

Response to DA-EPOCH-R is associated with activation of ‘fitter’ cytotoxic T cells in patients with newly diagnosed double and triple hit high-grade B-cell lymphoma

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<https://doi.org/10.3324/haematol.2024.285170>

SUPPLEMENTS

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Running title: Immune effector cells during treatment of HGBL

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Supplementary files:

- Supplementary Table 1 & 2
- Legend Supplementary Figure 1
- Supplementary Figure 1

Supplementary Table S1

Baseline characteristics of newly diagnosed high grade B-cell lymphoma patients stratified for achieving complete metabolic remission (CMR) after treatment with DA-EPOCH-R (left columns) and all patients (right column). Statistical tests used for group comparisons include the Pearson χ^2 test and the Kruskal–Wallis test for categorical and continuous variables, respectively. P-values < 0.05 are considered statistically significant.

	CMR (N=40)	No CMR (N=30)	P-value	All patients (N=70)
Age (years)				
Mean (SD)	59.3 (10.5)	61.5 (9.16)	0.35	60.2 (9.93)
Median [Min, Max]	60.0 [35.0, 79.0]	62.5 [36.0, 78.0]		62.0 [35.0, 79.0]
Sex				
Male	27 (67.5%)	19 (63.3%)	0.91	46 (65.7%)
Female	13 (32.5%)	11 (36.7%)		24 (34.3%)
WHO Performance score				
0	29 (72.5%)	15 (50.0%)	0.03	44 (62.9%)
1	11 (27.5%)	11 (36.7%)		22 (31.4%)
2	0 (0%)	4 (13.3%)		4 (5.7%)
3	0 (0%)	0 (0%)		0 (0%)
LDH				
Within reference range	17 (42.5%)	6 (20.0%)	0.002	23 (32.9%)
Elevated	23 (57.5%)	24 (80.0%)		47 (67.1%)
Extranodal localisations (no.)				
None	17 (42.5%)	12 (40.0%)	0.62	29 (41.4%)
1	9 (22.5%)	5 (16.7%)		14 (20.0%)
2 or more	14 (35.0%)	12 (40.0%)		26 (37.1%)
Unknown	0 (0%)	1 (3.3%)		1 (1.4%)
IPI-score				
Low	7 (17.5%)	3 (10.0%)	0.02	10 (14.3%)
Low-intermediate	21 (52.5%)	8 (26.7%)		29 (41.4%)
High-intermediate	8 (20.0%)	17 (56.7%)		25 (35.7%)
High	4 (10.0%)	2 (6.7%)		6 (8.6%)
DH/TH status				
BCL2 DH	31 (77.5%)	18 (60.0%)	0.18	49 (70.0%)
BCL6 DH	4 (10.0%)	3 (10.0%)		7 (10.0%)
TH	5 (12.5%)	9 (30.0%)		14 (20.0%)
COO classification (Hans)				
GCB	31 (77.5%)	28 (93.5%)		59 (84.3%)
Non-GCB	4 (10%)	2 (6.7%)		4 (5.7%)
Unknown (poor morphology)	5 (12.5%)	0		7 (10%)

Supplementary Table S2.

Flow-cytometric phenotyping of cryopreserved PBMCs and the number of patient samples measured for each flow-cytometry panel per time point.

Human normal immunoglobulins (final concentration 0.1 mg/ml, Nanogram, Sanquin Plasma Products B.V.) were added before staining to reduce unwanted non-specific FC-receptor binding of fluorochrome-conjugated antibodies. Spectral overlap was automatically calculated and compensated using compensation beads and FACSDiva™ software. Flow cytometer performance and standardization were monitored daily with fluorescent-labeled CS&T beads (BD, 655051). Laser voltages were optimized within a <2% deviation using BD OneFlow™ Setup Beads (BD, 658620) and Alignflow™ Flow cytometry Alignment Beads (Thermo Fisher Scientific, A16502) on a daily setting.

Raw flow cytometry data (fcs 3.1 files) were manually analyzed using FCS Express Flow Cytometry Software (version 6) to identify, gate and export single T-cell and NK-cell populations. Pre-processing included data cleaning (PeacoQC18), hyperbolic arcsin transformation, approximated min-max scaling and batch correction (quantile normalization) in R and R studio (version 4.0.3). Computational data analysis was performed using UMAP and FlowSOM on 10.000 cells per fcs file.

DA-EPOCH-R Flow cytometry panels	Day 1 first cycle START	Day 1 third cycle MID	After fifth cycle END
I. T-cell phenotype Differentiation, senescence, exhaustion (PD-1, TIM3, TIGIT, CD27, CD28)	CMR N=40	CMR N=35	CMR N=37
	No CMR N=30	No CMR N=23	No CMR N=12
II. T-cell phenotype Tregs, gd-T-cells Exhaustion (PD-1, LAG3) Activation (CD38, HLA-DR)	CMR N=40	CMR N=30	CMR N=34
	No CMR N=26	No CMR N=18	No CMR N=12
III. NK-cell phenotype NK-cell subsets, senescence, activation (DNAM-1, HLA-DR)	CMR N=35	CMR N=23	CMR N=31
	No CMR N=23	No CMR N=15	No CMR N=11
IV. NK-cell phenotype NK-cell subsets, receptors (NKG2A, NKG2C, NKG2D and TIM-3)	CMR N=31	CMR N=18	CMR N=26
	No CMR N=18	No CMR N=13	No CMR N=9

Legend

Supplementary Figure 1. NK-cell and T-cell phenotypes associated with response to DA-EPOCH-R.

(A) Total white blood cell (WBC) count as $\times 10^9/L$ collected at day 1 of the first (start) and third (mid) and after the fifth DA-EPOCH-R cycle (end) for patients who at the end of DA-EPOCH-R achieved complete metabolic remission (CMR, blue) or did not achieve CMR (red). (B) Dimensionality reduction by UMAP of the flow cytometry data of NK-cells. Color overlays for CD16, CD56, CD57, KLRG1 and TIGIT. (C) Percentages of kill of K562 and degranulation as measured by CD107a/b surface expression on NK-cells after 4 hours co-culture of peripheral blood mononuclear cells (PBMCs) with K562 cell line at day 1 of the first (start) and after the fifth DA-EPOCH-R cycle (end) for patients who at the end of DA-EPOCH-R achieved complete metabolic remission (CMR, blue) or did not achieve CMR (red). Cytotoxicity is calculated relative to the amount of K562 cells without PBMCs.

(D) Percentages of CD4⁺ T-cells, CD8⁺ T-cells, regulatory T-cells (Tregs), CD4⁺CD8⁻ and innate-like $\gamma\delta$ -T-cells as percentages of total T-cells at day 1 of the first (start) and third (mid) and after the fifth DA-EPOCH-R cycle (end) for patients who at the end of DA-EPOCH-R achieved complete metabolic remission (CMR, blue) or did not achieve CMR (red).

(E) Dimensionality reduction by UMAP of the flow cytometry data of T-cells. Color overlays for CD8, CD38, HLA-DR and PD-1.

(F-G) LAG3⁺ (left panels) or TIGIT⁺ (right panels) (F) and CD4⁺ (left panels) and CD8⁺ (right panels) expressing markers associated with T-cell senescence (loss of CD27 and CD28, and gain of KLRG1 and CD57) (G) as percentages of total T-cells at day 1 of the first (start) and third (mid) and after the fifth DA-EPOCH-R cycle (end) for patients who at the end of DA-EPOCH-R achieved complete metabolic remission (CMR, blue) or did not achieve CMR (red). (H) Percentages of CD4⁺ (upper panels) CD8⁺ (lower panels) T-cells expressing markers associated with T-cell activation (CD38, HLA-DR and CD25), immune checkpoints (LAG3 and TIGIT), and markers associated with T-cell senescence (loss of CD27 and CD28, and gain of KLRG1 and CD57) at third (mid) and after the fifth DA-EPOCH-R cycle (end) for patients who at the end of DA-EPOCH-R achieved complete metabolic remission (CMR, blue) or did not achieve CMR (red).

For all box plots, the lower upper hinges correspond to the 25th and 75th percentiles. The middle hinge corresponds to the median. The whiskers extend from the largest to smallest value $\pm 1.58 \times$ interquartile range. Outliers are plotted individually.

Linear mixed effect models (A,D, F, G) and non-parametric Mann-Whitney U test between two groups (C, H) were used for statistical analysis in which $P < 0.05$ was considered significant. No multiple correction was applied.

Supplementary figure 1

