

Polytypic B cells, monotypic/monoclonal B-cell proliferations, and neoplastic T cells diverge from *TET2*-/*DNMT3A*-mutant clonal hematopoiesis in follicular helper T-cell lymphomas

Authors

Natasha E. Lewis,^{1*} Kseniya Petrova-Drus,^{1,2*} Rohan Sardana,¹ Sarah Huet,^{1°} Qi Gao,¹ Shenon Sethi,¹ Chad Vanderbilt,² Wenbin Xiao,¹ Mikhail Roshal,¹ Jeeyeon Baik,¹ Himanshu Bhurtel,¹ Alison J. Moskowitz,³ Steven M. Horwitz³ and Ahmet Dogan¹

¹Hematopathology Service, Department of Pathology and Laboratory Medicine; ²Diagnostic Molecular Pathology Service, Department of Pathology and Laboratory Medicine and ³Lymphoma Service, Department of Medicine, Memorial Sloan Kettering Cancer Center, New York, NY, USA

**NEL and KP-D contributed equally as first authors.*

°NEL current address: Division of Hematopathology, Department of Laboratory Medicine and Pathology, Mayo Clinic Arizona, Phoenix, AZ, USA

°SH current address: Laboratory of Haematology, CHU Lyon-Sud, Hospices Civils de Lyon, Pierre-Benite, France

Correspondence:

N.E. LEWIS - lewis.natasha@mayo.edu

A. DOGAN - dogana@mskcc.org

<https://doi.org/10.3324/haematol.2024.285183>

Received: January 31, 2024.

Accepted: July 12, 2024.

Early view: July 18, 2024.

©2025 Ferrata Storti Foundation

Published under a CC BY-NC license 

Polytypic B cells, monotypic/monoclonal B-cell proliferations, and neoplastic T cells diverge from *TET2*-/*DNMT3A*-mutant clonal hematopoiesis in follicular helper T-cell lymphomas

Natasha E. Lewis, Kseniya Petrova-Drus, Rohan Sardana, Sarah Huet, Qi Gao, Shenon Sethi, Chad Vanderbilt, Wenbin Xiao, Mikhail Roshal, Jeeyeon Baik, Himanshu Bhurtel, Alison J. Moskowitz, Steven M. Horwitz, Ahmet Dogan

Supplemental materials

Supplemental Table 1. Genomics of follicular helper T-cell lymphomas, polytypic B cells, monotypic/monoclonal B cell proliferations, and myeloid compartments in 25 follicular helper T-cell lymphoma patients

Supplemental Figure 1. Comparisons of the incidences and allele frequencies of shared and/or private mutations in polytypic B cells and monotypic/monoclonal B cell proliferations among various groups

Supplemental Figure 2. Comparison of mutations detected in samples containing high levels of polytypic B cells and monotypic/monoclonal B cell proliferations obtained from the same patients

Supplemental Table 1. Genomics of follicular helper T-cell lymphomas, polytypic B cells, monotypic/monoclonal B cell proliferations, and myeloid compartments in 25 follicular helper T-cell lymphoma patients																								
		T-Cell Lymphoma							B Cells/Monotypic/Monoclonal B Cell Proliferations							Myeloid Cells								
Patient	Variant	VAF	TCL Type	Genotyping Assay Utilized	Sample Type	Flow Sorted	Neoplastic T Cells in		VAF	B Cell Type	Genotyping Assay Utilized	EBV	Sample Type	Flow Sorted	Combined Myeloid and Neoplastic T Cells in Sample		IgH/TCR Gene Rearrangement (Method)	VAF	Myeloid Cell Type	Genotyping Assay Utilized	Sample Type	Flow Sorted	Combined B and Neoplastic T Cells in	
							Sample	TCR Gene Rearrangement							Cells in Sample	Rearrangement (Method)							Sample	TCR Gene Rearrangement (Method)
1	DNMT3A p.R882H TET2 p.M1333Nfs*6 TET2 p.G1145Vfs*7 RHOA p.G17L IDH2 p.R172K	0.52 0.49 0.45 0.51 0.54	TFHL-AI	MSK-IMPACT	BM	Y	NA	NT	0.49 0.10 0.00 0.00 0.00	Polytypic	MSK-IMPACT	Neg	BM	Y	NA	IgH: NT	0.46 0.06 0.00 0.00 0.00	Myeloid Cells	MSK-IMPACT	BM	Y	NA	NA	
2	DNMT3A p.W795C TET2 p.Y1337* RHOA p.G17V TET2 p.H1904Y	0.49 0.55 0.49 0.00	TFHL-AI	ddPCR*	BM	Y	NA	Pos	0.04 0.01 0.00 0.11	Polytypic	ddPCR*	Pos	BM	Y	NA	IgH: NT	0.07 0.00 0.00 0.00	Monocytes	ddPCR*	BM	Y	NA	NA	
3	TET2 p.Y592fs DNMT3A exon11 splicing variant (c.1279+1G>A) RHOA p.G17V IDH2 p.R172K	0.51 0.47 0.48 0.49	TFHL-AI	ddPCR*	PB	Y	NA	Pos^	0.06 0.03 0.00 0.00	Polytypic	ddPCR*	Pos	PB	Y	NA	IgH: NT	0.01 0.00 0.00 0.00	Monocytes	ddPCR*	PB	Y	NA	NA	
4	TET2 p.E1106Vfs*23 TET2 p.S280* RHOA p.G17V IDH2 p.R172K	0.56 0.44 0.46 0.46	TFHL-AI	MSK-IMPACT	LN	Y	NA	NT	0.02 0.00 0.00 0.00	Polytypic	MSK-IMPACT	Neg	LN	Y	NA	IgH: NT	NT							
5	TET2 p.P570fs RHOA p.G17V IDH2 p.R172G TET2 p.L873fs PIK3CG p.V74M	0.13 0.06 0.04 0.06 0.13	TFHL-AI	MSK-IMPACT	LN	N	10%	NT	0.15 0.00 0.00 0.00 NT NT	Polytypic	ddPCR*	NA	PB	Y	NA	IgH: NT	0.22 0.00 0.00 0.00 NT NT	Monocytes	ddPCR*	PB	Y	NA	NA	
6	TET2 p.C973fs TET2 p.R1216* RHOA p.G17V IDH2 p.R172S	0.07 0.08 0.05 0.06	TFHL-AI	MSK-IMPACT#	LN	N	10-15%	Pos	0.27 0.07 0.00 0.00	Polytypic	ddPCR*	Pos	PB	Y	NA	IgH: NT	0.00 0.00 0.00 0.00	Monocytes	ddPCR*	PB	Y	NA	NA	
7	TET2 N427Vfs*4 TET2 X1268_splice STAT5A p.N398K	0.47 0.44 0.47	TFHL-NOS	MSK-IMPACT#	LN	Y	NA	Pos	0.13 0.13 0.00	Polytypic	MSK-IMPACT#	Pos	LN	Y	NA	IgH: NT	0.43 0.47 0.00	Granulocytes	MSK-IMPACT#	BM	Y	NA	NA	
8	TET2 p.Q1537* TET2 p.N1610fs*6 DNMT3A p.L681M TP53 p.MA160IS	0.12 0.11 0.10 0.01	TFHL-AI	MSK-IMPACT#	LN	N	20%	Pos^	0.00 0.00 0.00 0.00	Polytypic	MSK-IMPACT#	Neg	LN	Y	NA	IgH: NT	NT							
9	TET2 p.M1656fs*36 TET2 p.G1154D DNMT3A p.C557* RHOA p.G17V IDH2 p.R172S IDH2 p.R172S CD28 p.T195P	0.44 0.30 0.46 0.46 0.39 0.00 0.41	TFHL-AI	MSK-IMPACT	LN	Y	NA	Pos	0.00 0.00 0.00 0.00 0.00 0.00 0.01	Polytypic	MSK-IMPACT	Pos	LN	Y	NA	IgH: NT	0.00 0.00 0.00 0.00 0.00 0.00 0.00	Granulocytes	MSK-IMPACT	PB	Y	NA	NA	
10	TET2 p.Q1539* RHOA p.G17V	0.51 0.51	TFHL-AI	MSK-IMPACT	LN	Y	NA	Pos^	0.00 0.00	Polytypic	MSK-IMPACT	Pos	LN	Y	NA	IgH: NT	NT							
11	TET2 p.Q1138Kfs*14 TET2 p.C1271Wfs*29 RHOA p.G17V ARID5B p.E456Kfs*23 CDK12 p.L873dup	0.48 0.46 0.59 0.32 0.41	TFHL-AI	MSK-IMPACT#	PB	Y	NA	NT	0.00 0.00 0.00 0.00 0.00	Polytypic	MSK-IMPACT#	Pos	PB	Y	NA	IgH: NT	0.00 0.00 0.00 0.00 0.00	Myeloid Cells	MSK-IMPACT#	PB	Y	NA	NA	
12	TET2 p.Q778* TET2 p.D1384N DNMT3A p.V296M RHOA p.G17V IDH2 p.R172S NCOR2 p.A989T SETD1B p.E612* XBP1 p.S52N BCR p.G6D ACTG1 p.S60T	0.42 0.28 0.41 0.07 0.06 0.07 0.05 0.00 0.00 0.25 0.25 0.00	TFHL-AI	MSK-IMPACT#	LN	N	15%	NT	0.43 0.41 0.39 0.00 0.00 0.00 0.00 0.00 0.25 0.25 0.22	LBCL-like MBP	MSK-IMPACT#	Pos	GE junction	N	≤5%	IgH/TCR: NT	0.48 0.45 0.48 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00	Bulk BM	MSK-IMPACT#	BM	N	≤5%	IgH/TCR: NT	
13	TET2 p.W1233* TET2 p.Q674* DNMT3A p.V872F RHOA p.G17V CTNNB1 p.G48C CTNNB1 p.S45F GRIN2A p.X336_splice EPHA7 p.A816V	0.31 0.33 0.29 0.24 0.32 0.30 0.24 0.00	TFHL-AI	MSK-IMPACT#	BM	Y	NA	Pos	0.28 0.22 0.27 0.00 0.01 0.00 0.00 0.20	LBCL-like MBP	MSK-IMPACT#	Pos	Skin	N	≤5%	IgH/TCR: NT	0.07 0.00 0.08 0.00 0.00 0.00 0.00 0.00	Myelomonocytic cells	MSK-IMPACT#	BM	Y	NA	NA	

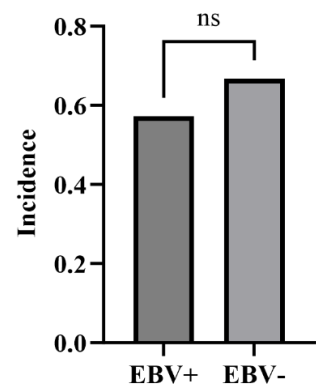
14	TET2 p.N427fs TET2 p.Q745X DNMT3A p.M801V IDH2 p.R172G TNFAIP3 p.R271* PAK7 p.P442Q MED12 p.R1343S TSC2 p.P542L BRAF p.K601N ATM p.P2842T BCORL1 p.V866F DTX1 p.A120T CD58 p.P125Hfs*5	0.17 0.16 0.21 0.04 0.18 0.12 0.08 0.08 0.03 0.00 NT NT NT	TFHL-AI MSK-IMPACT [#] LN N 15% Pos	0.10 0.09 0.12 0.00 0.00 0.00 0.00 0.10 0.10 0.07 0.05	LBCL-like MBP MSK-IMPACT [#] Pos Skin/Subq N ≤5% IgH/TCR: NT	0.05 0.06 0.14 0.00 NT NT NT NT NT NT NT NT	Bulk BM 28 gene panel BM N ≤1% IgH and TCR: Neg (PCR)
15	TET2 p.Y1661* TET2 p.I1873T TET2 p.T1554Sfs*16 PRDM1 p.Q171* KMT2D p.S2910Rfs*32 TET2 p.N1746Kfs*5 SF3B1 p.K666N TP53 p.H193R	0.44 0.22 0.11 0.00 0.00 0.00 0.00 0.00	TFHL-AI MSK-IMPACT LN N 40% Pos	0.42 0.03 0.06 0.30 0.25 0.23 0.00 0.00	LBCL-like MBP MSK-IMPACT [#] Neg Skin/Subq N ≤10% IgH/TCR: NT	0.38 0.00 0.00 0.00 0.00 0.00 0.05 0.03	Bulk PB MSK-IMPACT PB N ≤5% IgH/TCR: NT
16	TET2 p.R1452* TET2 p.L1276Wfs*87	0.85 0.00	TFHL-AI MSK-IMPACT PB Y NA Neg [^]	0.00 0.45	LBCL-like MBP MSK-IMPACT [#] Pos PB [†] Y NA IgH: NT	NT	
17	TET2 p.S631* TET2 p.G641W DNMT3A p.K455* TET2 p.V1232Gfs*21 PIK3C3 p.S460P TET3 p.C693G TET3 p.C695Y MAP2K1 p.K57N MAP2K1 p.Q45_E62del SPEN p.S147* FAT1 M1? CCND3 p.R271Pfs*53 HNF1A p.M154I CHEK2 p.C284* PTCH1 p.L39Cfs*41 TET2 p.Q1030*	0.45 0.49 0.47 0.00 0.00 0.52 0.00 0.47 0.08 0.02 0.07 0.08 0.27 0.21 0.00 0.00 0.00	TFHL-NOS MSK-IMPACT [#] PB Y NA Pos [^]	0.49 0.47 0.50 0.48 0.52 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00	Polymorphic MBP MSK-IMPACT [#] Pos PB [†] Y NA IgH: NT	0.11 0.13 0.18 0.09 0.09 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.03 0.03 0.01	Myeloid Cells MSK-IMPACT [#] PB Y NA NA
18	TET2 p.L1740* DNMT3A p.X518_splice RHOA p.G17V CTNNB1 p.S45F TBX3 p.A280T	0.38 0.40 0.06 0.06 0.06	TFHL-AI MSK-IMPACT [#] LN N 10% Pos	0.27 0.28 0.00 0.00 0.00	Polymorphic MBP MSK-IMPACT [#] Pos LN N ≤10% IgH: Pos (PCR, NGS) [^] , TCR: NT	NT	
19	TET2 p.S577Pfs*3 TET2 p.F1300V TET2 p.C1289F CUX1 p.E555L SETBP1 p.L1421P VAV1 p.Y174C CHEK2 p.D134G CBFB p.I102F KRAS p.A146T SETD1A p.G444R SETD5 p.A232V SOCS1 p.NQ5K* SOCS1 p.F79_Y80delinsLH ARID1B p.G357dup DNMT3A p.R736H SRSF2 p.P95H FBXO11 p.Y692H	0.37 0.07 0.24 0.30 0.30 0.33 0.02 0.00 0.00 0.01 0.00 0.00 0.00 0.01 0.00 0.01 0.00 0.01	TFHL-AI MSK-IMPACT [#] LN N 60% Pos	0.39 0.28 0.02 0.00 0.00 0.00 0.28 0.14 0.13 0.13 0.12 0.11 0.10 0.09 0.05 0.11 0.09	Polymorphic MBP MSK-IMPACT [#] Pos LN N ≤10% TCR: Neg (PCR), TCRg: Pos (NGS, 0.1%), IgH: NT	0.43 0.47 0.00 0.00 0.00 0.00 0.44 0.00 0.01 0.00 0.00 0.00 0.00 0.00 0.48 0.44	Bulk BM MSK-IMPACT [#] BM N ≤1% TCRg: Pos (NGS, 0.09%), IgH: NT
20	TET2 p.Q324Hfs*23 TET2 p.Q831* SOCS1 p.R179P TET2 p.L1276Sfs*22 DTX1 p.G58V FAT1 p.I1302M DAXX p.F79L RPTOR p.R616H	0.33 0.38 0.37 0.05 0.01 0.00 0.00 0.00	TFHL-NOS MSK-IMPACT [#] LN Y NA Pos	0.47 0.46 0.00 0.00 0.47 0.15 0.09 0.08	Polymorphic MBP MSK-IMPACT [#] Pos LN Y NA IgH: Pos (PCR)	0.01 0.00 0.00 0.00 NT 0.00 0.00 0.00	Bulk saliva and nails MSK-IMPACT [#] Saliva and nails N NA IgH/TCR: NT

21	TET2 p.A1373E TET2 p.V1136Cfs*6 DNMT3A p.P700L RHOA p.G17V CD28 p.T195P SOCS1 p.I194N PLCG1 p.D1169G TET2 p.C1271* TBX3 p.G218V	0.10 0.06 0.10 0.09 0.10 0.08 0.06 0.00 0.00	TFHL-AI MSK-IMPACT# LN N 20% NT						0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.38 0.06	Polymorphic MBP MSK-IMPACT# Pos PB ¹ Y NA IgH: NT					0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00	Granulocytes MSK-IMPACT# PB Y NA NA
22	TET2 p.H1382D (C>G) TET2 p.S1494* (C>A)	0.45 0.47	TFHL-NOS MSK-IMPACT# LN Y NA Pos [^]					0.00 0.00	Polymorphic MBP MSK-IMPACT# Pos PB ¹ Y NA IgH: Pos (PCR) [^]					0.00 0.00	Bulk BM 49 gene panel BM N ≤5% IgH: Pos (PCR), TCR: NT	
23	TET2 p.Y1628* TET3 p.T355fs*45 RHOA p.G17V KRAS p.A146V NRAS p.G12V ARID2 p.R13Efs*22	0.40 0.21 0.14 0.01 0.00 0.00	TFHL-AI MSK-IMPACT LN Y NA NT					0.56 0.32 0.00 0.26 0.10 0.00	FL-like MBP, grade 3B MSK-IMPACT Neg LN Y NA IgH: NT					0.97 0.47 0.00 0.06 0.09 0.06	Bulk BM MSK-IMPACT BM N ≤5% IgH and TCR: Neg (PCR)	
24	TET2 p.R1400* SRSF2 p.P95H TET2 p.R1465* TET2 p.Q734* ARID1A p.G105Efs*8 BCL2 p.E13D CRLF2 p.C180Y CSF1R p.V38L DTX1 p.G58D DTX1 p.V70L EPHA7 p.I11N KMT2D p.Q3961* KSR2 p.S184I LTB p.V36L MEF2B p.D83V PIM1 p.S75P PTCH1 p.P299L TNFAIP3 p.R136Qfs*3 TNFAIP3 p.L324Qfs*7 TNFRSF14 p.G60D TNFRSF14 p.N116Rfs*117	0.48 0.46 0.43 0.04 0.00 0.00 0.00 0.00 0.01 0.01 0.01 0.01 0.00 0.01 0.01 0.01 0.02 0.00 0.00 0.00 0.00	TFHL-AI MSK-IMPACT# LN Y NA Pos					0.00 0.00 0.00 0.00 0.28 0.43 0.51 0.39 0.49 0.52 0.52 0.35 0.46 0.46 0.99 0.47 0.45 0.34 0.30 0.69 0.18	FL-like MBP, grade 3A MSK-IMPACT# Pos LN Y NA IgH: Neg (PCR)					0.26 0.36 0.02 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.00	Bulk saliva MSK-IMPACT Saliva N NA IgH/TCR: NT	
25	TET2 p.Y1128* TET2 p.Q810* DNMT3A p.R736C RHOA p.G17V IDH2 p.R172K ATR p.D1409N KMT2B p.Q757* BRCA2 p.A2306S BTG1 p.K29* EPHA5 p.G1033E KMT2D p.H1525Pfs*37 SETD5 p.Y987F	0.46 0.46 0.48 0.46 0.41 0.21 0.25 0.00 0.00 0.00 0.00 0.00	TFHL-AI MSK-IMPACT# BM Y NA Pos					0.00 0.00 0.00 0.00 0.00 0.00 0.00 0.48 0.48 0.23 0.25 0.35	MM MSK-IMPACT# Neg BM Y NA IgH: Pos (PCR)					0.05 0.05 0.07 NT 0.00 NT NT NT NT NT NT NT	Bulk PB 49 gene panel PB N ≤1% IgH/TCR: NT	

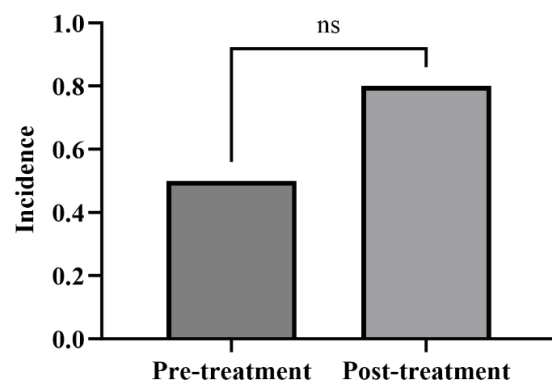
Footnote: Targeted next-generation sequencing (NGS)-based mutational analysis was performed using Memorial Sloan Kettering laboratory-developed hybrid-capture-based 400 or 410 gene panels (MSK-IMPACT) with or without a matched germline control (minimum depth of coverage 100x) or microdroplet amplicon-based 28 or 49 gene panels (minimum depth of coverage 500x), all containing at least 28 genes commonly mutated in hematopoietic neoplasms. Mutations with allele frequencies (VAFs) ≤0.02 are included here but were not included in the final analysis. Percentages of neoplastic T-, total B-, and/or myeloid cells in a sample (based on morphology, immunohistochemistry, flow cytometry, and/or gene rearrangement analysis) are reported only for unsorted samples. Variants with VAFs >2x the percentage of neoplastic T- and myeloid cells combined in bulk monotypic/monoclonal B cell proliferation (MBP) samples and of neoplastic T- and total B cells in bulk blood (PB) or bone marrow (BM) samples were considered present in MBP or myeloid compartments, respectively, as variants at those levels would generally be too high to be explained by the T/myeloid or T/B cells alone and would most likely be present in the remaining compartment (MBP or myeloid, respectively). Clonality assessment was performed during clinical case work up using polymerase chain reaction (PCR)-based (T cell receptor [TCR] gamma [TCRg] and beta, immunoglobulin heavy chain [IgH]) and/or NGS-based (TCRg, IgH) gene rearrangement assays. IgH gene rearrangement results are reported on all polytypic B cell and MBP samples (to provide clonality evaluation) and on bulk myeloid samples (to provide sample purity evaluation). TCR gene rearrangement results are reported on T-cell lymphoma (TCL) samples (PCR only) (to provide clonality evaluation) and on bulk MBP and myeloid samples (to provide purity evaluation). For TCR gene rearrangement studies using NGS, the percentage indicates the percentage of clonal reads of all rearranged TCR reads. *Droplet digital polymerase chain reaction (ddPCR) was performed utilizing custom primers targeting mutated genes detected by targeted NGS performed on corresponding bulk TCL samples (see Figure 2 for genes evaluated). For patients 2 and 3, the targeted NGS was performed on a separate TCL sample (data not shown). [^]Performed on a separate similar sample. [#]Matched germline control (nails or saliva) utilized. [†]The histologic type of MBP was determined on a concurrent or recent (obtained within 3 months) tissue sample that demonstrated the same B cell immunophenotype by flow cytometry and/or immunohistochemistry as the immunophenotypically abnormal B cell population identified in the genotyped PB sample by flow cytometry. [‡]Only a PB sample was available for evaluation, which showed an abnormal, light chain restricted B cell population with plasmacytic differentiation by flow cytometry and a lack of a significant population of large lymphocytes on the PB smear. EBV indicates Epstein-Barr virus; FL, follicular lymphoma; GE, gastroesophageal; LBCL, large B-cell lymphoma; LN, lymph node; MM, plasma cell myeloma; N, no; NA, not applicable; Neg, negative; NT, not tested; Pos, positive; TFHL-AI, follicular helper T-cell lymphoma, angioimmunoblastic type; TFHL-NOS, follicular helper T-cell lymphoma, not otherwise specified; Subq, subcutaneous tissue; Y, yes.

Supplemental Figure 1

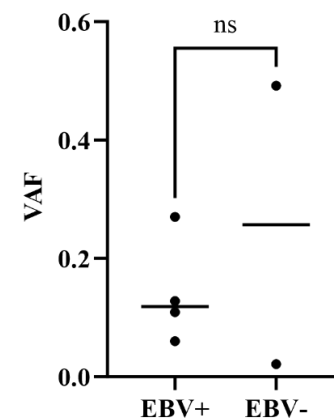
A Incidence of shared mutations in PolyBCs and TFHLs, EBV+ vs EBV-



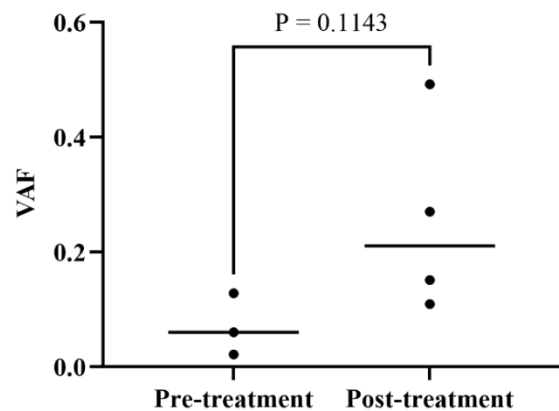
B Incidence of shared mutations in PolyBCs and TFHLs, pre- vs post-treatment



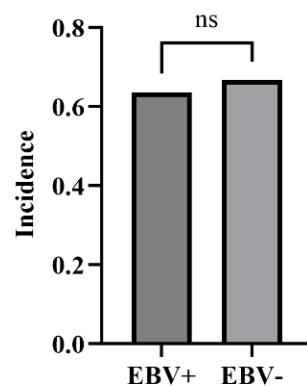
C VAFs of *TET2/DNMT3A* mutations in PolyBCs, EBV+ vs EBV-



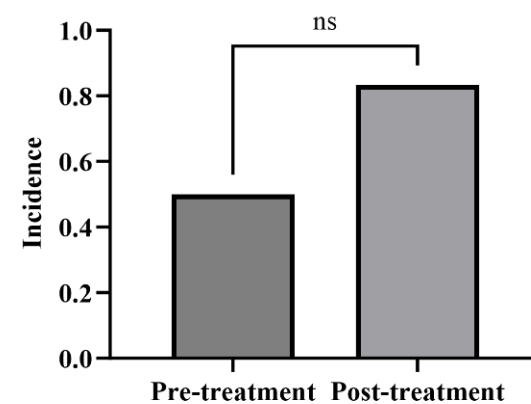
D VAFs of *TET2/DNMT3A* mutations in PolyBCs, pre- vs post-treatment



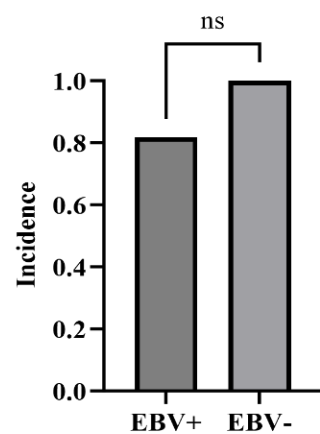
E Incidence of shared mutations in MBPs and TFHLs, EBV+ vs EBV-



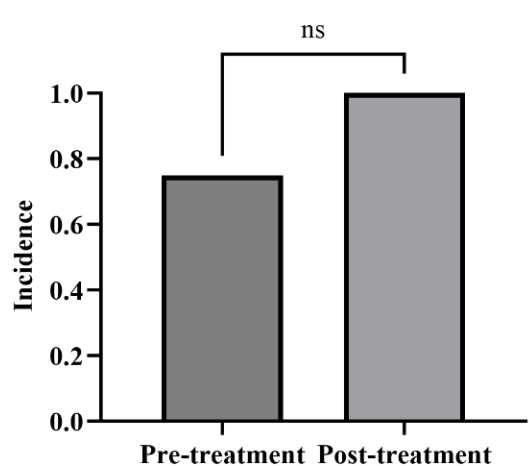
F Incidence of shared mutations in MBPs and TFHLs, pre- vs post-treatment



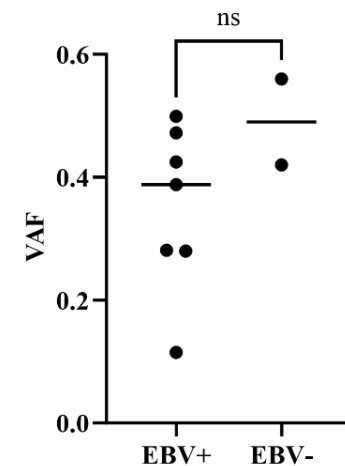
G Incidence of private mutations in MBPs EBV+ vs EBV-



H Incidence of private mutations in MBPs, pre- vs post-treatment

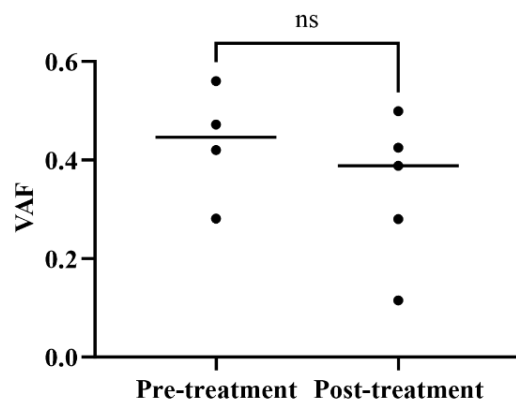


I VAFs of shared *TET2/DNMT3A* mutations in CH+ MBPs, EBV+ vs EBV-

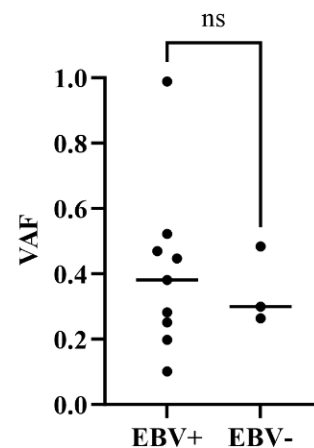


Supplemental Figure 1, Continued

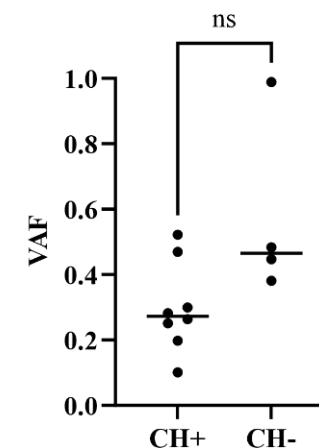
J VAFs of shared *TET2/DNMT3A* mutations in CH+ MBPs, pre- vs post-treatment



K VAFs of private mutations in MBPs, EBV+ vs EBV-

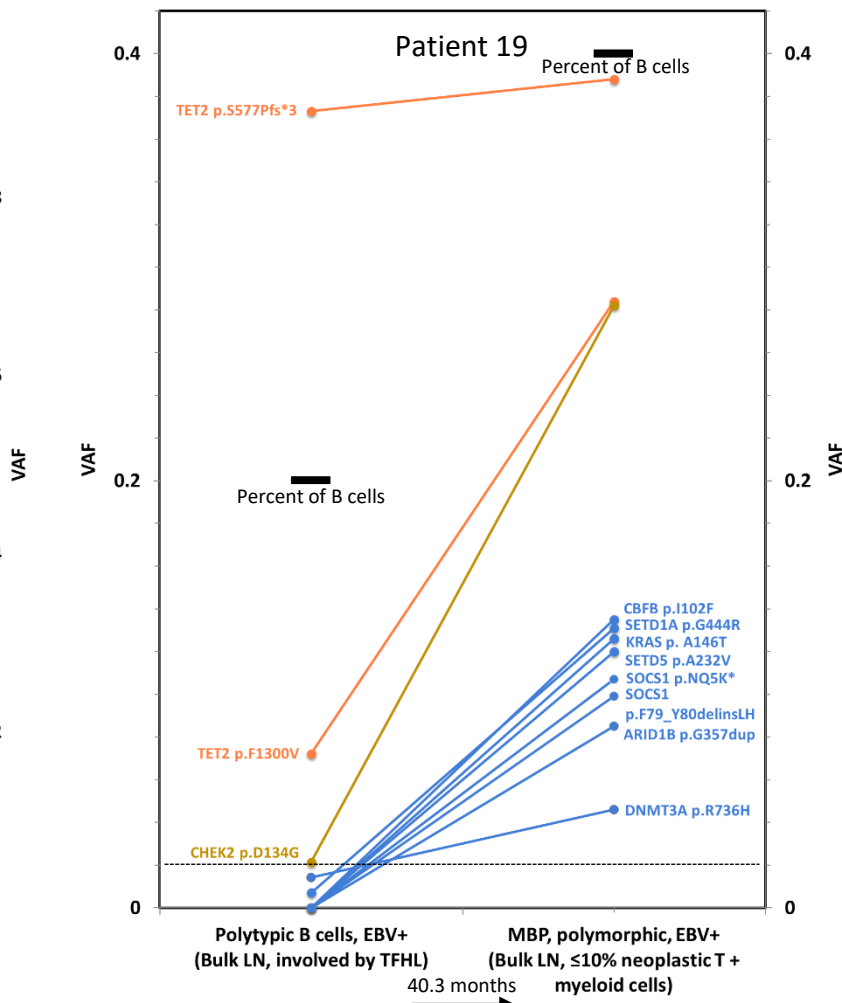
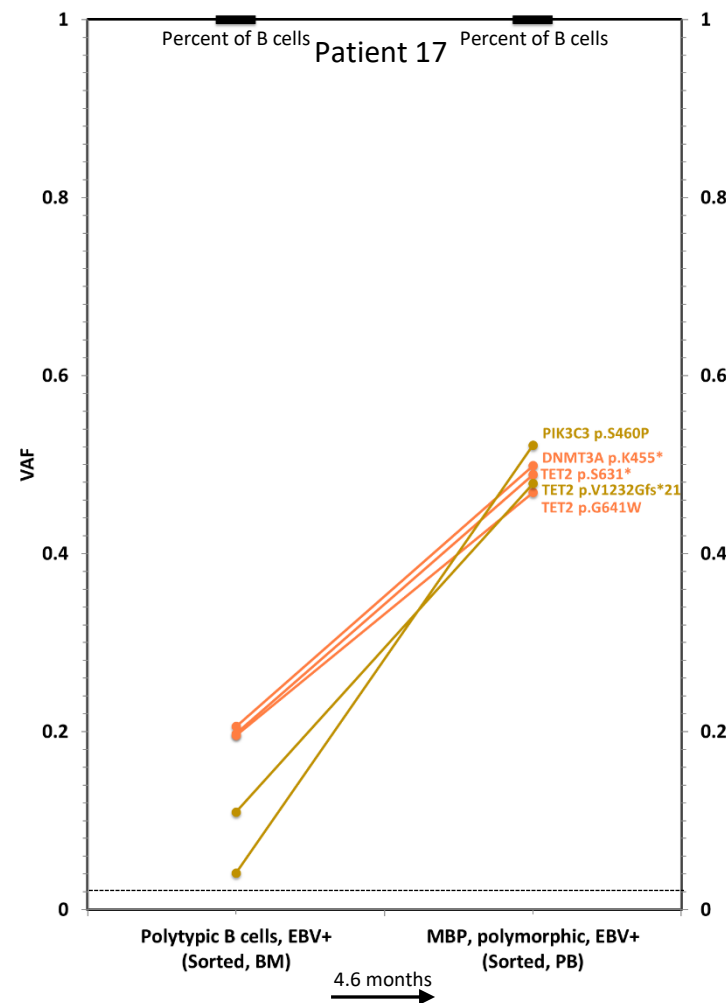
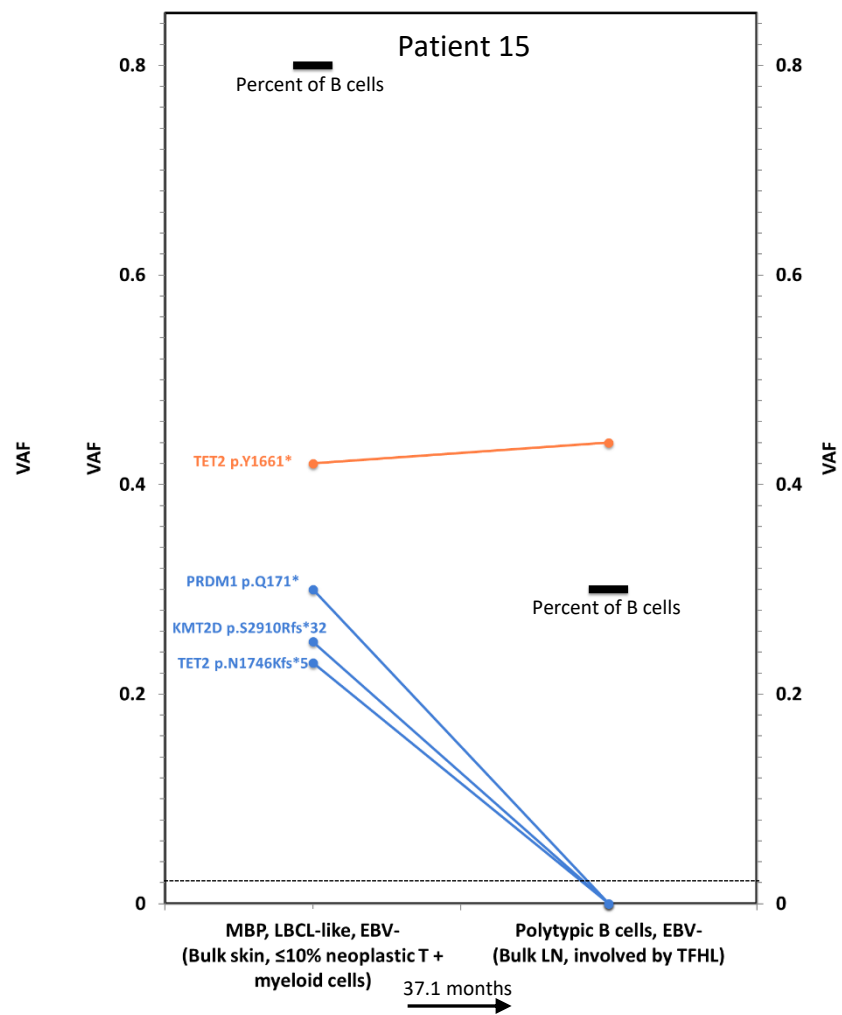
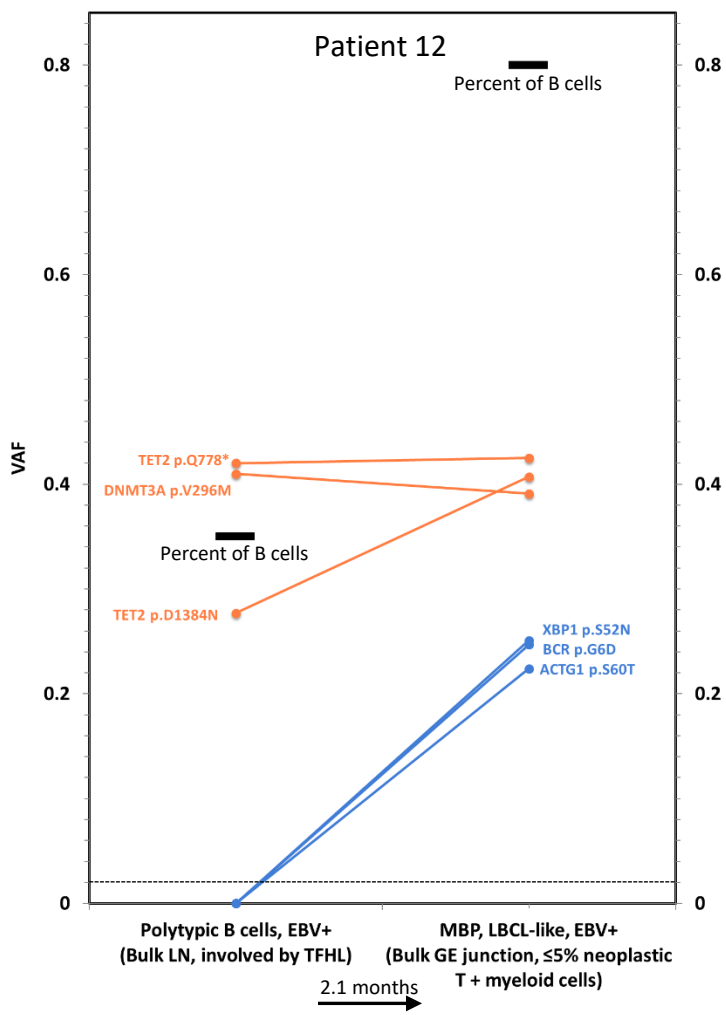


L VAFs of private mutations in MBPs, CH+ vs CH-



Supplemental Figure 1. Comparisons of the incidences and allele frequencies of shared and/or private mutations in polytypic B cells and monotypic/monoclonal B cell proliferations among various groups. (A) The proportion of patients with shared mutations in polytypic B cells (PolyBCs) and follicular helper T-cell lymphomas (TFHLs) was not significantly different among Epstein-Barr virus-positive (EBV+) and EBV-negative (EBV-) patients or (B) among those with PolyBCs sampled pre- and post-treatment. (C) The allele frequencies (VAFs) of the *TET2/DNMT3A* mutations in PolyBCs did not significantly differ among EBV+ and EBV- patients harboring those mutations. (D) A trend towards lower VAFs was present among samples obtained pre-treatment. (E) The incidence of shared mutations in monotypic/monoclonal B cell proliferations (MBPs) and TFHLs did not significantly differ among EBV+ and EBV- MBPs or (F) among MBP samples obtained pre- and post-treatment. (G, H) The incidence of private mutations in MBPs also did not differ among these groups. (I) Among clonal hematopoiesis-positive (CH+) MBPs (those that shared mutations with corresponding TFHLs), no significant difference in VAFs of the shared *TET2/DNMT3A* mutations was seen among EBV+ and EBV- MBPs or (J) among pre- and post-treatment samples. (K) Private mutant VAFs in MBPs did not significantly differ among EBV+ and EBV- cases or (L) among CH+ and CH-negative (CH-) MBPs. Horizontal lines in (C-D) and (I-L) indicate medians; ns, not significant.

Supplemental Figure 2



Supplemental Figure 2. Comparison of mutations detected in samples containing high levels of polytypic B cells and monotypic/monoclonal B cell proliferations obtained from the same patients. Separate samples containing high levels of polytypic B cells (PolyBCs) from 4 monotypic/monoclonal B cell proliferation (MBP) patients (12, 15, 17, 19) were sequenced. Such samples consisted of lymph node (LN) involved by follicular helper T-cell lymphoma (TFHL) (patients 12, 15, 19) or bone marrow (BM) (patient 17), which either pre-dated (patients 12, 17, 19) or followed (patient 15) MBP development at time intervals ranging from 2.1 to 40.3 months. Each patient received interval systemic therapy in-between the PolyBC- and MBP-containing samples. At PolyBC sampling, 3 patients were Epstein-Barr virus-positive (EBV+) (patients 12, 17, 19) and 1 EBV-negative (EBV-) (patient 15), the same EBV status as each patient's corresponding MBP. As the samples containing PolyBCs from patients 12, 15, and 19 were sequenced in bulk, determination of whether the shared mutations were present in the PolyBCs could not be performed. However, the private mutations seen in the corresponding MBPs were not identified despite high levels of PolyBCs in these samples. The *CHECK2* mutation in patient 19's PolyBC-containing sample was present at a low allele frequency (VAF) just below our cutoff (0.02) and whether it was present in background myeloid cells or in the PolyBCs could not be determined in this bulk sample. The PolyBCs from patient 17 were flow cytometry-sorted and harbored both the shared and private mutations detected in the MBP but at significantly lower VAFs. The PolyBC sample predated the MBP sample by a short time interval (4.6 months) in this case. The Y axes indicate the VAFs of the mutations detected as well as the fraction of B cells among total cells in each sample (horizontal black bars). The sample descriptions (including tissue site, EBV status, flow cytometry sorted vs bulk, and degree of involvement by other cell types) are indicated on the X axes with the time in-between the samples indicated on the bottom. Dotted lines indicate the VAF cut off of >0.02 used in this study for variant calling. Mutations in orange indicate those shared among the MBPs, TFHLs, and myeloid compartments, those in blue indicate private mutations found in MBPs and not in corresponding TFHLs or myeloid cells, while those in yellow indicate mutations detected in MBPs and myeloid cells but not in TFHLs. Private mutations detected in the TFHLs are not displayed. GE indicates gastroesophageal; LBCL, large B-cell lymphoma; PB, peripheral blood.