Inherited platelet diseases with normal platelet count: phenotypes, genotypes and diagnostic strategy

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INTRODUCTION

Inherited platelet disorders resulting from platelet function defects and a normal platelet count cause a moderate or severe bleeding diathesis. Since the description of Glanzmann thrombasthenia resulting from defects of ITGA2B and ITGB3, new inherited platelet disorders have been discovered, facilitated by the use of high throughput sequencing and genomic analyses. Defects of RASGRP2 and FERMT3 responsible for severe bleeding syndromes and integrin activation have illustrated the critical role of signaling molecules. Important are mutations of P2RY12 encoding the major ADP receptor causal for an inherited platelet disorder with inheritance characteristics that depend on the variant identified. Interestingly, variants of GPε encoding the major subunit of the collagen receptor GPVI/FcRγ associate only with mild bleeding. The numbers of genes involved in dense granule defects including Hermansky-Pudlak and Chediak Higashi syndromes continue to progress and are updated. The ANO6 gene encoding a Ca2+-activated ion channel required for phospholipid scrambling is responsible for the rare Scott syndrome and decreased procoagulant activity. A novel EPHB2 defect in a familial bleeding syndrome demonstrates a role for this tyrosine kinase receptor independent of the classical model of its interaction with ephrins. Such advances highlight the large diversity of variants affecting platelet function but not their production, despite the difficulties in establishing a clear phenotype when few families are affected. They have provided insights into essential pathways of platelet function and have been at the origin of new and improved therapies for ischemic disease. Nevertheless, many patients remain without a diagnosis and requiring new strategies that are now discussed.

Glanzmann thrombasthenia

With a worldwide distribution, Glanzmann thrombasthenia (GT) is the most common platelet function disorder.12 Most importantly, studies on it led to the characterization of the αIIbβ3 integrin mediator of platelet aggregation (Figure 1);
**Table 1. Principal named inherited platelet function disorders with a normal platelet count**

<table>
<thead>
<tr>
<th>Disease name</th>
<th>Gene (transmission - OMIM)</th>
<th>Platelets count - volume - granules</th>
<th>Isolated or syndromic bleeding severity</th>
<th>Principal platelet function defects</th>
<th>Biology</th>
<th>Ref</th>
</tr>
</thead>
<tbody>
<tr>
<td>Glanzmann thrombasthenia 273800</td>
<td>ITGA2B/ITGB3 (biallelic) c11bb3 integrin</td>
<td>N/N/N</td>
<td>Isolated Major bleeding</td>
<td>Absence of platelet aggregation with all agonists. Positive response to ristocetin Clot retraction often defective</td>
<td>Defects of the c11bb3 integrin: Type 1: &lt;5% Type 2: 5% -20% Variant: &gt;20% but non functional No binding of Fg</td>
<td>12-22</td>
</tr>
<tr>
<td>RASGRP2- (CalDAG-GEFI)-RD 613888</td>
<td>RASGRP2 (biallelic) CalDAG-GEFI</td>
<td>N/N/N</td>
<td>Isolated Major bleeding syndrome</td>
<td>Reduced or absent aggregation Positive response to ristocetin Normal response with PMA</td>
<td>Normal presence but defective activation of c11bb3.</td>
<td>25-28</td>
</tr>
<tr>
<td>Leukocyte adhesion deficiency-III (LAD-III syndrome) 612940</td>
<td>FERM3 (biallelic) Kindlin-3</td>
<td>N/N/N</td>
<td>Leukocytosis</td>
<td>Syndromic with severe infections Osteopetrosis (inconstant) Major bleeding</td>
<td>Absence of platelet aggregation with all agonists Positive response to ristocetin. Defective white cell functions</td>
<td>29-34</td>
</tr>
<tr>
<td>Scott syndrome 262690</td>
<td>ANO6 (TREM16F) (biallelic) Anoctamin-6</td>
<td>N/N/N</td>
<td>Isolated Bleeding after surgery</td>
<td>Defective platelet-derived thrombin formation Defect of microvesiculation</td>
<td>Lack of Ca2+-dependent translocation of PS to the surface of platelets and red cells</td>
<td>35-42</td>
</tr>
<tr>
<td>EPHB2-RD 614862</td>
<td>EPUB2 (biallelic) EPUB2</td>
<td>N/some elongated platelets/N</td>
<td>Isolated Spontaneous bleeding and after lesions</td>
<td>Defective platelet response to collagen, ADP, AA and TXA2 analogs</td>
<td>Tyrosine phosphorylation of intermediates involved in GPVI and G-protein-coupled receptor mediating signaling</td>
<td>44-53</td>
</tr>
<tr>
<td>P2Y12 receptor-RD 699821 AD</td>
<td>P2RY12 (biallelic) P2Y12</td>
<td>N/N/N</td>
<td>Isolated Bleeding can be severe. For AD disease bleeding frequently mild</td>
<td>ADP reversible aggregation even at high doses. Response reduced with collagen, AA, reversible, with TRAP. Defective VASP de-phosphorylation after ADP stimulation</td>
<td>Defect of Gq-coupled P2Y12 receptor linked to adenylyl cyclase and PI3-kinase activation</td>
<td>44-53</td>
</tr>
<tr>
<td>GPVI-RD 614201</td>
<td>GP6 (biallelic) GPVI</td>
<td>N/N/N</td>
<td>Isolated Bleeding absent or mild.</td>
<td>Absence of response with collagen, Cx or CRP. Decreased binding of fibrin and other ligands</td>
<td>Lack or nonfunctioning of GPVI in complex with FcRγ. Impaired signaling through a pathway involving PLCγ2</td>
<td>54-59</td>
</tr>
<tr>
<td>TXA2 receptor-RD 614009</td>
<td>TXB2 (biallelic) TXB2</td>
<td>N/N/N</td>
<td>Isolated Post-surgery bleeding</td>
<td>Absence of response to AA and TXA analogs.</td>
<td>Defective TPcx-coupled signaling via Gq-protein; signaling pathway involves PLCβ</td>
<td>60-64</td>
</tr>
<tr>
<td>Hermansky-Pudlak syndrome (HPS): 10 subtypes OMIM: HPS1:203300 HPS2:608233 HPS3: 614072 HPS4: 614073 HPS5: 614074 HPS6: 614075 HPS7: 614076 HPS8: 614077 HPS9: 614171 HPS10: 617050</td>
<td>10 Genes according subtypes HPS1,3,4,5,6 encoded for HPS3,4,5,6 HPS2 for AP3B1 HPS7 for Dysbindin HPS8 for BLOS3 HPS9 for Pallidin HPS10 for AP3D1</td>
<td>N/N/defect of dense granules</td>
<td>Syndromic OCA HPS1-10 -Pulmonary fibrosis and granulomatous colitis, HPS1,4 -Immunodeficiency and neutropenia, HPS2,10 -Mild to severe bleeding syndrome HPS1-10</td>
<td>Defect of aggregation to ADP, collagen, AA Defect of secretion whatever the agonist used Decreased or absence of dense granules Defect of CD63</td>
<td>Platelet storage pool disease affecting dense granules member of the lysosomal-related organelle family. Genes manifest via altered function of BLOC1-3 and AP-3 complexes during organelle biogenesis</td>
<td>67-73</td>
</tr>
<tr>
<td>Chediak-Higashi syndrome (CHS) 214500</td>
<td>LYST (biallelic) LYST</td>
<td>N/N/defect of dense granules</td>
<td>Mild bleeding Multi-syndromic OCA, recurrent infections, immune dysregulation with lymphoproliferative histiocytosis</td>
<td>Defect of aggregation to ADP, collagen, AA Defect of secretion whatever the agonist Decrease or absence of dense granules</td>
<td>Lysosomal storage disorder affecting platelet dense (β) granules, member of the lysosomal-related organelle family. LYST is a member of the BEACH protein family</td>
<td>67,69,74</td>
</tr>
<tr>
<td>Cytosolic phospholipase A2- syndrome 606522</td>
<td>PLA2G4A (biallelic) cPLA2</td>
<td>N/N/N</td>
<td>Isolated Moderate bleeding Recurrent Gluceration</td>
<td>Defect of platelet response to AA, collagen, and absence of second wave with ADP</td>
<td>Signaling pathway defect Impaired liberation of AA from membrane phospholipids</td>
<td>84-86</td>
</tr>
</tbody>
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this was accompanied by the emergence of a new category of powerful anti-ischemic drugs.6,14

Definition
Classically, GT is a severe bleeding diathesis with autosomal recessive transmission caused by an absence of platelet aggregation induced by all physiologic agonists.

Clinical phenotype
The clinical phenotype is marked by life-long spontaneous easy bruising, epistaxis and gum bleeding, often requiring platelet transfusions. Gastro-intestinal hemorrhage, menorrhagia and severe bleeding at menarche feature prominently, as does trauma-related hemorrhage. The first signs usually appear at birth, are intense during childhood but tend to decrease in later life. GT is prevalent in ethnic communities.15

Biological phenotype
 Patients have a normal platelet count and morphology. Most have platelets lacking αIIbβ3 although some express residual amounts but insufficient to support platelet aggregation: these have been termed type I and type II thrombasthenia.12 A third category consists of phenotypic variants in which platelets have reduced to normal amounts of αIIbβ3 unable to function. While type I GT patients possess platelets that fail to promote clot retraction, this can be subnormal or normal for some with residual or variant αIIbβ3. Platelets respond to ristocetin and adhere to other adhesive proteins can participate. Occupied fibrinogen is the principal ligand for aggregation although other adhesive proteins can participate. Occurred αIIbβ3 undergoes further conformational changes, including clustering, providing “outside-in” signaling pathways essential for thrombus consolidation, platelet spreading and clot retraction.15 Variants within the cytoplasmic domains of αIIb or β3 interfere with the binding of talin and kindlin-3 during early αIIbβ3 activation.17 Rare gain-of-function monoallelic variants in cytoplasmic and membrane proximal domains promote spontaneous conformational changes that result in macrothrombocytopenia, as detailed in our companion paper.11 Significantly, recent data obtained by next-generation sequencing have begun to reveal combinations of variants in different genes that modify the GT phenotype.18

Target for anti-platelet therapy
Because of the importance of αIIbβ3 in the formation of arterial thrombi, a generation of antiplatelet drugs block-
The integrin αIIbβ3 function was developed some 20 years ago. The main drugs are abciximab, a humanized Fab fragment of the murine monoclonal antibody 7E3 (c7E3), integrilin, a KGD-based cyclic heptapeptide and tiotibaf a synthetic non-peptide inhibitor; all are given intravenously. These inhibitors have basically been used in acute coronary syndromes. While the associated bleeding risk has now restricted their use, they are still recommended for bailout in shunt thrombosis or as a bridge to supplement platelet inhibition by orally taken drugs such as anti-P2Y12 inhibitors.

**RASGRP2-(CalDAG-GEFI)-related disease**

In 2014 a missense mutation (p.G248W) in RASGRP2 coding for calcium and diacylglycerol-regulated guanine nucleotide exchange factor I (CalDAG-GEFI) was reported for siblings with a severe bleeding disorder and much reduced platelet aggregation despite normal αIIbβ3 levels. Since this report, a characteristic GT-like defect of platelet function has been confirmed worldwide for 23 cases, as recently reviewed by Canault and Alessi.

**Definition**

Gene variants of RASGRP2 represent a new autosomal recessive inherited bleeding disorder linked to major defects of platelet signaling resulting in impaired αIIbβ3 activation. CalDAG-GEFI activates Rap1, which initiates “inside-out” signaling for αIIbβ3 promoting the binding of talin to shift the integrin into its high-affinity state. Both mouse and human platelets lacking functional CalDAG-GEFI undergo a slow but sustained activation that supports the formation of small thrombi at low shear such as those found in the venous system but not the large thrombi characteristic of arterial blood flow.

**Clinical phenotype**

Bleeding manifestations start early in life and can be severe. Epistaxis is common and most patients have other cutaneous bleeds. Gastrointestinal bleeding has been reported in four patients and menorrhagia in seven out of nine women. The bleeding diathesis is not truly syndromic; CalDAG-GEFI is found in other blood cell lineages, vascular cells and the brain striatum and has been implicated in Huntington disease. It often occurs in ethnic groups. One case, a male of Jamaican origin, successfully underwent surgery for a brain tumor under recombinant activated factor VII administration despite inefficacy of platelet transfusions.

**Biological phenotype and genetics**

A critical characteristic is that residual platelet aggregation and elevation of Ca²⁺ level, which is key for CalDAG-GEFI activity, is retained for high doses of some physiological agonists. Furthermore, αIIbβ3 fails to activate normally with decreased binding of the PAC1 IgM monoclonal antibody. Phorbol 12-myristate 13-acetate (PMA) activates Rap1 independently of surface receptors and CalDAG-GEFI and PMA-induced platelet aggregation is a diagnostic test for this disease. Interestingly, platelet spreading on a collagen surface is much decreased and while platelets attach to fibrinogen, lamellipodia are not formed. In the profile of RASGRP2 variants updated by Canault & Alessi in 2020, 12 of the 23 families had a premature stop codon or frameshift mutations while the others possessed missense mutations in functional domains important for CalDAG-GEFI activity.

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**Figure 1. Integrin αIIbβ3 and Glanzmann thrombasthenia.** The model illustrates the bent resting mature integrin and shows the many interactions between αIIb (green) and β3 (violet). Much information has been obtained from variant forms (noted in blue) that block functional sites in the extracellular and cytoplasmic domains while allowing αIIbβ3 expression. Other rare mutations (in red) give rise to activated integrin. Full information on mutations giving rise to Glanzmann thrombasthenia is found in Nurden & Pillois. Fg: fibrinogen; GT: Glanzmann thrombasthenia.
Leukocyte adhesion deficiency III syndrome

Around 2009, several groups reported that genetic variants of \textit{FERMT3} encoding kindlin-3, which is expressed in all hematopoietic cells, caused leukocyte adhesion deficiency III (LAD-III) syndrome.\textsuperscript{29} Kindlin-3 together with its paralogs kindlin-1 and kindlin-2 are key regulators of cellular functions and cell-matrix interactions but kindlin-3 predominates in platelets.

Definition

LAD-III syndrome, with an autosomal recessive inheritance, combines a rare immunodeficiency and bleeding syndrome of extreme severity characterized by leukocytosis, platelet dysfunction and recurrent infections. The genetic defects result in loss of activation of $\beta_1$, $\beta_2$ and $\beta_3$ integrins.

Clinical phenotype

Patients are detected when children have GT-like bleeding with severe non-purulent infections and often osteopetrosis. The clinical phenotype is based on the description of a limited number of largely homozygous patients mostly from ethnic Turkish and Arab populations.\textsuperscript{29}

Biological phenotype and genetics

As in GT, platelets of patients with LAD-III syndrome fail to aggregate in response to all physiological agonists and PMA.\textsuperscript{30} This translates into markedly reduced thrombus formation when whole blood is perfused over collagen, von Willebrand factor or fibrinogen-containing microspots.\textsuperscript{30} $\alpha$IIb$\beta$3 integrin is present but fails to bind fibrinogen or the PAC-1 monoclonal antibody when platelets are activated (Figure 2). Most disease-causing variants in \textit{FERMT3} abrogate kindlin-3 expression.\textsuperscript{29} As first shown in mice, kindlin-3 is an essential cytoplasmic cofactor for the activation of $\beta_1$, $\beta_2$ and $\beta_3$ integrins.\textsuperscript{32} Kindlin-3 binds in a phosphorylation-dependent manner to a subterminal NITY motif of the $\beta_3$ cytoplasmic tail; it coordinates with talin, which binds independently to trigger “inside-out” integrin activation.\textsuperscript{27,32} Initially there was controversy about the role of a CaLDAG-GEFI mutation in this disorder but this was dispelled when a splice site mutation in \textit{RASGRP2} observed in seven subjects with LAD-III syndrome was shown to be in linkage disequilibrium with \textit{FERMT3}; the two genes are located very closely on chromosome 11q.\textsuperscript{33,34} Most mutations are nonsense or involve frameshifts.

Figure 2. Loss of CalDAG-GEFI and kindlin-3 function abrogates $\alpha$IIb$\beta$3 activation. (A) A schematic representation showing examples of agonistic receptors and $\alpha$IIb$\beta$3 in the resting and activated forms. Intracytoplasmic signaling pathways induced by the binding of appropriate ligands lead to parallel roles of CalDAG-GEFI and kindlin-3 in promoting $\alpha$IIb$\beta$3 activation. CalDAG-GEFI is critical for RAP1 activation both in the circulation and at sites of vascular injury. It responds to changes in cytoplasmic Ca$^{2+}$ providing sensitivity and speed to the activation response; its absence leads to a Glanzmann thrombasthenia-like phenotype. Kindlin-3 binds directly to $\beta$-integrin cytoplasmic tails in platelets but also in white blood cells, explaining the susceptibility to infections and immune disorders in its absence. (B) Key elements of the syndromes resulting from \textit{RASGRP2} and \textit{FERMT3} defects are summarized. Bleeding in both cases can be very severe but defects in \textit{FERMT3} are syndromic and life-threatening. TRAP: thrombin receptor agonist peptide; PAR: protease activated receptor; Cvc: convulxin; GPVI: glycoprotein VI; TXA2: thromboxane A2; PLC$\gamma$: phospholipase C$\gamma$; DAG: diacylglycerol; CalDAG-GEFI: calcium and diacylglycerol-regulated guanine nucleotide exchange factor I; RAP1: RAS-related protein 1; RD: related disease; AR: autosomal recessive; LTA: light transmission aggregometry; ADP: adenosine triphosphate; Col: collagen; Cvc: convulxin; AA: arachidonic acid; PMA: phorbol 12-myristate 13-acetate.
Defect of platelet procoagulant activity: Scott syndrome

The original patient reported many years ago in New York had impaired platelet procoagulant activity and fibrin formation but other platelet functions were normal.\textsuperscript{35} The affected gene ANO6 (also called TMEM16F) was discovered in 2010 when a constitutively active mutant was identified in a cDNA library obtained from a Ba/F3 cell line overexpressing phosphatidyserine on the cell surface; a homozygous deleterious variant of ANO6 in a patient with Scott syndrome validated the findings.\textsuperscript{36}

\textbf{Definition}

Scott syndrome is characterized by autosomal recessive transmission, a lack of phosphatidyserine exposure and phosphatidyserine-bearing microparticle production from platelets and red blood cells.\textsuperscript{37,38} Loss of Ca\textsuperscript{2+}-dependent binding of the intrinsic tenase (FVIIIa, IXa, X) and prothrombinase (FVa, Xa, prothrombin) complexes results in reduced thrombin generation.

\textbf{Clinical phenotype}

Only six cases of Scott syndrome have been detailed and all are women.\textsuperscript{39} Bleeding is predominant after surgery or trauma; menorrhagia and post-partum hemorrhage are marked. Although spontaneous mucocutaneous bleeding is rare, it was severe in one young woman and included epistaxis, gum and gastrointestinal bleeding.\textsuperscript{39}

\textbf{Biological phenotype}

The major diagnostic clues are elevated residual levels of prothrombin in sera and a marked defect of platelet procoagulant activity. Full diagnosis requires flow cytometry measurements of platelet phosphatidyserine exposure (annexin V-binding) and microparticle release upon activation by thrombin in combination with collagen or by the ionophore A23187.\textsuperscript{39} Loss of mitochondrial membrane potential, a major factor in phosphatidyserine exposure, late phase procoagulant ballooning and capping of adhesive proteins all still occur but may be reduced. Caspase-dependent apoptotic or necrotic pathways of phosphatidyserine expression are normal.\textsuperscript{39} Platelet numbers, morphology, adhesion, secretion and aggregation are largely unaffected.

\textbf{Genotype}

Gene variants of ANO6 encoding a multi-pass transmembrane protein that is an essential component of the Ca\textsuperscript{2+}-dependent “scramblase” activity cause Scott syndrome.\textsuperscript{39} For the original patient a homozygous variant at a splice acceptor site in intron 12 resulted in a premature stop codon.\textsuperscript{39} Two French siblings showed compound heterozygosity for a large deletion of exons 1-10 in one allele and a premature stop codon on the other.\textsuperscript{40} Compound heterozygosity with premature termination was also seen in a Welsh family.\textsuperscript{40} All mutations result in abrogated TMEM16F expression.

\textbf{Ephrin type-B receptor 2-related disorder}

Recently, we demonstrated that defects in Ephrin type-B receptor 2 (EPHB2) cause a bleeding syndrome and IPD in two siblings from a consanguineous French family.\textsuperscript{41}

\textbf{Definition}

Recruent bleeding, normal platelet counts but abnormal function were linked to a variant of EPHB2, encoding a member of the EPH receptor family of transmembrane tyrosine kinases. The molecular basis of this IPD confirms a specific role for EPHB2 in platelet function via tyrosine phosphorylation of intermediates involved in GPVI and G-protein–coupled receptor-mediated signaling.

\textbf{Clinical phenotype}

Both affected family members showed excessive spontaneous subcutaneous and heavy bleeding upon minor wounds.\textsuperscript{40} One sibling developed anemia after chronic gastrointestinal bleeding requiring iron supplementation. Due to its rarity the clinical phenotype remains poorly defined.

\textbf{Biological phenotype}

Ephrin receptors consist of a N-terminal glycosylated ligand-binding domain, a transmembrane region, and an intracellular kinase domain. Interactions largely occur between neighboring cells; studies using mice revealed a role in thrombus formation and in both contact-dependent and -independent signaling.\textsuperscript{42} Platelets from our family showed decreased aggregation and secretion in response to ADP, arachidonic acid, collagen, and analogs of thromboxane A\textsubscript{2}. The mutation did not affect EPHB2-ephrin interactions but phosphorylation of Lyn, Syk and FcR\textgamma, the initial steps in GPVI signaling and of Src were drastically impaired as was inside-out cDlb3 activation.

\textbf{Genotype}

A homozygous missense variant (p.R745C) in the EPHB2 gene was identified by whole exome sequencing in both siblings; their asymptomatic parents were heterozygous. The p.R745C substitution is located within the tyrosine kinase domain.

\textbf{P2Y12 receptor-related disorder}

The use of ticlopidine as an anti-platelet agent in the 1970s showed the benefit of blocking the interaction between ADP and platelets in anti-ischemic treatment.\textsuperscript{43} The recognition in 1992 of an Italian family with a ticlopidine-like platelet function profile and in 1995 of a French family with a similar defect was crucial to identifying how ADP activates platelets.\textsuperscript{44} Some 20 years later only a few such families have been identified.

\textbf{Definition}

P2Y12-related disease affects the full aggregation response of platelets to ADP and the stabilization of aggregates. Human platelets possess two ADP receptors that act in synergy: (i) P2Y1, a G\textsubscript{\alpha}-coupled receptor, which initiates platelet shape changes and ADP-induced aggregation through the mobilization of internal Ca\textsuperscript{2+} stores, and (ii) P2Y12, a G\textsubscript{\alpha}-coupled receptor linked to adenylyl cyclase and phosphoinositide 3-kinase activation (Figure 3).

\textbf{Clinical phenotype}

The disease is inherited in an autosomal recessive manner and is manifested clinically as moderate to severe mucocutaneous bleeding, including after surgery.\textsuperscript{45,46} Beyond the classical forms, a family with severe hemorrhage has been recently observed with autosomal domi-
nant transmission; other cases with a monoallelic form have mild bleeding.47,50

**Biological phenotype**

ADP, even at high doses, fails to induce full and irreversible platelet aggregation. With other agonists such as thrombin receptor agonist peptide (TRAP), the snake venom protein convulxin or ristocetin, platelet aggregation may remain partially reversible. A key feature is defective Gia-mediated inhibition of platelet adenyl cyclase by ADP with loss of vasodilator-stimulated phosphoprotein (VASP) de-phosphorylation, readily evaluated in a diagnostic test.48 Variants can result in qualitative or quantitative defects of P2Y12, the latter can also be attributed to defective receptor recycling.

**Genotype**

The patient that we first described in 1995 has a two nucleotide deletion in the coding region of P2RY12 moving the reading frame before the introduction of a premature stop (p.I240fs*29).10 Subsequently described mutations with autosomal recessive transmission include those of the initially reported Italian family, heterozygous for two missense mutations, p.R256Q and p.R265W (Figure 3).48,49 Here, two asymptomatic family members carried the heterozygous p.R265W variant. This is important because recently a single allele p.R265P variant, affecting the same amino acid but with a different substitution, was associated with a severe phenotype and autosomal dominant transmission in an unrelated family (Figure 3).47 Significantly, platelet mRNA for this allele was three-fold overexpressed, limiting wild-type homodimer formation (key to ADP-receptor signaling) and introducing a dominant-negative effect.47 Other mutations with a single affected P2RY12 allele and mild bleeding in families with autosomal dominant transmission are shown in Figure 3.46 Missense mutations allowing normal P2Y12 synthesis and affecting domains close to or part of the ADP-binding pockets cause qualitative defects (Figure 3).48,50 Special mention should be made of a novel cytoplasmic domain p.F341A variant affecting the PDZ-binding domain and causing abnormal endosomal recycling of platelet internal pools leading to a surface deficit of P2Y12.21 Interestingly, p.R122C and p.K174E P2RY12 variants have been associated with an intronic polymorphism within FPR (encoding PAR-1) in a patient with chronic bleeding and in a patient with type 1 von Willebrand disease showing that hemorrhage can be part of a complex trait.52,53

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**Figure 3. G-protein-coupled ADP receptors and signaling pathways highlighting P2RY12 variants and their different subgroups.** (A) A schematic representation of ADP-induced signaling pathways. Binding of extracellular ADP to the Gαq-coupled P2Y1 receptor initiates aggregation whereas the Gαi-coupled P2Y12 receptor enhances and sustains the platelet response. Specific signaling pathways are involved with PLCβ and PI3K enabling both RAP1 activation and RASA3 inactivation; both are required for the formation of a stable hemostatic thrombus. The release of ADP from dense granule storage pools after protein kinase C (PKC) activation contributes to the stabilization of thrombi. (B) The schematic representation shows the structure of P2Y12; variants reported as causal for a bleeding syndrome are highlighted (red spots). (C) Summarizes the genotype/phenotype relationship when the variants are regrouped according to bleeding severity and their mode of transmission. The first group consists of patients with autosomal recessive transmission and platelets showing defects of both the binding of ADP analogs and/or receptor cycling. The p.H187Q variant located in the fifth transmembrane domain has normal receptor expression but suppressed function.50 Interestingly p.R256 has a side chain that inserts into a hydrophobic pocket and in docking models it potentially makes contact with the phosphate groups of ADP;52 p.R265W impairs receptor activation and probably alters the conformational state of the receptor. Significantly, a severe phenotype-monoallelic form has a different amino acid substitution on the same residue, p.R265P associated with an increased expression of mutated mRNA and an impaired wild-type homodimer formation, possibly accounting for the bleeding severity. The third subgroup has a mild phenotype and monoallelic expression of mutations that affect receptor function; note that p.P341A located in the PDZ intracellular domain is associated with both abnormal ligand binding and a defect of re-sensitization of the receptor after ADP agonist-induced desensitization. (D) Electron microscopy of platelet aggregates stimulated with 10 μM ADP examined at the peak of aggregation. For the control the platelets are in close contact with some of the cells having lost their granule content. For our patient with the p.I240fs*29 mutation, platelets remain loosely bound and their granule contents are still present, as shown by the immuno gold (black dots) localization of fibrinogen.
Antiplatelet therapy

P2Y12 identification was key to the development of next-generation therapies including the orally available thienopyridines (clopidogrel and prasugrel). These are prodrugs requiring cytochrome P450-based conversion to a short-lived active metabolite that binds covalently to P2Y12 although drug-resistance to clopidogrel can affect up to a third of patients.14 Ticagrelor and cangrelor are direct-acting and rapidly reversible antagonists; cangrelor given intravenously has the advantage of acting immediately.

Other platelet primary receptor defects

Glycoprotein VI-related disease

Rare inherited autosomal recessive defects of the GP6 gene cause a mild bleeding syndrome. GPVI is a receptor for collagen as well as other extracellular matrix components and, as recently identified, fibrin.29,30 A member of the immunoglobulin superfamily, GPVI forms a non-covalent complex with FcRγ and acts in synergy with integrin α2β1. Early evidence of an inherited GPVI deficiency came from a Japanese woman with lifelong although mild mucocutaneous bleeding and platelets refractory to collagen despite responding to other agonists; immunological studies confirmed the specific absence of GPVI.31 The first genotyped patients with GP6 variants belonged to families from France and Belgium with autosomal recessive compound heterozygous mutations.32,33 However, most studies have centered on a cluster of patients from Chile with a homozygous 2 bp insertion followed by a stop at position 242.34 All evidence suggests a founder effect. The Chilean patients had mild bleeding and heterozygous family members were asymptomatic. A defective platelet interaction with fibrin and immobilized fibrinogen helps to explain the diminished thrombus build up under flow.35

Thromboxane prostanoid type α receptor-related disease

The TBXIDA2R gene encodes the thromboxane prostanoid type α receptor (TPα), the major thromboxane A2 (TXA2) receptor on platelets.36 Its activation leads to Ca++ mobilization and platelet aggregation. It associates mainly with G, and principally modulates platelet activation through the PLCβ pathway. Platelet aggregation is characterized by a decreased response to arachidonic acid or TXA2 analogs such as U46619 and I-BOP.37 From as early as 1994, Japanese families were described with a bleeding disorder due to a non-responsive TPα receptor.38,39 A p.R60L variant in the first intracytoplasmic loop of TPα with autosomal dominant transmission predominated, but it was homozygous in one member with mild mucocutaneous bleeding. Globally, three more families were later reported with autosomal dominant transmission; two had missense mutations with a dominant-negative effect linked to impaired receptor dimerization while a third had a frameshift variant, a stop codon and reduced TPα expression.40,41

Antiplatelet therapy

The apparent preservation of hemostasis in the absence of GPVI has prompted research for anti-ischemic drugs and, in particular, treatment for stroke.42 Revaccet is a recombinant soluble dimeric GPVI-Fc fusion protein blocking GPVI-reactive sites in collagen and is the object of ongoing clinical trials. Fab fragments of a monoclonal antibody blocking GPVI, ACT017, have exhibited excellent safety and tolerability profiles in a phase 1 study in humans.43 An exciting challenge for anti-GPVI drugs will be to block thrombus build up through inhibition of the GPVI-TXα2R interaction while preserving the initial interaction with collagen. Anti-GPVI agents may also have a role in cancer therapy by inhibiting platelet-tumor cell interactions; galactin-3 is a novel counter-receptor for GPVI on tumor cells.44 Attempts at treating ischemic disease based on inhibition of TXA2R have largely failed compared to the efficiency of aspirin and related drugs.14

Classic dense granule defects

Much information has been obtained from studies on disorders of platelet lysosome-related organelles.45,46

Hermansky-Pudlak syndrome

Hermansky-Pudlak syndrome (HPS) is named after the Czechoslovakian physicians who in 1959 described two patients with oculocutaneous albinism, prolonged bleeding and pigmented macrophages in the bone marrow.47 Many of the advances regarding HPS have been linked to the numerous mouse and other animal models of storage pool diseases.48,49

Definition

HPS is a heterogeneous autosomal recessive multisystem disorder resulting from defects of ten genes encoding proteins essential for the biogenesis of platelet dense granules, a member of the lysosomal-related organelle family.50,51 Association with albinism reflects defects in the biogenesis of melanosomes in the melanocytes of the skin, hair and choroid of the eye while pigment epithelial cells of the retina can also be affected.

Clinical phenotype

The bleeding diathesis includes easy bruising, epistaxis, gingival bleeding, menorrhagia, postpartum hemorrhage and prolonged bleeding after surgery.52 Platelet transfusions prevent the hemorrhagic risk during surgery, while tranexamic acid is often used preventively or to stop moderate bleeding. Oculocutaneous albinism involves hypopigmentation of the skin and hair associated with characteristic ocular findings. Granulomatous colitis, neutropenia, immunodeficiency and fatal pulmonary fibrosis can feature according to the subtype.53-56

Biological phenotype

Platelet aggregation is characterized by an impaired second wave.57-60 The specific absence of dense granules means that the platelet secretory pool of ADP, ATP, serotonin, calcium, and polyphosphate is reduced or lacking. Dense granules can be evaluated by whole mount electron microscopy due to their opacity to electrons (Figure 4). Total platelet ADP and ATP and the secretion of ATP during platelet aggregation is now mostly measured by bioluminescence. Quantifying mepacrine uptake by flow cytometry is an indirect method for assessing dense granule content. Granule secretion can also be measured by flow cytometry based on the translocation of P-selectin and lysosomal-associated membrane protein (LAMP-3, CD63) to the platelet surface; exposure of LAMP-3 can be spontaneous.
Genotype

Identification of pathogenic variants for HPS involves ten subtypes (HPS1-10) and ten genes: AP3B1, AP3D1, BLOC1S3 encoding for BLOS-3, BLOC1S6 encoding for palladin, DTNBP1 encoding for dysbindin, HPS1, HPS3-6. All of the genes encode subunits organized in four multi-subunit protein complexes, named biogenesis of lysosome-related organelles complex (BLOC)-1, -2 and -3 and the adaptor protein-3 complex (AP-3). The HPS phenotype is related to the BLOC complex affected (Figure 4). BLOC-1 consists of eight subunits of which dysbindin (HPS7), BLOC-3 (HPS8) and palladin (HPS9) are mutated in rare patients with a mild or absent bleeding diathesis and variable hypopigmentation. HPS3, HPS5, and HPS6 are subunits of BLOC-2; mutations in these genes are more frequent than those in the BLOC-1 and affected individuals have a mild clinical phenotype with a bleeding syndrome and hypopigmentation only. HPS1 and HPS4 are subunits of BLOC-3; mutations in these cases are predominantly associated with severe manifestations (severe oculocutaneous albinism, pulmonary fibrosis, granulomatous colitis). BLOC-3 defects are the most frequent form of HPS with p.His497Glnfs*90 in HPS1, the first HPS mutation to be discovered; this form is common in Puerto Rico. The AP-3 complex has four subunits with AP3B1 encoding β3A mutated in HPS-2 and AP3D1 encoding AP-3β8 mutated in HPS-10. AP-3 is important for dense granule biogenesis and protein delivery to the forming granule (e.g., the serotonin transporter, VMAT2). Variants of AP3B1 are said to cause HPS type II. Interestingly, Enders et al. detected a homozygous nonsense mutation in AP3B1 together with a heterozygous bystander RAB27A mutation in a child with bleeding, albinism, developmental delay and a susceptibility to infections; HPS type II was diagnosed rather than Griscelli syndrome as would indicate a RAB27A mutation. HPS-10 is a very rare form of HPS characterized by partial oculocutaneous albinism, bleeding, neutropenia and immunodeficiency. Defective protein disulfide isomerase release from platelets and endothelial cells in HPS contributes to the impaired thrombus formation and could represent a novel target for antithrombotic drugs.

Chediak-Higashi syndrome

The most important clinical problem of patients with Chediak-Higashi syndrome is immunodeficiency with life-

Figure 4. Hermansky-Pudlak syndrome. Syndromic platelet defect resulting from the abnormal biogenesis of dense granules with an autosomal recessive inheritance. (A) Summarizes the genotype/phenotype relationship of different types of Hermansky-Pudlak syndrome (HPS) regrouped according to the four multi-protein complexes involved in different steps of lysosome-related organelle (LRO) biogenesis including vesicle formation and/or trafficking: BLOC1, 2 and 3 and AP-3 with each containing several subunits. Variants in HPS 6, 7 and 8 are causal in BLOC1 but with mild clinical consequences. For BLOC2, variants in HPS 3, 5 or 6 give syndromes of moderate severity; they are rare but a variant of HPS 3 occurs in Puerto Rican communities. The most frequent causes of HPS are BLOC3 defects (variants in HPS 1 or 4). The syndromes are severe and are associated with lung fibrosis and gastrointestinal defects; in Puerto Rico a community is affected by a common HPS3 variant. In the last group, defects of AP-3 are associated with neutropenia and infections. (B) A scheme illustrates the major steps of granule biogenesis. Dense granules may derive from the endolysosomal system, including either early or late endosomes. As secretory granules the membrane constituents including membrane transporters come from the Golgi complex. A network of interconnected and functionally distinct tubular subdomains transports their cargoes along microtubule tracks from the endosomes. Tubules ferry contents from the trans-Golgi network and the plasma membrane to LRO, a system that requires the coordination of numerous effectors. Components stored in dense granules or associated with their membranes are shown. (C) Platelets from a control and a patient with HPS3 examined by electron microscopy as whole mounts are shown. The dense granules observed in controls as black spots are absent from the patient’s platelet.

AP-3: adaptor protein-3; BLOC: biogenesis of lysosome-related organelles complex; LRO: lysosome-related organelle; OCA: oculocutaneous albinism; MVB: multivesicular body.
threatening infections. There is mild bleeding with a partial to severe reduction of dense granules. It is a rare but fatal autosomal recessive disorder with multiple clinical features, including hypopigmentation of skin and hair, reduction of leukocytes, giant organelles in circulating granulated cells, and neurological dysfunction. Recurrent infections are associated with defective function of natural killer cells. Immune dysregulation often leads to lymphoproliferative histiocytosis with an accelerated phase that determines the prognosis. Platelet function defects resemble those seen in HPS. The LYST gene (also called CHS1) alone is responsible for this disease and a wide variety of mutations have been described. Chediak-Higashi syndrome is particularly prevalent in Puerto Rico. LYST is a member of the BEACH protein family and is required for sorting resident endosomal proteins into late multivesicular endosomes (Figure 4B).

Other defects involving platelet granules

As well as the above disorders, qualitative defects can lead to decreased dense granule content, inability to secrete their contents or combined dense- and α-granule deficiency. An absent second wave of aggregation in response to ADP and collagen is characteristic. This can be secondary to mutations of genes for membrane receptors: P2RY12, GP6, TXBA2R, EPHB2, G6b-B; or genetic defects often accompanied by thrombocytopenia (Online Supplementary Table S1). Applying next-generation sequencing procedures targeting 529 preselected genes, Leo et al. analyzed 18 patients including six with secretion defects, highlighting 49 genes not previously known to be causal of IPD. Gorski et al. performed a similar study using whole exome sequencing of 14 patients with bleeding caused by primary platelet secretion defects. Both studies exposed the complications of whole exome sequencing and next-generation sequencing when isolated gene variants are identified as potentially causal without statistical validation or comprehensive biological studies or usage of mouse models.

A specific condition with defective granule secretion from platelets is familial hemophagocytic lymphohistiocytosis, a rare primary immunodeficiency syndrome in which autosomal recessively inherited variants of UNC13D, STX11 and STXBP2 can result in defects in the secretory machinery of dense bodies and α-granules as well as lysosomes.

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**THERAPEUTIC TOOLS TO PREVENT OR TREAT BLEEDING SYNDROMES**

**DAILY LIFE**

- Preventive actions:
  - Provide a card to the patient indicating essential medical information to prevent or treat bleeding and with their bleeding score
  - Establish a list of medications to avoid
  - Recommend a lifestyle in line with the hemorrhagic risk e.g. avoid contact sports, etc...
  - Recommend regular dental care, hormonal therapy to reduce menorrhagia
  - Evaluation of the need for iron supplementation

**SURGERY**

- Bleeding risk must be established by a multidisciplinary staff and a protocol planned

**PREGNANCY**

- Evaluation of the risk of bleeding during delivery
- Prophylactic platelet transfusions and/or anti-fibrinolytics

**RECURRENT SEVERE BLEEDING WITH POOR RESPONSE TO TREATMENT**

- Stem cell transplantation: GT, LAD-III
- Gene therapy for GT patients on the horizon

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*Figure 5. Schema with management options for patients with inherited defects of platelet function. It is important to limit risks in daily life. It is also important to provide each patient with a card detailing the type of disorder and proposing the medication to be used according to the bleeding risk, in particular when platelet expert medical centers are unavailable to take charge of the patient’s needs. While the main therapeutic approaches for all disorders, including those patients with Glanzmann thrombasthenia (GT), RASGRF2 (CaldAG-GEFI)-related disease, and leukocyte adhesion deficiency III (LAD-III) syndrome for whom bleeding can be severe, are similar, some situations (e.g., isocytoband formation in GT) or infections in LAD-III syndrome need special care. Surgery requires a multidisciplinary consensus to evaluate the risk of bleeding, the benefit-risk ratio of prophylaxis, and to assess therapeutic efficiency. Pregnancy is always a challenge, from women with mild bleeding risks for whom minimal measures are needed to those with a severe bleeding history and for whom maximal prophylactic care is required. Here again, the management must be planned between obstetricians, anesthetists and hematologists. The contraindication of epidural or spinal anesthesia for those with mild risk remains a difficult problem, as is the choice of vaginal delivery or Cesarean section for the most severe forms. In the figure, it is noted that for mild bleeding risk, an antifibrinolytic medication or desmopressin is preferred as the first line of bleeding prevention and that platelet transfusions should be considered only when these approaches are insufficient. There is a risk of allo- or iso-immunization which, if it occurs, can be a lifelong handicap, and recombinant activated factor VII is increasingly recommended. Finally, stem cell transplantation is reserved for young patients with recurrent, severe bleeding that fails to respond to standard treatments and when the survival of the patient is at risk. Gene therapy is not yet an option for the disorders covered in this review but will probably become available in the future.*
Cytosolic signaling pathways

Rao and his colleagues were pioneers in identifying candidate proteins in patients with mild bleeding diatheses deficient in key signaling pathways and second messengers.31 Geq, PLC-β2, as well as defects in Ca2+-mobilization and protein kinase C-induced phosphorylation were highlighted. Abnormalities extended to granule secretion as well as αIIbβ3 activation. However, most of the early patients still await full genetic analysis. We have chosen to highlight three genes involved in arachidonic acid metabolism in platelets.

The first is a defect of PLA2G4A encoding cytosolic PLA2 in three families. In the first family compound heterozygous missense mutations induced marked structural changes within cPLA2. In the second family heterozygous p.S111P and p.R485H missense variants respectively affected cPLA2 trafficking and abrogated the enzyme’s catalytic activity; while for the third family a homozygous p.D575H variant was present.24-26 In all three cases, platelet aggregation and dense granule secretion with collagen and ADP were impaired; gastrointestinal bleeding was a feature.

An inherited defect of cyclooxygenase-1 (COX-1) is more specifically linked to defective platelet aggregation with arachidonic acid and absence of the second wave response to epinephrine, ADP and collagen. While failing to respond to arachidonic acid, platelets aggregate normally with TXA2 agonists. Several such patients with mild bleeding have been reported either with autosomal dominant or, more recently, autosomal recessive transmission of variants in PTGS1.27-29 Patients may lack COX-1 or have a functionally defective but normally expressed enzyme. Bastida et al., using a high-throughput sequencing gene panel, found a heterozygous variant with an aspirin-like platelet functional defect linked to a heterozygous p.N143S missense mutation in PTGS1.30 Very recently, whole genome sequencing helped to identify a homozygous missense variant (p.W522S) in a consanguineous family of Iranian descent with mild bleeding.

The third disease is Ghosal syndrome, a disorder with increased bone density that was linked to a deletion mapped to chromosome 7q33/34. The deletion includes TBXAS1, the gene for thromboxane synthase, a terminal enzyme in the arachidonic acid cascade converting prostaglandin H2 into TXA2. A key finding was homozygous missense mutations alone in TBXAS1 in consanguineous families with Ghosal syndrome in ethnic groups; the affected residues were all highly conserved.30 Platelets from the patients had reduced aggregation and secretion in response to arachidonic acid; in contrast the response to U46619 directly activating TPα was normal. A role for TXA2 in bone remodeling is also indicated. Somewhat surprisingly, none of the patients had a bleeding diathesis (distinguishing them from patients with congenital defects of TBXA2R).

Therapy

The management of IPD involves both prevention of bleeding and treatment of mild and severe bleeding episodes and differs little for disorders of platelet count with the exception that the recent use of thrombopoietin mimetics is not applicable.31 As detailed in Figure 5, strategies involve the maintenance of a lifestyle that minimizes risk and being prepared in case of an emergency. Procedures range from local measures to the use of platelet transfusions or recombinant activated FVII to stop major bleeding as well as the establishment of and adherence to standardized protocols for prophylaxis prior to surgery or childbirth.15,21-26 A special case is type I GT in which the absence of αIIbβ3 can lead to the formation of isoantibodies, making patients refractory to normal platelets.

Conclusions

Of the disorders covered by this review, GT is the most frequent and widely screened disease.45-47 Genotyping has now identified several hundred disease-causing mutations in a constantly changing spectrum, so the classification of patients into type I and type II subgroups based on residual platelet αIIbβ3 content should be updated to take into account mutation analysis.41,43 From 2001, the year in which P2RY12 was first characterized,48 newly identified genes include FERMT3, ANO6, RASGRP2 and EPHB2 thereby expanding the gene repertoire for IPD.25,26,33,34,35,43 The development of next-generation sequencing platforms targeting IPD genes,6,88 together with widening access to whole exome sequencing,47 and now whole genome sequencing,4 has both increased the number of patients with a genetic diagnosis and the speed at which this is performed. The multiplication of data has underlined the great diversity of variants responsible for gene dysfunction, the rarity of hot-spot mutations, the frequency of compound heterozygosity within the same gene (limiting the notion of consanguinity for recessive diseases); and raised the likelihood of additive effects of heterozygous variants on different genes modulating the phenotype. It is also worthy of mention that some inherited thrombocytopenias with associated functional platelet defects (Online Supplementary Table S1) may not have complete penetrance and some patients may therefore not be thrombocytopenic (e.g., those with mutations of RUNX1 and FLI1).44 The increasing understanding of the gene spectrum causing IPD is leading to better care of patients and it should be underlined how major anti-ischemic treatments have been initiated or improved as a result of this.

Future strategies

Despite the progress in gene identification a significant proportion of patients still fail to be genotyped. One successful strategy for the identification of causal genes in patients with inherited thrombocytopenia was by comparison with mouse models with similar phenotypes.60-62 Another approach is ongoing screening of local whole exome or whole genome sequencing databases of patients for variants of each newly published IPD gene. A third strategy involves screening large families or those with consanguinity, as done for RASGRP2 and EP HB2, or statistical analysis of large cohorts of patients, aligning variant detection in unrelated families with a similar phenotype.63 The final proof that variants in new genes are causal will still require their expression in heterologous cells, the use of in silico modeling to assess changes in protein structure,
or their reproduction in mouse, zebrafish or other animal models. When no genetic diagnosis has been achieved with standard approaches, gene analysis should be expanded to evaluate complex copy number variations, deep variants in non-coding regions and secondary defects involving the action of microRNA or retrotransposons, as we have detailed elsewhere.11

Currently the diagnosis of IPD is based largely on the concept of a monogenic heritable platelet disease, but a proportion of these diseases may be di- or polygenic in origin. Families with unexplained heritability may have unrecognized variants or other changes in regulatory regions in combinations that only advanced technologies may help to resolve. Thus in a large cohort of patients with primary immunodeficiency, genome-wide association studies have revealed loci that modulate platelet function with the co-localization of and interplay between novel high-penetrate monogenic variants and common variants.24 It is reassuring that IPD without thrombocytopenia are not associated with a propensity to develop hematological malignancies or bone marrow aplasia or fibrosis, as in some inherited thrombocytopenias such as RUNX1-related disease and ETV6-related disease.

Current advances suggest the need to redefine the circuit for patient screening from the first consultation. An information-gathering first consultation, blood cell counts and basic coagulation screening remain essential as is assigning a bleeding score and determining whether the platelet function abnormalities are isolated or syndromic.25 But flow charts with extensive biological testing in order to select the gene(s) to analyze need to be revisited and the choice of extensive biological testing before genotyping questioned. Platelet aggregation, flow cytometry and the examination of blood smears provide early information to guide the diagnosis.26,27 Nevertheless, when clinical and biological data are in favor of an IPD, in selected cases upfront DNA analysis may save time, money and improve patients’ care: however longitudinal studies are required to substantiate this expectation. For patients without access to a specialized platelet center, sending blood and DNA samples to expert centers is recommended. Validation of novel variant(s) on next-generation sequencing targeted gene platforms or after whole exome or whole genome sequencing requires confirmation by co-segregation or other studies as discussed above and it is here that selected biological phenotyping may be necessary.

As part of this roadmap, national centers regrouping platelet experts have a key inter-connecting role between the patients, local hematology laboratories and genomic platforms. In France, our creation of a Reference Center for IPD, led to the organization, on a national basis, of platelet studies with European links to next-generation sequencing and the preparation of personalized follow-up both for the patient and the local general practitioner. In this context it is indispensable that the medical personnel directly responsible for the patient remain in control of the diffusion of information both to the patient and in the scientific literature and especially take into account ethical considerations.26 In everyday clinical practice a patient-centered approach governed by the overarching principle of patient utility versus in-depth, detailed laboratory and genetic characterization is acceptable and in general sufficient for patients’ care.28 On the other hand, our understanding of these disorders cannot be expanded without advanced research involving modern technologies. Here, informed explicit collaboration of the patient should be obtained.

Disclosures
No conflicts of interests to disclose.

Contributions
All the authors participated in the manuscript preparation. PN and ATN supervised the writing of the text.

Acknowledgment
We thank Xavier Pillois for his contribution to the section on GT and for the Pymol model of cDIIIβ3.

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Inherited platelet function disorders


