1.19 (0.8 – 1.76)	0.4

		Acute GvHD		Chronic GvHD		
		<i>Adjusted for disease type, risk score, conditioning regimen, T-cell depletion, source of transplant, transplant center</i>		<i>Adjusted for age of recipient, disease type, transplant center</i>		
Variable	<i>n</i>	HR (95%-CI)	<i>p</i>	<i>n</i>	HR (95%-CI)	<i>p</i>
KIR genotype						
AA	348	1		377	1	
Bx	750	0.95 (0.74 – 1.21)	0.65	766	1.12 (0.91 – 1.39)	0.28
<i>Cen</i> AA	527	1		563	1	
<i>Cen</i> Bx	571	1.09 (0.87 – 1.37)	0.45	580	1.19 (0.98 – 1.44)	0.08
<i>Tel</i> AA	643	1		682	1	
<i>Tel</i> Bx	455	0.9 (0.72 – 1.14)	0.39	461	1.03 (0.83 – 1.24)	0.8
Centromeric genotype						
AA	508	1		545	1	
cA01/cB01	189	1.24 (0.91 – 1.69)	0.17	287	1.31 (1.0 – 1.7)	0.047
cA01/cB02	242	1.06 (0.79 – 1.43)	0.68	253	1.26 (0.99 – 1.6)	0.06
cB01/x	54	0.8 (0.43 – 1.48)	0.47	59	1.01 (0.64 – 1.6)	0.96
cB02/cB02	37	0.85 (0.43 – 1.68)	0.63	31	1.32 (0.79 – 2.21)	0.29
B content score						
Neutral	748	1		797	1	
Better	240	1.08 (0.82 – 1.43)	0.56	237	1.19 (0.95 – 1.5)	0.14
Best	110	0.78 (0.51 – 1.19)	0.25	109	0.95 (0.68 – 1.32)	0.74
KIR2DS1 – HLA-C2						
Absence	829	1		869	1	
Presence	269	1.01 (0.77 – 1.31)	0.96	274	1.07 (0.86 – 1.33)	0.55
KIR2DS1 – HLA-C2						
Absence	829	1		869	1	
2DS1 – C1/x	209	0.99 (0.74 – 1.33)	0.95	213	1.21 (0.96 – 1.52)	0.11
2DS1 – C2/C2	60	1.07 (0.65 – 1.75)	0.8	61	0.63 (0.37 – 1.05)	0.08

KIR2DS2*00101 – HLA-C*16,C*01:02, A*11						
Absence	912	1		946	1	
Presence	186	0.99 (0.74 – 1.34)	0.97	197	1.11 (0.87 – 1.42)	0.4
KIR2DS4*00101 – C1/C2/A11						
Absence	874	1		917	1	
Presence	208	1.23 (0.94 – 1.62)	0.13	208	1.29 (1.02 – 1.64)	0.035
KIR3DS1						
Absence	679	1		716	1	
Presence	419	0.9 (0.71 – 1.14)	0.38	427	1.06 (0.87 – 1.29)	0.54
KIR3DS1 – Bw4 (HLA-B)						
Absence	832	1		716	1	
Presence	266	0.96 (0.73 – 1.27)	0.79	427	1.03 (0.82 – 1.28)	0.84
KIR2DL1 – HLA-C2						
Strong	586	1		619	1	
Weak	48	0.68 (0.35 – 1.35)	0.27	49	1.13 (0.7 – 1.8)	0.62
Missing ligand	379	0.94 (0.73 – 1.2)	0.62	397	1.05 (0.85 – 1.29)	0.64
KIR2DL2/L3 – HLA-C1						
≥ 1 KIR2DL2 – C1	492	1		497	1	
KIR2DL3 – C1	447	0.92 (0.72 – 1.18)	0.54	475	0.84 (0.68 – 1.03)	0.09
Missing ligand	159	0.94 (0.67 – 1.34)	0.74	171	0.69 (0.5 – 0.94)	0.018
KIR2DL2/L3 – HLA-C1						
Weak/weak	380	1		408	1	
Weak/strong	227	1.38 (1.01 – 1.87)	0.042	227	1.36 (1.05 – 1.76)	0.02
Strong/strong	309	1.08 (0.81 – 1.46)	0.59	310	1.27 (1.0 – 1.62)	0.049
Missing ligand	159	1.06 (0.74 – 1.53)	0.75	171	0.87 (0.63 – 1.2)	0.4
KIR2DL2/L3 – HLA-C1						
No strong 2DL3	844	1		869	1	
Strong 2DL3	77	1.13 (0.73 – 1.74)	0.58	81	1.59 (1.14 – 2.22)	0.006
Missing ligand	159	0.95 (0.69 – 1.32)	0.78	171	0.77 (0.57 – 1.04)	0.09
KIR3DL1 – Bw4 (HLA-B)						
Strong inhibiting	330	1		342	1	
Weak inhibiting	276	1.06 (0.78 – 1.44)	0.72	279	1.11 (0.86 – 1.43)	0.44

Non-inhibiting	355	1.08 (0.81 – 1.44)	0.62	382	0.99 (0.77 – 1.26)	0.95
KIR3DL1 – Bw4 (HLA-A and -B)						
Strong inhibiting	330	1		342	1	
Weak inhibiting	276	1.05 (0.77 – 1.43)	0.77	279	1.1 (0.86 – 1.42)	0.45
Non-inhibiting	375	1.05 (0.79 – 1.4)	0.73	405	0.99 (0.78 – 1.25)	0.91
HLA-A*24 and A*32	90	1.01 (0.66 – 1.56)	0.95	88	0.92 (0.63 – 1.37)	0.7
KIR3DL1 and KIR2DS1						
Strong + 2DS1	78	1		79	1	
Weak + 2DS1	67	0.91 (0.47 – 1.76)	0.77	68	1.52 (0.92 – 2.51)	0.1
None + 2DS1	98	1.17 (0.66 – 2.08)	0.59	102	1.07 (0.65 – 1.75)	0.8
Strong without 2DS1	252	0.97 (0.59 – 1.58)	0.89	263	1.1 (0.72 – 1.68)	0.67
Weak without 2DS1	209	1.06 (0.65 – 1.75)	0.81	211	1.08 (0.7 – 1.68)	0.73
None without 2DS1	257	1.0 (0.61 – 1.64)	0.99	280	1.07 (0.7 – 1.64)	0.77

Cen; centromeric, CI; confidence interval, GvHD; graft-versus-host disease, HR; hazard ratio, OS; overall survival, PFS; progression-free survival, Tel; telomeric, TRM; transplant-related mortality

Table S6. Multivariable cox regression analysis of KIR3DL1 and KIR2DS1 interactions tested on six different transplant outcomes in the AML subcohort ($n = 498$).

		Acute GvHD		Chronic GvHD		
		<i>Adjusted for disease type, risk score, conditioning regimen, T-cell depletion, source of transplant, transplant center</i>		<i>Adjusted for age of recipient, disease type, transplant center</i>		
Variable	<i>n</i>	HR (95%-CI)	<i>p</i>	<i>n</i>	HR (95%-CI)	<i>p</i>
KIR3DL1 – Bw4 (HLA-B)						
Strong inhibiting	144	1		144	1	
Weak inhibiting	127	1.17 (0.78 – 1.76)	0.87	127	1.17 (0.78 – 1.76)	0.45
Non-inhibiting	165	1.31 (0.9 – 1.91)	0.86	165	1.31 (0.9 – 1.91)	0.16
KIR3DL1 – Bw4 (HLA-A and -B)						
Strong inhibiting	144	1		144	1	
Weak inhibiting	127	0.96 (0.6 – 1.62)	0.88	127	1.1 (0.86 – 1.42)	0.51
Non-inhibiting	172	0.95 (0.58 – 1.56)	0.84	172	0.99 (0.78 – 1.25)	0.23
HLA-A*24 and A*32	42	2.18 (1.19 – 3.99)	0.01	42	0.92 (0.63 – 1.37)	0.98
KIR3DL1 and KIR2DS1						
Strong + 2DS1	38	1		38	1	
Weak + 2DS1	32	0.65 (0.25 – 1.7)	0.38	32	1.33 (0.62 – 2.85)	0.46
None + 2DS1	47	0.7 (0.29 – 1.66)	0.41	47	1.63 (0.82 – 3.24)	0.16
Strong without 2DS1	106	0.58 (0.27 – 1.23)	0.15	106	0.97 (0.51 – 1.84)	0.93
Weak without 2DS1	95	0.66 (0.32 – 1.38)	0.27	95	1.07 (0.56 – 2.06)	0.84
None without 2DS1	118	0.64 (0.31 – 1.31)	0.22	118	1.14 (0.61 – 2.14)	0.67

		Relapse/progression		PFS		
		<i>Adjusted for age of recipient, disease type, disease state, risk score, source of transplant, transplant center</i>		<i>Adjusted for Karnofsky score, disease state, disease type, presence of comorbidities, transplant center</i>		
Variable	<i>n</i>	HR (95%-CI)	<i>p</i>	<i>n</i>	HR (95%-CI)	<i>p</i>
KIR3DL1 – Bw4 (HLA-B)						
Strong inhibiting	144	1		144	1	
Weak inhibiting	127	1.8 (1.21 – 2.6)	0.004	127	1.37 (0.96 – 2.0)	0.08
Non-inhibiting	165	1.7 (1.16 – 2.4)	0.006	165	1.28 (0.92 – 1.8)	0.15
KIR3DL1 – Bw4 (HLA-A and -B)						
Strong inhibiting	144	1		144	1	
Weak inhibiting	127	1.77 (1.2 – 2.6)	0.004	127	1.37 (0.96 – 2.0)	0.08
Non-inhibiting	172	1.63 (1.13 – 2.4)	0.009	172	1.24 (0.89 – 1.7)	0.2
HLA-A*24 and A*32	42	0.59 (0.28 – 1.3)	0.168	42	0.73 (0.4 – 1.3)	0.31
KIR3DL1 and KIR2DS1						
Strong + 2DS1	38	1		38	1	
Weak + 2DS1	32	1.2 (0.49 – 3.1)	0.659	32	0.9 (0.42 – 1.9)	0.78
None + 2DS1	47	2.0 (0.91 – 4.4)	0.086	47	1.16 (0.59 – 2.3)	0.66
Strong without 2DS1	106	1.6 (0.75 – 3.3)	0.23	106	1.14 (0.63 – 2.1)	0.66
Weak without 2DS1	95	3.1 (1.53 – 6.3)	0.002	95	1.84 (1.01 – 3.4)	0.046
None without 2DS1	118	2.5 (1.25 – 5.2)	0.01	118	1.54 (0.86 – 2.8)	0.15

		OS		TRM		
		<i>Adjusted for Karnofsky score, disease state, disease type, presence of comorbidities</i>		<i>Adjusted for Karnofsky score, disease type, transplant center, source of transplant</i>		
Variable	<i>n</i>	HR (95%-CI)	<i>p</i>	<i>n</i>	HR (95%-CI)	<i>p</i>
KIR3DL1 – Bw4 (HLA-B)						
Strong inhibiting	144	1		144	1	
Weak inhibiting	127	1.39 (0.95 – 2.0)	0.09	127	0.8 (0.43 – 1.5)	0.49
Non-inhibiting	165	1.23 (0.85 – 1.8)	0.26	165	0.72 (0.4 – 1.28)	0.26
KIR3DL1 – Bw4 (HLA-A and -B)						
Strong inhibiting	144	1		144	1	
Weak inhibiting	127	1.4 (0.95 – 2.0)	0.09	127	0.8 (0.43 – 1.48)	0.47
Non-inhibiting	172	1.21 (0.84 – 1.7)	0.3	172	0.68 (0.38 – 1.2)	0.18
HLA-A*24 and A*32	42	0.82 (0.43 – 1.6)	0.57	42	0.69 (0.26 – 1.81)	0.45
KIR3DL1 and KIR2DS1						
Strong + 2DS1	38	1		38	1	
Weak + 2DS1	32	0.71 (0.32 – 1.6)	0.4	32	1.43 (0.46 – 4.47)	0.54
None + 2DS1	47	0.8 (0.39 – 1.6)	0.54	47	0.94 (0.31 – 2.82)	0.91
Strong without 2DS1	106	0.79 (0.42 – 1.5)	0.46	106	1.37 (0.55 – 2.43)	0.5
Weak without 2DS1	95	1.39 (0.75 – 2.6)	0.29	95	0.88 (0.32 – 2.47)	0.8
None without 2DS1	118	1.16 (0.63 – 2.1)	0.64	118	0.9 (0.34 – 2.37)	0.83

CI; confidence interval, GvHD; graft-versus-host disease, HR; hazard ratio, OS; overall survival, PFS; progression-free survival, TRM; transplant-related mortality

REFERENCES

1. Norman PJ, Hollenbach JA, Nemat-Gorgani N, et al. Defining KIR and HLA Class I Genotypes at Highest Resolution via High-Throughput Sequencing. *Am J Hum Genet.* 2016;99(2):375-91.
2. Gardiner CM, Guethlein LA, Shilling HG, et al. Different NK cell surface phenotypes defined by the DX9 antibody are due to KIR3DL1 gene polymorphism. *J Immunol.* 2001;166(5):2992-3001.
3. Pando MJ, Gardiner CM, Gleimer M, McQueen KL, Parham P. The protein made from a common allele of KIR3DL1 (3DL1*004) is poorly expressed at cell surfaces due to substitution at positions 86 in Ig domain 0 and 182 in Ig domain 1. *J Immunol.* 2003;171(12):6640-9.
4. Maxwell LD, Wallace A, Middleton D, Curran MD. A common KIR2DS4 deletion variant in the human that predicts a soluble KIR molecule analogous to the KIR1D molecule observed in the rhesus monkey. *Tissue Antigens.* 2002;60(3):254-8.
5. Bari R, Bell T, Leung WH, et al. Significant functional heterogeneity among KIR2DL1 alleles and a pivotal role of arginine 245. *Blood.* 2009;114(25):5182-90.
6. Bari R, Thapa R, Bao J, et al. KIR2DL2/2DL3-E(35) alleles are functionally stronger than -Q(35) alleles. *Sci Rep.* 2016;6:23689.
7. Robinson J, Halliwell JA, Hayhurst JD, et al. The IPD and IMGT/HLA database: allele variant databases. *Nucleic Acids Res.* 2015;43(Database issue):D423-31.
8. Hilton HG, Guethlein LA, Goyos A, et al. Polymorphic HLA-C Receptors Balance the Functional Characteristics of KIR Haplotypes. *J Immunol.* 2015;195(7):3160-70.
9. Graef T, Moesta AK, Norman PJ, et al. KIR2DS4 is a product of gene conversion with KIR3DL2 that introduced specificity for HLA-A*11 while diminishing avidity for HLA-C. *J Exp Med.* 2009;206(11):2557-72.
10. Moesta AK, Graef T, Abi-Rached L, et al. Humans differ from other hominids in lacking an activating NK cell receptor that recognizes the C1 epitope of MHC class I. *J Immunol.* 2010;185(7):4233-7.
11. Naiyer MM, Cassidy SA, Magri A, et al. KIR2DS2 recognizes conserved peptides derived from viral helicases in the context of HLA-C. *Sci Immunol.* 2017;2(15).
12. Liu J, Xiao Z, Ko HL, Shen M, Ren EC. Activating killer cell immunoglobulin-like receptor 2DS2 binds to HLA-A*11. *Proc Natl Acad Sci U S A.* 2014;111(7):2662-7.
13. Foley BA, De Santis D, Van Beelen E, et al. The reactivity of Bw4+ HLA-B and HLA-A alleles with KIR3DL1: implications for patient and donor suitability for haploidentical stem cell transplantations. *Blood.* 2008;112(2):435-43.
14. Gumperz JE, Litwin V, Phillips JH, Lanier LL, Parham P. The Bw4 public epitope of HLA-B molecules confers reactivity with natural killer cell clones that express NKB1, a putative HLA receptor. *J Exp Med.* 1995;181(3):1133-44.
15. Saunders PM, MacLachlan BJ, Widjaja J, et al. The Role of the HLA Class I alpha2 Helix in Determining Ligand Hierarchy for the Killer Cell Ig-like Receptor 3DL1. *J Immunol.* 2021;206(4):849-60.
16. van der Ploeg K, Le Luduec JB, Stevenson PA, et al. HLA-A alleles influencing NK cell function impact AML relapse following allogeneic hematopoietic cell transplantation. *Blood Adv.* 2020;4(19):4955-64.

17. Blokhuis JH, Hilton HG, Guethlein LA, et al. KIR2DS5 allotypes that recognize the C2 epitope of HLA-C are common among Africans and absent from Europeans. *Immun Inflamm Dis.* 2017;5(4):461-8.
18. Martin MP, Gao X, Lee JH, et al. Epistatic interaction between KIR3DS1 and HLA-B delays the progression to AIDS. *Nat Genet.* 2002;31(4):429-34.
19. Amorim LM, Augusto DG, Nemat-Gorgani N, et al. High-Resolution Characterization of KIR Genes in a Large North American Cohort Reveals Novel Details of Structural and Sequence Diversity. *Front Immunol.* 2021;12:674778.