# Classification and diagnosis of iron overload

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## Abstract

Background and Objective. Iron overload is the result of many disorders and could lead to the development of organ damage and increased mortality. The recent description of new conditions associated with iron overload and the identification of the genetic defect of hereditary hemochromatosis prompted us to review this subject and to redefine the diagnostic criteria of iron overload disorders.

*Evidence and Information sources.* The material examined in the present review includes articles published in the journals covered by the Science Citation Index<sup>\*</sup> and Medline<sup>\*</sup>. The author has been working in the field of iron overload diseases for several years and has contributed ten of the papers cited in the references.

State of the art and Perpectives. Iron overload can be classified on the basis of different criteria: route of access of iron within the organism, predominant tissue site of iron accumulation and cause of the overload. Excess iron can gain access by the enteral route, the parenteral route, and placental route during fetal life. The different distribution of iron within parenchymal or reticuloendothelial storage areas indicates different pathogenetic mechanisms of iron accumulation and has relevant implications in terms of organ damage and prognosis of the patients. Iron overload may be either primary, resulting from a deregulation of intestinal iron absorption as in hemochromatosis or secondary to other congenital or acquired conditions. Diagnosis of iron overload can be suspected on the basis of clinical data, high transferrin saturation and/or serum ferritin values. However, several hyperferritinemic conditions are not related to iron overload, but may imply severe disorders (inflammations, neoplasia) or a deregulation of ferritin synthesis (hereditary hyperferritinemiacataract syndrome), and iron overload secondary to aceruloplasminemia, and the recently described dysmetabolic-associated liver iron overload syndrome, are characterized by low or normal transferrin saturation levels. Liver biopsy is still very useful in the diagnostic approach to iron overload disorders, by defining the amount and the distribution of iron within the liver. The analysis of HFE gene mutations (C282Y and H63D) is a simple and strong tool in the diagnostic work out of iron overload conditions. ©1998, Ferrata Storti Foundation

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Phone: international +39-39-2333361 • Fax: international +39-39-322274 • E-mail: alberto.piperno@unimi.it ron overload is the result of many disorders and could lead to the development of organ damage and increased mortality. The recent description of new conditions associated with iron overload and the identification of the genetic defect of hereditary hemochromatosis prompted us to review this subject and to redefine the diagnostic criteria of iron overload disorders.

# **Classification of iron overload**

Iron overload is the result of many disorders and could lead per se to the development of organ damage and increased mortality. In humans total body iron stores is maintained normally within the range of 200-1500 mg (in men, the normal concentration of iron in the storage pool is 13 mg/Kg and in women 5 mg/kg) by adequate adjustment of intestinal iron absorption, since no excretory mechanisms exist.<sup>1-4</sup> In normal individuals, feedback mechanisms inhibit iron absorption as storage iron increases.<sup>1</sup> Each condition that induces an increased net entry of iron within the body inevitably leads to iron overload.<sup>2</sup> Iron overload could be defined as an increase of storage iron, regardless of the presence or absence of tissue damage. It can be classified on the basis of different criteria: route of access of iron within the organism, predominant tissue site of iron accumulation and cause of the overload.

## Route of access of iron

Excess iron can gain access to the organism in three ways: the enteral route, through absorption of dietary heme and non-heme iron, the parenteral route, through transfusions or injections of iron-containing compounds, and placental route during fetal life (Table 1).

## Enteral route

Hereditary hemochromatosis (HHC) leads to absorption of excess dietary iron. Besides the recent discovery of the genetic defect in HHC, the pathogenic mechanism leading to inappropriately high iron absorption in this disorder is still undefined.<sup>4,5</sup> The excess of daily iron uptake is not large, but over time, due to the absence of iron excretory pathways in humans, substantial body overload develops.<sup>4</sup> In juvenile hemochromatosis, iron absorption is also increased, but the genetic as well as the biochemical defect of this kind of hemochromatosis is still

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unknown.<sup>6</sup> Ineffective erythropoiesis is associated with increased uptake of dietary iron, presumably in response to the increased demand from an expanded pool of erythrocyte precursors.<sup>1-3,7</sup> However, in most of such disorders transfusion therapy is needed; in that case the contribution of increased enteral absorption to iron overload is relatively small. Iron absorption is increased in the hemolytic state, although a significant iron overload (in the absence of over-transfusions) is commonly evident only when heterozygosity for HHC or ineffective erythropoiesis coexists.7-11 A puzzling finding is the apparent relationship between splenectomy and increased absorption and accumulation of iron in hemolytic patients.<sup>11-13</sup> Increased dietary iron from traditional beer brewed in steel drums has been regarded as the sole cause of iron overload in African populations, but recent data suggest that a genetically determined defect in the regulation of iron absorption exists in these patients and that this defect is necessary for the development of iron overload.<sup>14-16</sup> According to the model presented in that study, heterozygotes for the iron-loading locus develop iron overload only in the face of high dietary iron, but homozygotes may become iron-loaded without increased dietary iron.<sup>16</sup>

Similarly, increased iron absorption has been regarded as a cause of liver iron overload in chronic hepatic disorders<sup>17-20</sup> (see below). However, it is possible that the coexistence with heterozygous HHC may further increase iron absorption in these patients.<sup>17,20</sup>

Iron absorption is increased in the hypotransferrinemic mouse,<sup>21</sup> probably as the result of the iron deficient erythropoiesis. Although not proved, this probably occurs also in congenital hypotransferrinemia in humans. In systemic hemosiderosis associated with aceruloplasminemia,<sup>22,23</sup> the cause of iron overload does not primarily depend on defective regulation of iron absorption, but it is likely that the mechanisms that normally inhibit iron absorption as iron storage increases do not operate in this rare disease, due to the low serum iron levels secondary to the impaired iron release from cells to plasma. However, functional studies on iron metabolism in this human hereditary disorder have not yet been reported.

Cases of iron overload induced by prolonged oral administration of iron-containing compounds are exceptionally rare<sup>24</sup> and are probably the results of an interaction with other genetically or acquired disorders able to increase intestinal iron absorption.

## Parenteral route

Transfused blood is disposed through the macrophages of the reticuloendothelial (RE) system, which breaks down the hemoglobin of ingested erythrocytes.<sup>25</sup> Thus, large amounts of iron accumulate as clumps of hemosiderin within the RE cells and excess iron, by some means, makes its way into the extracellular fluid and in the plasma until the capacity of apotransferrin to take up iron becomes saturated. At this point excess iron is delivered to the hepatocytes and other parenchymas leading to the development of organ damage.<sup>1,2</sup>

Iron overload from parenteral administration of iron compounds is rare and is the result of inaccurate determination of body iron stores.<sup>26</sup>

#### Placental route

Throughout pregnancy, iron is taken up from maternal plasma by trophoblasts and transferred into the fetoplacental circulation.27 This process is thought to accelerate during the last trimester. In the fetus, the hepatocyte is the main site of storage iron. Thus, based on the tests used in adults, the term infant is markedly iron overloaded and iron stores may even increase within the first post-natal weeks.<sup>27</sup> It is then difficult to dissect whether systemic or liver diseases that develop during fetal or neonatal life are merely superimposed on an otherwise physiological iron overload status or they are the result of excess transplacental delivery of iron. Perinatal or neonatal hemochromatosis is a syndrome of severe idiopathic liver disease of fetal onset associated with marked hepatic and extra hepatic siderosis that is not a variant of HHC.<sup>28</sup> It has been argued that this disease is due to abnormal iron handling in the throphoblast and fetal liver, but no specific defect has yet been identified.27,28

#### Sites of iron accumulation

Parenchymal vs. RE iron overload

The different distribution of iron within parenchymal or RE storage areas indicates different pathogenetic mechanisms of iron accumulation and has relevant implications for organ damage and prognosis of the patients (Table 1). In fact, organ damage is related to the amount of iron present in the parenchymal cells, whereas iron within RE cells appears to be relatively innocuous.<sup>1,7</sup> Liver parenchymal iron overload is usually the result of excessive iron absorption by the enteral route, such as in HHC and anemias with ineffective erythropoiesis (iron loading anemias), but may also reflect enhanced internal redistribution of transfused erythrocyte iron recycled from the RE cells, as observed in the more advanced stage of transfusional iron overload.<sup>1,2,4,7</sup> In hypotrasferrinemic mice as well as in congenital hypotransferrinemia, tissue iron distribution is almost identical to that seen in HHC. There is no accumulation of iron in macrophages either in bone marrow or spleen.<sup>21</sup> This indicates that transferrin is not required for the recycling of iron by macrophages and that any conditions, genetic or environmental, that lowers plasma apotransferrin levels (e.g. advanced liver cirrhosis) will result in increased parenchymal iron uptake and eventual tissue damage.<sup>17,19,21</sup>

RE iron overload is the result of enhanced phagocytosis of red cells (chronic hemolysis and transfusions) and uptake of iron-containing compound.<sup>1,25</sup> Table 1. Iron overload disorders divided according to route of iron access and site of predominant iron deposition.

Route of access	Predominant storage areas			
	RE	Parenchyma	Both	
Enteral route	Chronic hemolytic disorders	Hemochromatosis	African iron overload	
		Iron-loading anemias	Chronic hepatic disorders	
		Juvenile hemochromatosis	Ceruloplasmin deficiency	
		Congenital hypotransferrinemia		
Parenteral route	Transfusional (early stage)		Transfusional	
	lron-containing compounds			
Placental route		Perinatal hemochromatosis		

Chronic hemolytic disorders: hereditary spherocytosis, stomatocytosis, and pyruvate kinase deficiency (9-11). Iron loading anemias: thalassemia, congenital and acquired sideroblastic anemias, congenital and acquired dyserythropoietic anemias (1,2). Chronic hepatic disorders: alcoholic liver disease, chronic viral hepatitis, end-stage liver cirrhosis, porto-caval shunts, porphyria cutanea tarda, non-alcoholic liver steatohepatitis, dysmetabolicassociated liver iron overload (17,19,48,49).

In the liver, iron may accumulate in Kupffer cells because of phagocytosis of dead iron-laden hepatocytes (sideronecrosis) or uptake of ferritin and hemosiderin from damaged hepatocytes.<sup>17,25,29,30</sup> In inflammatory disorders, iron accumulates in macrophages because of the *RE iron block*; this disease state is characterized by a low to normal transferrin saturation and a progressive increase in serum ferritin and marrow hemosiderin with time.<sup>1,25</sup>

In African and African-American iron overload, iron deposition is similar to that seen in trasfusional iron overload: iron is deposited equally in the spleen, bone marrow and both liver parenchymal cells and macrophages.<sup>14-16</sup> A full explanation of the accumulation of iron in Kupffer cells and macrophages in this form of iron overload is still lacking. In both aceruloplasminemia, and ascorbic acid deficiency there is an impaired delivery of iron from iron-storing cells.<sup>1,23,25</sup> Ascorbic acid deficiency may occur as a consequence of massive iron overload and would inhibit either iron transfer to plasma or entry of iron into ferritin, but does not influence the total amount of body iron.<sup>18,20</sup>

By contrast, aceruloplasminemia is the cause of parenchymal and RE hemosiderosis.<sup>22,23</sup> Ceruloplasmin catalyzes the oxidation of ferrous iron to the ferric form which combines to apotransferrin to form transferrin. The lack of the ferroxidase activity of ceruloplasmin would therefore results in impairment of iron release that explains both low serum iron levels and systemic iron accumulation.<sup>22,23</sup>

#### Systemic vs. focal iron overload

The most typical and common iron overload disorders, if left untreated, cause progressive iron accumulation in the liver and then in other tissues and organs leading to a multi-visceral disease.<sup>2</sup> However, as mentioned before, the pattern of body iron distribution may vary in different iron overload disorders. Gonads and brain are generally preserved from iron accumulation since they are sheltered from circulating transferrin.<sup>31</sup> Conversely, in aceruloplasminemia, in which iron overload depends on a defect on cellular export of iron, brain is a main site of iron accumulation together with liver and pancreas.<sup>23</sup> Accordingly, the clinical features are characterized mainly by progressive dementia, extrapyramidal symptoms, cerebral ataxia and diabetes mellitus.<sup>23</sup>

Iron accumulation can be confined to a specific organ. In several chronic hepatic disorders (alcoholic liver disease, chronic viral hepatitis, porto-caval shunts, liver cirrhosis, porphyria cutanea tarda), mild to severe iron overload is frequent in the liver, both in parenchyma and RE cells, but iron deposits are minimal in extra hepatic sites.<sup>17,19,30,32,33</sup> In Gaucher's disease, there is a marked iron accumulation in Gaucher's cells.<sup>34</sup> In late stages of Alzheimer's disease and other degenerative neurological disorders, excessive amounts of iron have been found in some areas of the brain, where iron appears to be a component of neuronal cell death.<sup>35</sup> In some rare conditions, such as idiopathic pulmonary hemosiderosis, renal hemosiderosis and superficial siderosis of the central nervous system, iron accumulation is the results of sequestration and degradation of hemoglobin from damaged erythrocytes by macrophages because of inflammatory, hemolytic or hemorrhagic processes strictly confined to a specific organ.<sup>3,36,37</sup>

#### Causes of iron overload

From a general point of view, iron overload can be classified as primary or secondary depending whether it results from a primary defect in the regulation of iron balance or is secondary to other genetic or acquired disorders. In some cases the specific cause of iron overload has already been defined, whereas in others the mechanism leading to iron overload is not yet elucidated and these iron overload disorders remain idiopathic.

#### Primary iron overload

The best-known example of primary iron overload is HHC, in which iron is absorbed in excess because of increased iron transfer from the enteral cells to the blood.<sup>2,4</sup> Recent advances in molecular genetics, clinical aspects and diagnosis of HHC have been previously reviewed in this journal.<sup>38</sup> A new gene, now called HFE, has been isolated and two *missense* mutations, C282Y and H63D, have been identified.<sup>5</sup> Whereas C282Y mutation in the homozygous state seems undoubtedly sufficient to cause HHC, some confu

sion exists regarding the H63D variant.<sup>5,38,39</sup> However, several data prove the relationship of the H63D mutation to the disease and that compound heterozygotes of this mutation with the C282Y variant are at special risk for the development of HHC, but with a low penetrance and milder phenotype expression.<sup>39</sup>

A variable proportion of patients with a HHC phenotype did not show any mutation in the HFE gene in different populations.<sup>5,40-42</sup> It is hypothesized that some of these patients may represent forms of non-HFE related hemochromatosis in which a primary defect, different from that observed in HHC, exists. Accordingly, recent data showed that juvenile hemochromatosis is not linked to HFE gene,<sup>6</sup> and that African iron overload is caused by an interaction between high dietary iron content and a common iron-loading gene not linked to the HLA locus.<sup>16</sup> These results suggest that defects of other still undefined genes may be responsible for primary derangement of iron metabolism in humans.

#### Secondary iron overload

This group includes iron overload either due to or associated with ineffective erythropoiesis, chronic liver diseases, parenteral administration or ingestion of excessive amounts of iron. In some of these disorders it is possible, as previously mentioned, that interactions with causes of primary iron overload exist.

Thalassemia major and sideroblastic anemia are the two best studied examples of iron overload secondary to blood transfusions and ineffective erythropoiesis. Because abnormalities in hemoglobin can decrease erythrocyte life span, the pool of erythrocyte precursors is markedly expanded in certain hemoglobinopathies, leading to increased enteral absorption of dietary iron.<sup>2,3,7</sup>

Aggressive transfusion therapy suppresses endogenous erythopoiesis and corrects the severe anemia, but leads to its own complications, the worst of which is iron overload.<sup>43</sup> Sideroblastic anemias are a heterogenous group of inherited and acquired hematopoietic disorders characterized by the association of anemia with the presence of non-heme nonferritin iron deposits within the mitochondria of erythroid precursors in the bone marrow (ringed sideroblasts). It has been generally assumed that mitochondrial iron deposits are secondary to the failure of heme synthesis that would lead to ineffective erythopoiesis and increased iron absorption.<sup>3,7,44</sup>

The main causes of iron overload in chronic hepatic diseases are shown in Table 2. In these cases liver iron overload is frequent, often mild or moderate, and iron deposits are usually found in both Kupffer cells and hepatocytes.<sup>2,17,19,30,33</sup> In these circumstances, even relatively low amounts of iron may amplify and propagate the initial toxic effect of alcohol and viruses, with rapid acceleration of the fibrotic response in the liver.<sup>17</sup> In porphyria cutanea tarda, liver iron overload is the result of an interaction

#### Table 2. Causes of iron overload in chronic hepatic disorders.

Related to high alcohol intake	Related to hepatocyte damage	Related to chronic liver failure
Alcohol-induced increased iron absorption	Increased iron and ferritin release to extracellular fluid and plasma	Reduced transferrin synthesis
Increased ability of desialylated transferrin to deliver iron to the liver	Cytokine-mediated increased hepato- cellular iron uptake	Spontaneous or surgically- constructed porto- systemic shunts
Chronic hemolysis		Chronic hemolysis
Ineffective erythropoiesis		Ineffective erythropoiesis

References: (17,19,30,34,48,61)

between several factors: heavy alcohol intake, chronic viral hepatitis and heterozigosity for the HFE gene mutations.<sup>45,46</sup> Iron overload in end-stage liver cirrhosis is a rather frequent condition characterized by a continuum from slight and spotty iron deposition to diffuse and marked siderosis.<sup>19,47</sup> This post cirrhosis siderosis seems to be acquired rather than genetically determined. However, data on HFE gene mutations in these patients are lacking.

A new syndrome of liver iron overload has been recently described as generally modest, associated with increased ferritin, normal transferrin saturation and presence of metabolic disorders.<sup>48</sup> The mechanism leading to parenchymal iron overload in these patients is still undefined.

## **Diagnosis of iron overload**

Iron overload is suspected when biochemical iron indices are increased and confirmed by the demonstration of increased iron deposits either by liver biopsy, magnetic resonance imaging (MRI) and spectroscopy or subsequent quantum interference device (SQUID), or by retrospective evaluation of total iron removed by phlebotomy.<sup>38,43,49</sup> The differential diagnosis among the different causes of iron overload could be difficult and should take into account clinical, biochemical and histological data. The recent identification of the HFE gene mutations added a simple and strong tool in the differential diagnostic strategy of iron overload conditions. Figure 1 illustrates a practical approach to the diagnosis of iron overload disorders.

# **Biochemical iron indices**

Three iron markers can be considered: serum iron, transferrin saturation and serum ferritin. Serum iron

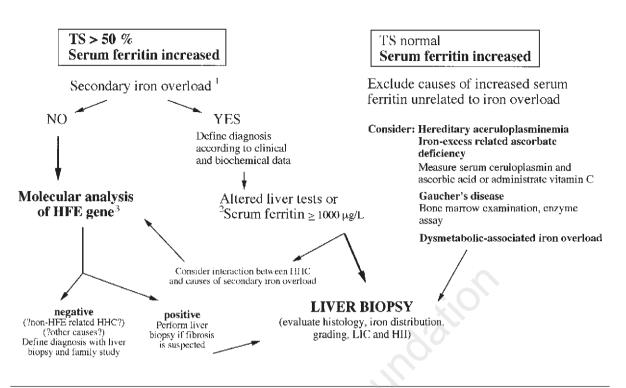


Figure 1. Diagram illustrating a practical approach to the differential diagnosis of iron overload disorders. Legend: 1. iron loading anemias, transfusional iron overload, chronic hemolytic disorders; 2. this level indicate an increased risk of liver fibrosis, but lower levels do not exclude it; 3. see Table 4)

has no value *per se* in the diagnosis of iron overload, but it remains an obligatory index because it is necessary for measuring transferrin saturation. Approaching the diagnosis of iron overload disorders, one should consider that some iron indices might be elevated in non-iron overloaded situations. Moreover, in some iron-overload disorders transferrin saturation and serum ferritin may behave differently in relationship to the different mechanism leading to iron overload. Figure 2 schematically shows the different mechanisms leading to high transferrin saturation and/or serum ferritin values in iron overload disorders.

#### Transferrin saturation

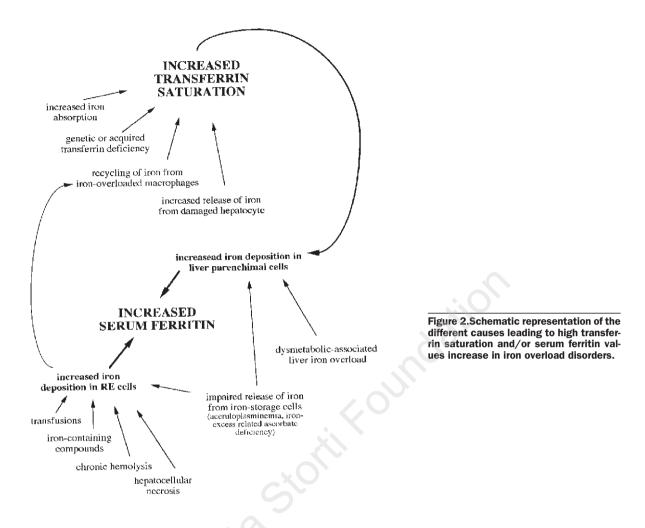
It corresponds to the ratio of serum iron and totaliron binding capacity. However, in most laboratories, total-iron binding capacity is not directly measured, but is deduced from serum transferrin concentration, after correction for a constant value (mg of transferrin multiplied by 1.24 or 1.25).<sup>27,50</sup> Thus, there is a need for homogeneity among different laboratories.

Transferrin saturation is influenced by the same variations of serum iron (within day and day to day variability and inflammation), which can limit its clinical usefulness.<sup>25</sup> In *non-iron overloaded* conditions transferrin saturation might be high in presence of liver dysfunction, due to increased serum iron (through hepatocellular necrosis) and decreased

transferrin synthesis (through liver failure).<sup>17,19,25</sup> However, with time this condition may be a cause of hemosiderosis secondary to liver cirrhosis, due to the increase of non-transferrin bound iron in the plasma (see first section of the review).<sup>19,21</sup> In *iron overload* situations transferrin saturation is usually elevated (> 50%) before serum ferritin increases. It can be lowered by ascorbic acid deficiency (iron-excess related),<sup>18,20</sup> is low in hereditary aceruloplasminemia,<sup>23</sup> normal in the recently described iron overload syndrome associated with dysmetabolic disorders<sup>48</sup> and in Gaucher's disease (see below).<sup>34</sup>

#### Serum ferritin

It is increased in a number of *non-iron overloaded* situations, including infections, acute and chronic inflammatory disorders, hepatocellular necrosis, alcohol abuse (through hepatocellular necrosis and increased ferritin synthesis), and deregulation of L-ferritin synthesis (hereditary hyperferritinemia-cataract syndrome).<sup>1,2,51</sup> In *iron overloaded* situations serum ferritin is usually higher than the upper normal limits corrected for sex and age (Table 3) and is most often associated with high transferrin saturation,<sup>2,4</sup> excluding those situations previously mentioned.<sup>18,20,23,34,48</sup> In dysmetabolic associated liver iron overload, serum ferritin levels overestimate body iron stores.<sup>48</sup> Serum ferritin concentrations are usually related to the amount of body iron stores, but a variety of condi-



tions that may frequently associate with iron overload, may reduce (ascorbate deficiency), or increase serum ferritin levels (hepatocellular necrosis either ironinduced or related to coexistent alcohol abuse or viral infections), independently of changes of body iron burden.<sup>17,18</sup> In hemochromatosis, ferritin concentrations above 1000 µg/L suggests liver damage (fibrosis or cirrhosis).<sup>29</sup> The same level is associated with an increasing risk of developing iron-induced complications in thalassemia major.<sup>43</sup>

Table 3. Upper normal values of serum ferritin corrected for sex and age (reported from Adams *et al.* $^{62}$ )

Age	Women	Men
10-19 years	40 µg/L	100 µg/L
20-29 years	65 µg/L	350 µg/L
30-39 years	80 µg/L	350 µg/L
40-49 years	100 µg/L	350 µg/L
> 50 years	200 µg/L	350 µg/L

## Assessment of iron overload

#### Liver biopsy

It is still essential in the diagnostic pathway of iron overload disorders. Liver biopsy ascertains iron overload, defines its distribution within the hepatic lobules, provides a semi-quantitative evaluation of iron excess by different grading systems,<sup>29,52</sup> enables the quantitation of iron by measurement of liver iron concentration (LIC), and informs on the degree of tissue iron-dependent damage and associated lesions (i.e., alcoholism, chronic viral hepatitis, steatosis). The distribution of iron may vary in different iron overload disorders and is related to the underlying mechanisms responsible for iron accumulation (see also paragraph Site of iron accumulation). Sometimes, liver iron distribution can be more useful that LIC in differential diagnosis of iron overload disorders. Nonhomogeneous iron deposition within the liver without intrabiliary iron deposits is the typical pattern of iron overload associated with alcoholic and posthepatitis cirrhosis.<sup>19,47</sup> In HHC as well as in iron overload secondary to ineffective erythropoiesis, iron is deposited in the liver in a lobular distribution with a decreasing periportal-to-pericentral gradient,<sup>2,4,29</sup>

Table 4. Relationship between HFE genotype and phenotype. Usefulness of HFE gene analysis in the diagnosis of hemochromatosis (data derived from the *Proceedings of the International Symposium on Iron in Biology and Medicine*, S.Malo, France, June 16-20, 1997, refs. #5, 40, 42, and Piperno *et al*, submitted)

Genotype	Phenotype	
C282Y homozygotes	Hemochromatosis with a wide range of expression related to both genetic and acquired factors. About 30% of women and 5% of men may not express the disease due to physiological and pathological blood losses.	
C282Y heterozygotes	Generally normal. Rarely may express a hemochromatosis phenotype, more fre- quently if other factors able to increase iron absorption are present.	
C282Y/H63D compound heterozygotes	May express a hemochromatosis pheno- type, generally mild, with a low penetration	
H63D heterozygotes or homozygotes	Generally normal. Frequent in patients with porphyria cutanea tarda and chronic viral hepatitis with mild/moderate iron overload in Italy. Rarely associated with a hemochro- matosis phenotype (see C282Y heterozy- gotes)	
Wild-type	Generally normal. Some cases with a hemochromatosis phenotype have been described suggesting the existence of non- HFE related hemochromatosis	

whereas in transfusional and African iron overload, iron deposition is panlobular and typically involves Kupffer cells and portal macrophages.<sup>14,15</sup> The upper normal limit of LIC ranges between 30 to 39  $\mu$ Mol/g in men and 18 to 28 µMol/g in women, in different studies.<sup>29,30,47</sup> In classical forms of iron overload either primary o secondary, LIC is generally higher that 90 µMol/g in men and 60 µMol/g in women. The hepatic iron index (HII), defined by Bassett et al.53 as the LIC to age (years) ratio, has been widely accepted as a tool for differentiating HHC patients from either heterozygotes and patients with alcoholic siderosis. Currently, many authors assume that in the absence of a cause of secondary iron overload, a HII greater than 1.9 or 2 is suggestive of HHC. However, this assumption is questionable when advanced cirrhosis is present, due to the wide intrahepatic variability of iron concentrations found in this condition.<sup>19</sup>

#### Other methods

The amount of iron removed by venesection (IR) is a reliable retrospective method for defining the amount of iron overload, but it gives no further information useful for differentiating among various iron overload disorders. In normal subjects IR is below 1.5 g, whether in major iron overload situations is generally higher than 5 g.<sup>1,2</sup>

Many studies show that MRI can reflect the presence of tissue iron *in vivo*, but this method has not been validated as one that provides measurements of tissue iron that are quantitatively equivalent to those determined at liver biopsy.<sup>43</sup> Conversely, SQUID is a rapid, safe, reliable and quantitative measurement of liver iron overload, but the instrumentation for this technique is scarcely available. However, none of these techniques provide information regarding iron distribution and histological findings.

#### Molecular analysis of HFE gene

In the presence of increased biochemical iron indices, once that major causes of secondary iron overload has been excluded, searching for the two HFE gene mutations should be the next and simplest diagnostic step. Homozygosity for the C282Y mutation is generally associated with expressed HHC with few exceptions.<sup>5,39,41,42</sup> Table 4 shows the relationship between HFE genotypes and phenotypes, as deduced from data presently available (refs #5, 39, 41, Proceedings of the International Symposium on Iron in Biology and Medicine, S. Malo, France, June 16-20, 1997, and ref. #54). HFE analysis allows to distinguish between HHC and chronic hepatic disorders with iron overload, or to define such patients in which both conditions exist. This is particularly useful in those countries, such as in Italy, where both heavy alcohol intake and hepatitis viral infections are frequent and may aggravate the prognosis of HHC patients.<sup>30,55</sup> A preliminary study indicates that the coexistence of a single HFE mutation with  $\beta$ -thalassemia trait, alcohol abuse, or chronic liver hepatitis, alone or in combinations, may give rise to a HHC phenotype<sup>54</sup> but further studies are needed to validate these observations.

# **Future directions**

The discovery of the HFE gene mutations provides the opportunity for a more precise classification of those heterozygous and homozygous for HHC. Moreover, HFE molecular testing will allow the clarification of the role of both mutations in some iron overload conditions of uncertain origin. It has been previously suggested that the heterozygous state for HHC may favour the development of significant iron overload in several diseases such as chronic hemolytic disorders, porphyria cutanea tarda, hemoglobinopathies, or chronic liver diseases<sup>8,9,11,30,38,45,46</sup> and that HHC heterozygotes may be at an increased risk of malignancies.<sup>56</sup> The availability of the molecular analysis of HFE gene is an important tool to test some of these associations.

The causes of iron overload remain to be clarified in several disorders (e.g. African-American iron overload, dysmetabolic associated iron overload, perinatal and juvenile hemochromatosis, post-cirrhosis hemosiderosis) and is hypothesized that both acquired and genetic factors are involved. Some studies suggest the existence of forms of genetically determined, non HFErelated hemochromatosis<sup>6,16,38</sup> that await the identification of their own molecular defects. Genes other than HFE are likely to be involved in the regulation of iron absorption, as recently shown by the cloning of iron transporters in mammalians.<sup>57,58</sup>

The recent description of a new condition associated with liver iron overload that seem to be related to metabolic disorders,48 open new perspectives on the possible link between iron excess and atherosclerotic and cardiovascular disorders<sup>59</sup> and raises guestions about the mechanisms of parenchymal iron loading in the presence of normal transferrin saturation. Further studies are needed to clarify the mechanisms leading to iron overload in patients with chronic viral hepatitis and end-stage liver disease and to understand the role of HFE gene in these disorders. In addition, based on the observation that increased liver iron concentrations may favor the progression to liver fibrosis in chronic viral hepatitis and negatively influence the response to interferon therapy, 17,33,60 careful studies and follow-up are needed to clarify whether early identification and treatment of iron overload may have a beneficial effect in the natural history of these common disorders.

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## References

- 1. Finch C, Huebers H. Perspectives in iron metabolism. N Engl J Med 1982; 306:1520-8.
- 2. Halliday J, Powell L. Iron overload. Semin Hematol 1982; 19:42-53.
- Pollycove M. Iron overload syndromes. Clin Physiol Biochem 1985; 4:61-77.
- Powell L, Jazwinska E, Halliday J. Primary iron overload. In: Brock, Halliday, Pippard and Powell, eds. Iron metabolism in health and disease. London: Saunders, 1994. p. 227-70.
- Feder J, Gnirke A, Thomas W, et al. A novel MHC class I-like gene is mutated in patients with hereditary haemochromatosis. Nature Genet 1996; 13:399-408.
- Camaschella C, Roetto A, Ciciliano M, et al. Juvenile and adult hemochromatosis are distinct genetic disorders. Eur J Hum Genet 1997; 5:371-5.
- 7. Pootrakul P, Kitcharoen K, Yansukon P, et al. The effect of erythroid hyperplasia on iron balance. Blood 1988; 71:1124-9.
- Fargion S, Piperno A, Panajotopoulos N, Taddei M, Fiorelli G. Iron overload in subjects with beta-thalassemia trait: role of the hemochromatosis gene. Br J Haematol 1985; 61:487-90.
- Fargion S, Cappellini M, Piperno A, Panajotopoulos N, Ronchi G, Fiorelli G. Association of hereditary spherocytosis and idiopathic hemochromatosis. A sinergistic effect in determining iron overload. Am J Clin Pathol 1986; 86:645-9.

- Stewart G, Amess J, Eber S, et al. Thrombo-embolic disease after splenectomy for hereditary stomatocytosis. Br J Haematol 1996; 93:303-10.
- Zanella Á, Berzuini A, Colombo M, et al. Iron status in red cell pyruvate kinase deficiency: study of Italian cases. Br J Haematol 1993; 83:485-90.
- Piperno A, Taddei M, Petrella G, Fiorelli G. Iron overload in hereditary spherocytosis. Haematologica 1984; 69:90-1.
- Parkin J, Rush B, Degroot R, Budd R. Iron absorption after splenectomy in hereditary spherocytosis. Aust NZ J Med 1974; 4:58-61.
- 14. Bacon B. Causes of iron overload. N Engl J Med 1992; 326:126-7.
- 15. Baer D. Hereditary iron overload in African Americans. Am J Med 1996; 100:5-8.
- Gordeuk V, Mukiibi J, Hasstedt S, et al. Iron overload in Africa. Interaction between a gene and dietary iron content. N Engl J Med 1992; 326:95-100.
- 17. Bonkovsky H, Banner B, Lambrecht R, Rubin R. Iron in liver diseases other than hemochromatosis. Semin Liver Dis 1996; 16:65-82.
- Chapman R, Hussain M, Gorman A, et al. Effect of ascorbic acid deficiency on serum ferritin concentration in patients with β-thalassemia major. J Clin Pathol 1982; 35:487-91.
  Deugnier Y, Turlin B, Le Quilleuc D, et al. A reappraisal
- Deugnier Y, Turlin B, Le Quilleuc D, et al. A reappraisal of hepatic siderosis in patients with end-stage cirrhosis: practical implications for the diagnosis of hemochromatosis. Am J Surg Pathol 1997; 21:669-75.
- 20. Tagliabue A, Turconi G, Allegrini M, et al. Ascorbic acid status in thalassemia major. Haematologica 1984; 69:542-8.
- Kaplan J, Craven C, Alexander J, Kushner J, Lamb J, Bernstein S. Regulation of the distribution of tissue iron. Lessons learned from the hypotransferrinemic mouse. In: Weintraub, Edwards and Krikker, eds. Hemochromatosis. Proceedings of the First International Conference. New York: The New York Academy of Sciences, 1988. p. 124-35.
- 22. Cox D. Genes of the copper pathways. Am J Hum Genet 1995; 56:828-34.
- 23. Yoshida K, Furihata K, Takeda S, et al. A mutation in the ceruloplasmin gene is associated with systemic hemosiderosis in humans. Nature Genet 1995; 9:267-72.
- Johnson BF. Hemochromatosis resulting from prolonged oral iron therapy. N Engl J Med 1968; 278: 1100-1.
- 25. Deiss A. Iron metabolism in reticuloendothelial cells. Semin Hematol 1983; 20:81-90.
- Saven A, Beutler E. Iron overload after prolonged intramuscolar iron therapy. N Engl J Med 1989; 321: 331-2.
- Knisely A. Iron and pediatric liver disease. Semin Liv Dis 1994; 14:229-35.
- Silver M, Beverley D, Valberg L, Cutz E, Phillips J, Shaheed W. Perinatal hemochromatosis: clinical, morphologic, and quantitative iron studies. Am J Pathol 1987; 128:538-54.
- 29. Deugnier YM, Loreal O, Turlin B, et al. Liver pathology in genetic hemochromatosis: a review of 135 homozygous case and their bioclinical correlation. Gastroenterology 1992; 102:2050-9.
- Piperno A, D'Alba R, Fargion S, et al. Liver iron concentration in chronic hepatitis: a study of 98 patients. Eur J Gastroenterol Hepatol 1995; 7:1203-8.
- Huggenvik J, Craven C, Idzerda R, Bernstein S, Kaplan J, McKnight G. A splicing defect in the mouse transferrin gene leads to congenital atransferrinemia. Blood 1989; 74:482-6.
- 32. Adams P, Bradley C, Frei J. Hepatic iron and zinc con-

centrations after portacaval shunting for nonalcoholic cirrhosis. Hepatology 1993; 19:101-5.

- Beinker N, Voigt M, Arendse M, Smit J, Stander I, Kirsch R. Threshold effect of liver iron content on hepatic inflammation and fibrosis in hepatitis B and C. J Hepatol 1996; 25:633-8.
- Lee R, Balcerzak S, Westerman M. Gaucher's disease: a morphologic study and measurements of iron metabolism. Am J Med 1967; 42:891-8.
- Connor J, Menzies S, St. Martin S, Mufson E. A histochemical study of iron, transferrin, and ferritin in Alzheimer's diseased brain. J Neurosci Res 1992; 31:75-83.
- Fearnley J, Stevens J, Rudge P. Superficial siderosis of the central nervous system. Brain 1995; 118:1051-66.
- Soergel K, Sommers S. Idiopathic pulmonary hemosiderosis and related syndromes. Am J Med 1962; 32:499-511.
- Camaschella C, Piperno A. Hereditary hemochromatosis: recent advances in molecular genetics and clinical management. Haematologica 1997; 82:77-84.
- Beutler E. The significance of the 187G (H63D) mutation in hemochromatosis. Am J Hum Genet 1997; 61:762-4.
- Borot N, Roth M, Malfroy L, et al. Mutations in the MHC class I-like candidate gene for hemochromatosis in French patients. Immunogenetics 1997; 45:320-4
- 41. Carella M, D'Ambrosio L, Totaro A, et al. Mutation analysis of the HLA-H gene in Italian hemochromatosis patients. Am J Hum Genet 1997; 60:828-32.
- Jouanolle A, Gandon G, Jézéquel P, et al. Hemochromatosis and HLA-H. Nature Genet 1996; 24:251-2.
- 43. Olivieri N, Brittenham G. Iron-chelating therapy and the treatment of thalassemia. Blood 1997; 89:739-61.
- 44. Ponka P. Tissue-specific regulation of iron metabolism and heme synthesis: distinct control mechanisms in erythroid cells. Blood 1997; 89:1-25.
- 45. Roberts A, Whatley S, Morgan R, Worwood M, Elder G. Increased frequency of the haemochromatosis Cys282Tyr mutation in sporadic porphyria cutanea tarda. Lancet 1997; 349:321-3.
- Sampietro M, Piperno A, Lupica L, et al. High prevalence of the Hys63Asp HFE mutation in Italian patients with porphyria cutanea tarda. Hepatology 1998; 27:181-4.
- 47. Ludwig J, Hashimoto E, Porayko M, Moyer T, Baldus

W. Hemosiderosis in cirrhosis: a study of 447 native livers. Gastroenterology 1997; 112:882-8.

- Moirand R, Mortaji A, Loreal O, Paillard F, Brissot P, Deugnier Y. A new syndrome of iron overload with normal transferrin saturation. Lancet 1997; 349:95-8.
- 49. Dixon R, Styles P, Al-Refaie F, et al. Assessment of hepatic iron overload in thalassemic patients by magnetic resonance spectroscopy. Hepatology 1994; 19:904-9.
- 50. Tsung S, Rosenthal W, Milewski K. Immunological measurement of transferrin compared with chemical measurement of total iron-binding capacity. Clin Chem 1975; 21:1063-6.
- Girelli D, Olivieri O, De Franceschi L, Corrocher R, Bergamaschi G, Cazzola M. A linkage between hereditary hyperferritinemia not related to iron overload and autosomal dominant congenital cataract. Br J Haematol 1995; 90:931-4.
- 52. Scheuer PJ, Williams R, Muir AR. Hepatic pathology in relatives of patients with hemochromatosis. J Pathol Bacteriol 1962; 84:53-64.
- Bassett ML, Halliday JW, Powell LW. Value of hepatic iron measurements in early hemochromatosis and determination of the critical level associated with fibrosis. Hepatology 1986; 6:24-9.
- Piperno A, Sampietro M, Pietrangelo A, et al. Heterogeneity of hemochromatosis in Italy. Gastroenterology 1998; in press.
  Piperno A, Fargion S, D'Alba R, et al. Liver damage in
- Piperno A, Fargion S, D'Alba R, et al. Liver damage in Italian patients with hereditary hemochromatosis is highly influenced by hepatitis B and C virus infection. J Hepatol 1992; 16:364-8.
- 56. Nelson R, Davis F, Persky V, Becker E. Risk of neoplastic and other diseases among people with heterozigosity for hereditary hemochromatosis. Cancer 1995; 76:875-9.
- Fleming M, Trenor III C, Su M, et al. Microcytic anemia mice have a mutation in Nramp2, a candidate iron transporter gene. Nature Genet 1997; 16:383-6.
- Gunshin H, Mackenzie B, Berger U, et al. Cloning and characterization of a mammalian proton-coupled metal-ion transporter. Nature 1997; 388:482-8.
- Conrad M. Excess iron and catastrophic illness. Am J Haematol 1993; 43:234-6.
- 60. Piperno A, Sampietro M, D'Alba R, et al. Iron stores, response to  $\alpha$ -interferon therapy, and effects of iron depletion in chronic hepatitis C. Liver 1996; 16:248-54.