

4. SPECIAL CONDITIONS

SPATIAL IMAGING UNLOCKS THE POTENTIAL OF CHARTING MULTIPLE MYELOMA AND EXTRAMEDULLARY DISEASE**V. Desantis¹, A. Andriano¹, G. De Martino¹, T. Düking², O. Hartwig², G. Ingravallo⁴, M. Biondo⁴, C. Botta⁵, R. Ria⁶, A. Vacca⁶, A. G. Solimando⁶**

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Multiple myeloma (MM) is a plasma cell (PCs) cancer marked by strong genetic, phenotypic, and microenvironmental heterogeneity. The emergence of extramedullary disease (EMD) signals an aggressive phase linked to treatment resistance and poor outcomes. The mechanisms driving plasma cells to escape the bone marrow and infiltrate extramedullary sites remain unclear. This study uses MACSima™ imaging cyclic staining (MICS), a high-dimensional spatial imaging technology, to map protein expression patterns and cellular interactions, revealing structural and network differences between bone marrow and EMD lesions. Using MICS, researchers stained 56 biomarkers across carefully prepared BM and EMD tissue samples from 5 newly diagnosed MM patients. Data from 231,186 cells were analysed, with 105,047 annotated as PCs. Computational tools processed and statistically validated the imaging data to generate spatial maps and cellular profiles. Distinct clustering patterns of PCs and immune infiltration profiles were identified via cell-type heatmaps and spatial distance analyses. This approach allowed for comprehensive 3D mapping of malignant PCs and key tumor microenvironment components, providing an integrated spatial framework for investigating cellular organization and intercellular interactions. MICS technology enabled the identification of 14 distinct cell types in BM and 9 in EMD, showcasing method's capacity to discern cellular diversity at a single-cell level. EMD lesions displayed disrupted architecture, characterized by a loss of stromal network integrity and diminished cross-talk with regulatory immune cells. Malignant PCs in EMD sites were organized into dense clusters, lacking canonical interactions with CD8⁺ T cells and dendritic cells, indicative of an immunologi-

cally permissive microenvironment (Figure1_EMN2026). PCs at EMD sites exhibited higher expression of drug resistance markers like BCL-2 and EZH2, and reduced CD38 expression, confirmed by immunohistochemistry. The spatial arrangement of immune cells such as T-cells and macrophages, was markedly different in EMD compared to BM. PCs were overall far distant from T-cells in EMD. This study provides the first evidence that spatial imaging is essential for understanding the structural complexity of MM and its progression to EMD. Three-dimensional reconstruction of cellular networks reveals the loss of immunostromal interactions and the formation of protective niches that sustain highly aggressive plasma cell subclones. This may explain the reduced effectiveness of CD38-targeted and immune-mediated therapies (such as CAR-T and bispecific antibodies) in EMD. Although larger studies are needed to confirm these findings, they offer valuable insights for precision medicine and microenvironment- focused strategies that could transform the treatment of MM with an extramedullary phenotype.

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