

Inducible deletion of CDK4 and CDK6 – deciphering CDK4/6 inhibitor effects in the hematopoietic system

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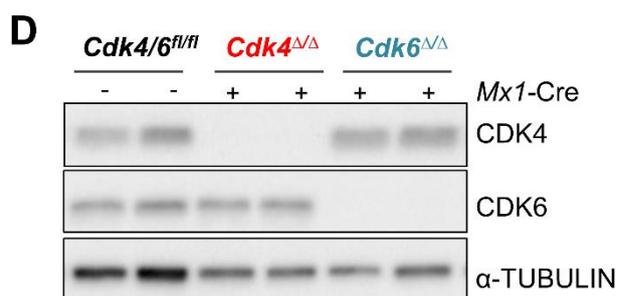
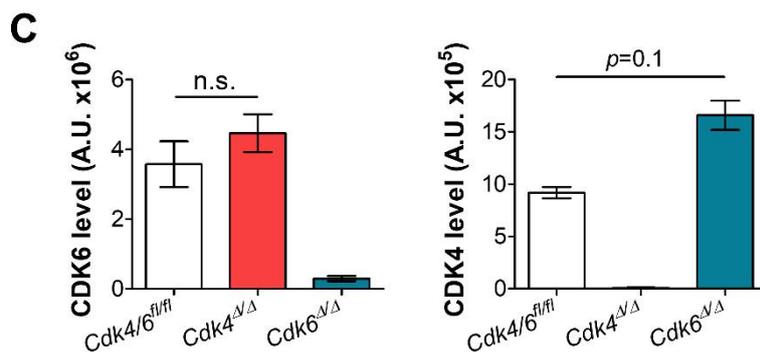
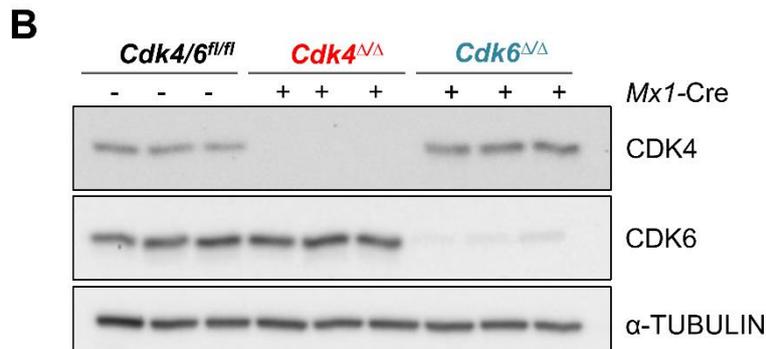
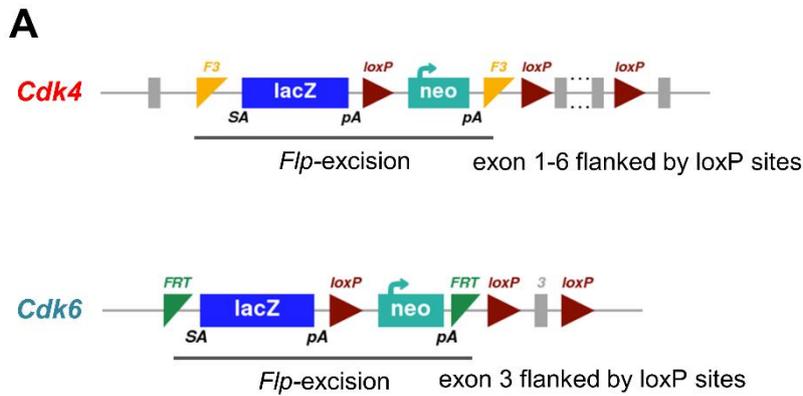
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Supplementary Information

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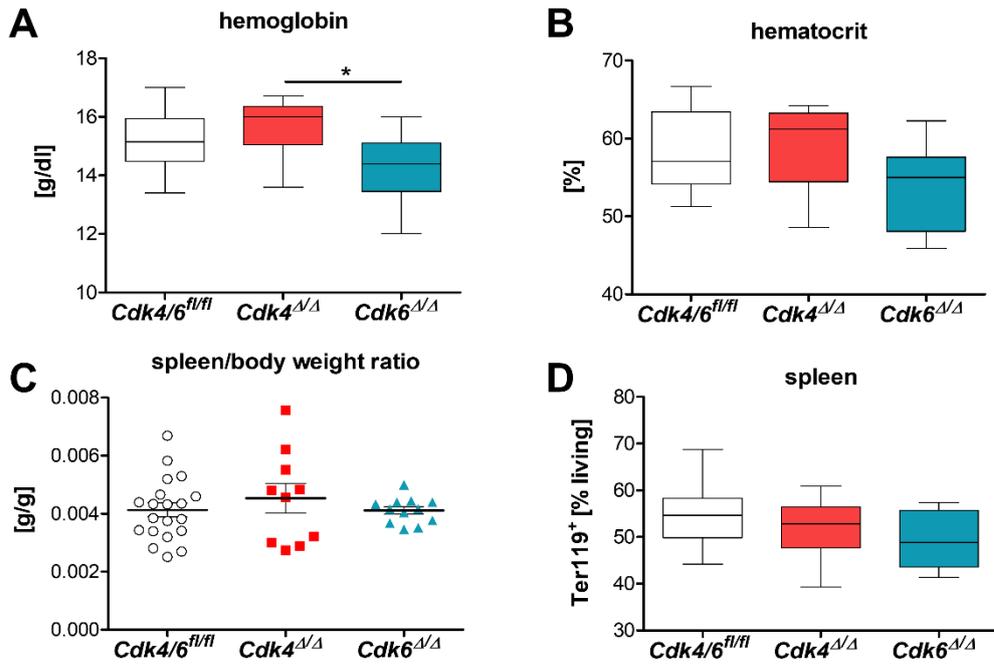


Supplementary Figure 1: Reduced CDK4 or CDK6 expression in *Cdk4^{fl/fl}* or *Cdk6^{fl/fl}* *Mx1-Cre* mice

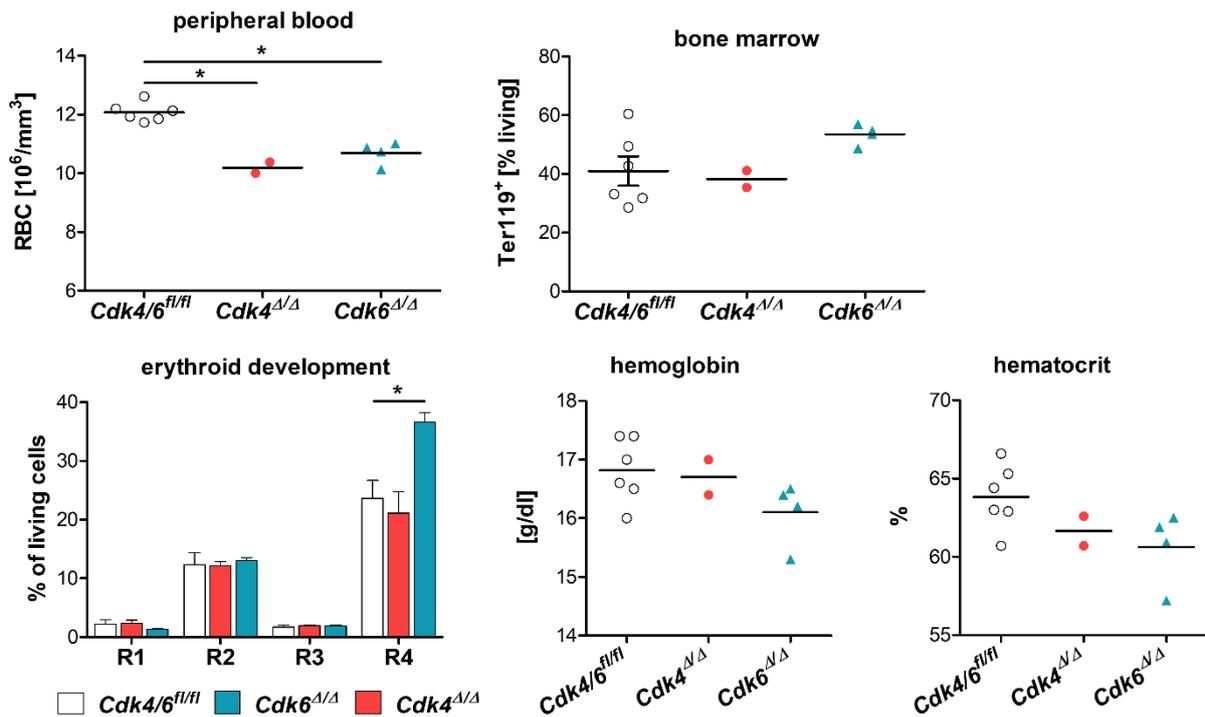
(A) Targeting strategy of *Cdk4* (top) or *Cdk6* (bottom). Respective floxed exon(s) were derived by *Flp*-excision of the neo selection cassette and lacZ reporter. (B) Immunoblotting: levels of CDK4 and CDK6 in bone marrow (BM) cells (*Cdk4^{fl/fl}* or *Cdk6^{fl/fl}*, *Cdk4^{Δ/Δ}*, *Cdk6^{Δ/Δ}*, n=3/genotype). α-TUBULIN served as a loading control. Analysis was performed three weeks post final poly(I:C) injection. A representative blot of at least three independent experiments is shown. (C) Quantification of immunoblot shown in (B),

signal intensities were normalized to α -TUBULIN levels (A.U.; arbitrary units). *Cdk4* ^{$\Delta\Delta$} or *Cdk6* ^{$\Delta\Delta$} protein levels were compared to the respective controls performing Mann-Whitney U tests. **(D)** Immunoblotting: levels of CDK4 and CDK6 in BM cells (*Cdk4*^{fl/fl} or *Cdk6*^{fl/fl}, *Cdk4* ^{$\Delta\Delta$} , *Cdk6* ^{$\Delta\Delta$} , n=2/genotype). α -TUBULIN served as a loading control. Analysis was performed six weeks post final poly(I:C) injection.

Analysis 3 weeks after poly I:C injection



E Analysis 6 weeks after poly I:C injection

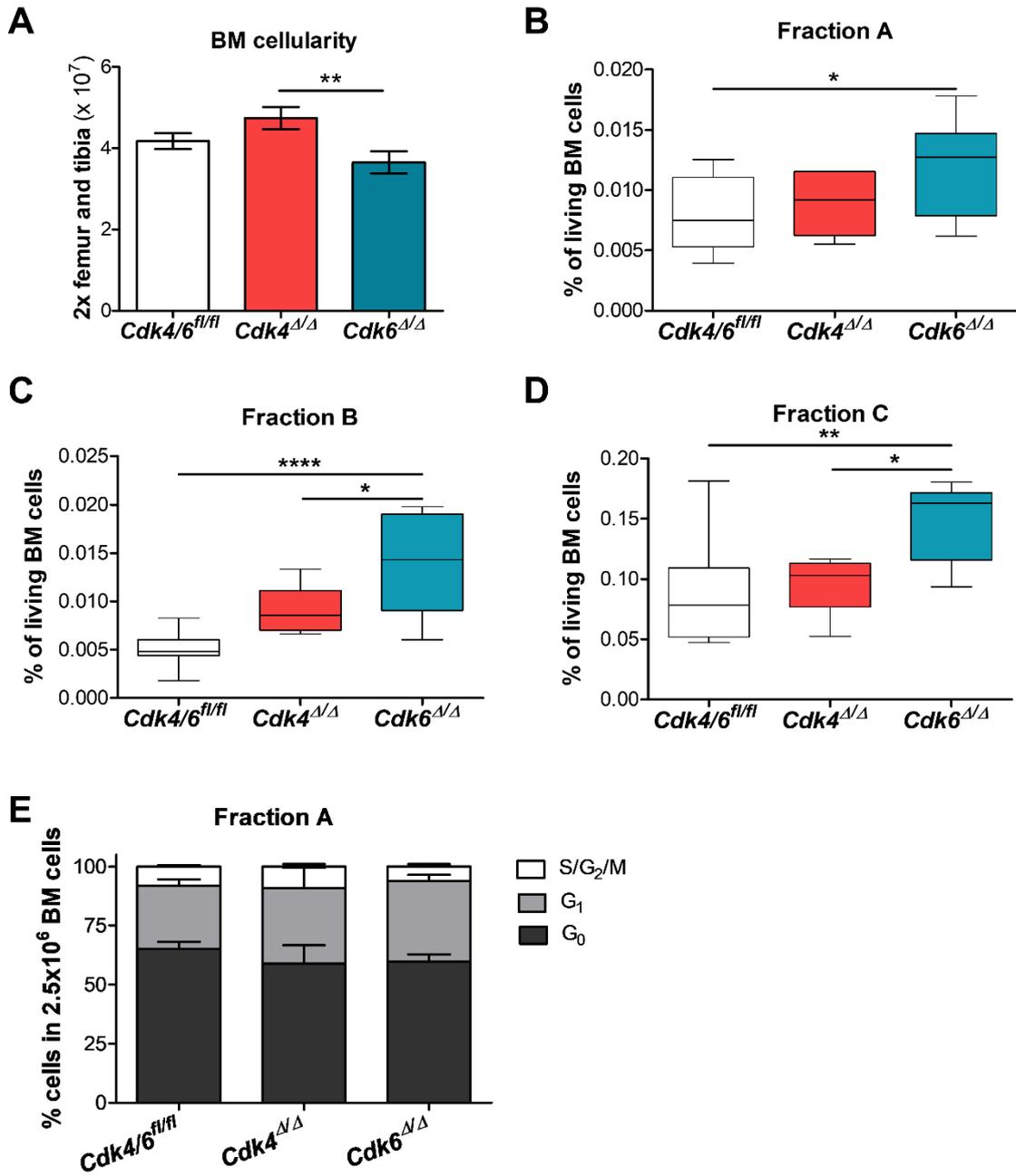


Supplementary Figure 2: *Cdk6^{Δ/Δ}* mice are anemic with compensatory upregulation of erythrocytes in the bone marrow

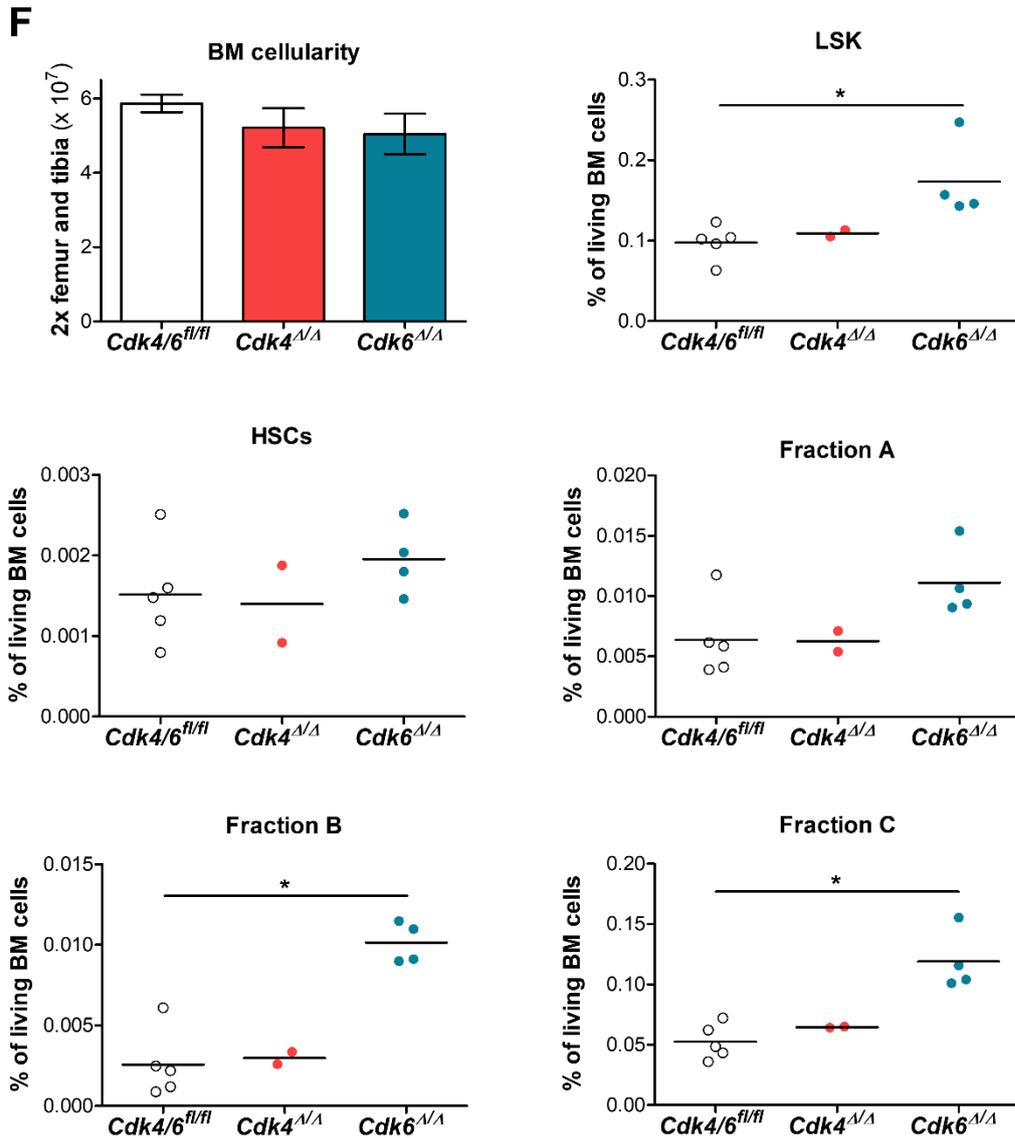
(A-D) Analysis was performed three weeks post final poly(I:C) injection. (A) Hemoglobin and (B) hematocrit levels in peripheral blood (*Cdk4^{Δ/Δ}*; *Cdk6^{Δ/Δ}*; *Cdk4/6^{fl/fl}*, $n \geq 8/\text{genotype}$). (C) Spleen to body weight ratio (*Cdk4^{Δ/Δ}*; *Cdk6^{Δ/Δ}*; *Cdk4/6^{fl/fl}*, $n \geq 10/\text{genotype}$). (D) Percentage of Ter119⁺ cells in spleen (*Cdk4^{Δ/Δ}*; *Cdk6^{Δ/Δ}*; *Cdk4/6^{fl/fl}*, $n \geq 8/\text{genotype}$). All comparisons were done with one-way ANOVA followed by Tukey's Multiple Comparison Test, $*P \leq 0.05$. (E) All analysis shown in Figure 2 and Supplementary

Figure 2A-B were repeated 6 weeks post final poly(I:C) injection. Comparisons were done with Kruskal-Wallis test followed by Dunn's Multiple Comparison Test, $*P \leq 0.05$.

Analysis 3 weeks after poly(I:C) injection

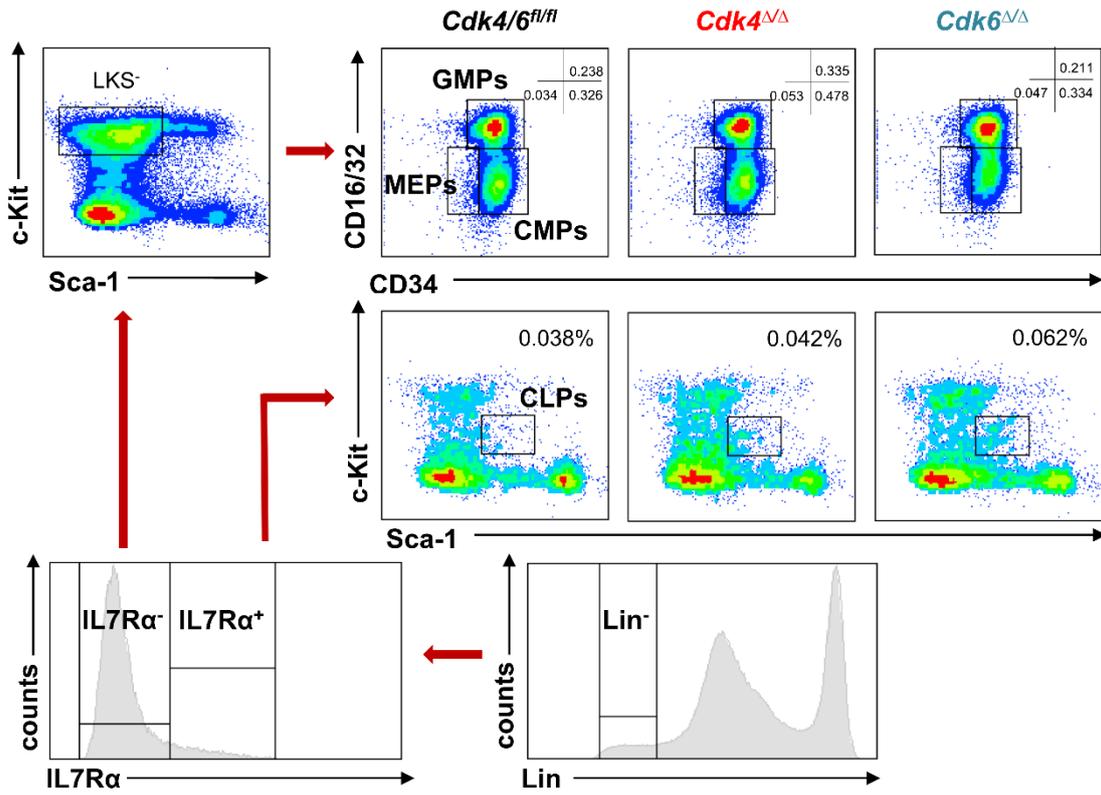
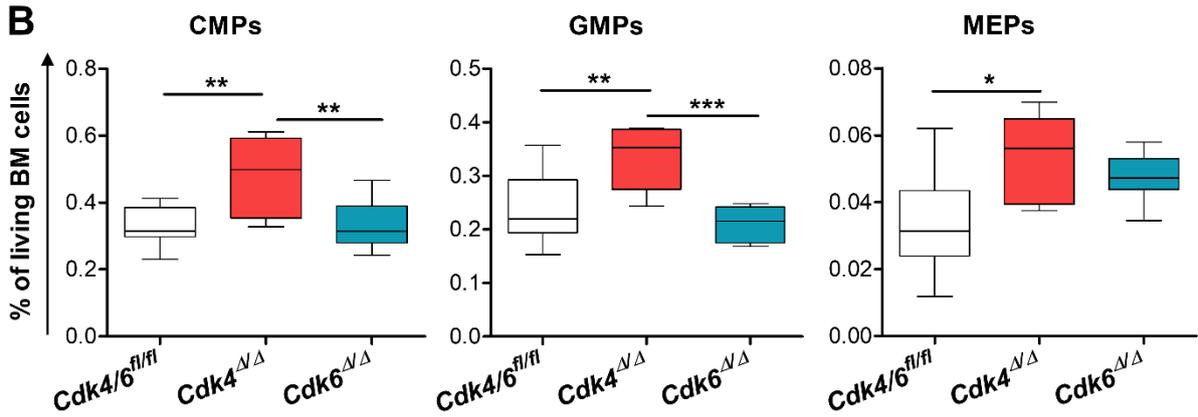


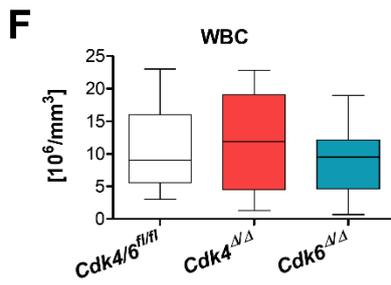
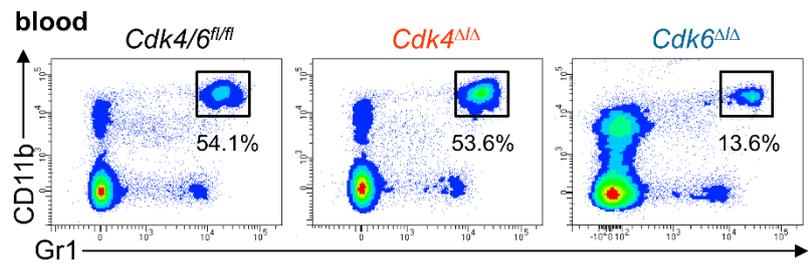
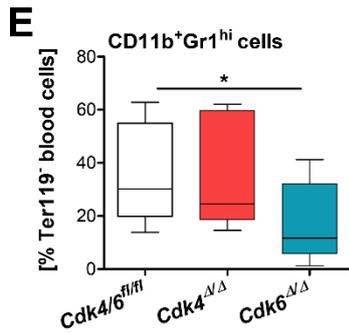
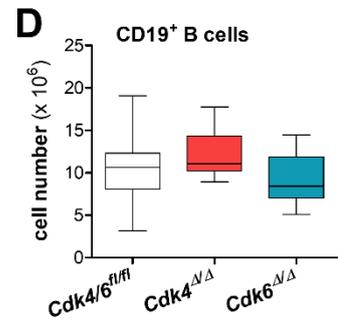
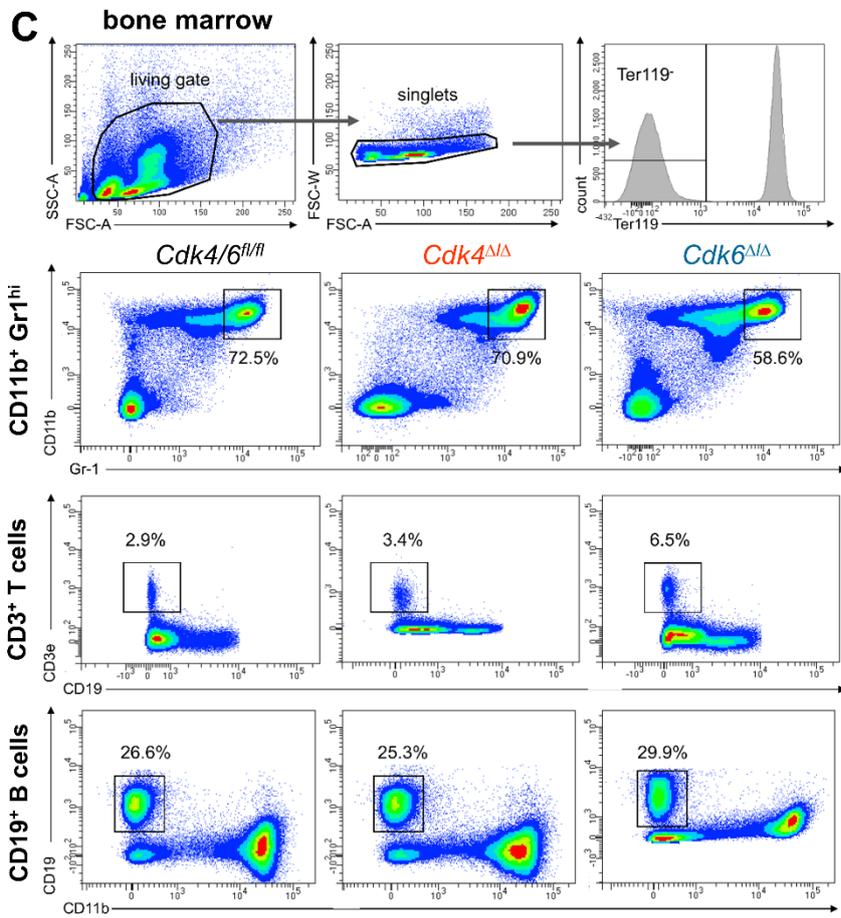
Analysis 6 weeks after poly(I:C) injection

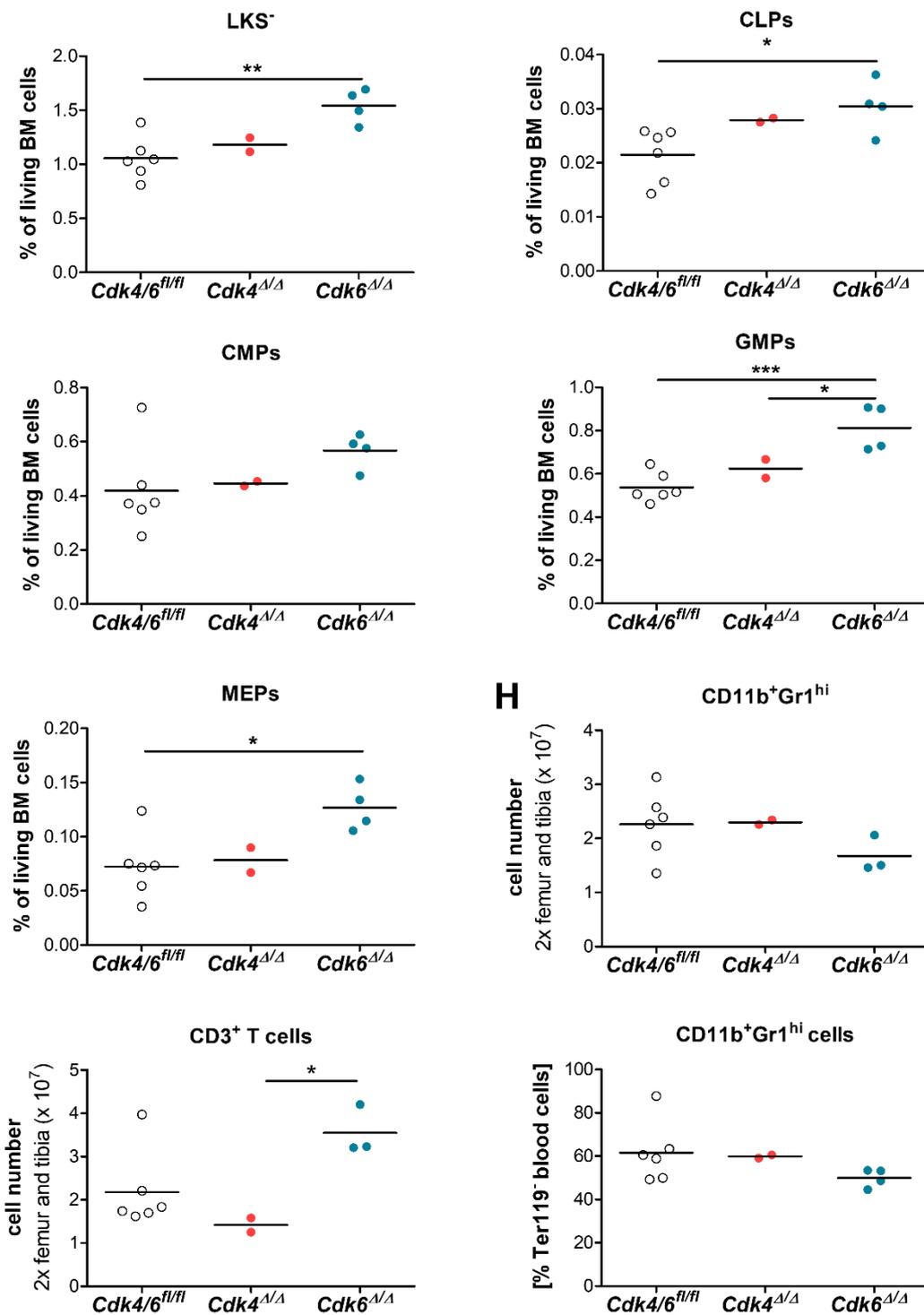


Supplementary Figure 3: CDK6 alters the composition of the hematopoietic stem and progenitor pool

(A-E) Analysis was performed three weeks post final poly(I:C) injection. (A) Bone marrow (BM) cellularity in both femur and tibia per mouse (*Cdk4^{Δ/Δ}*; *Cdk6^{Δ/Δ}*; *Cdk4/6^{fl/fl}*, $n \geq 6$ /genotype). Percentage of (B) Fraction A (CD150⁺CD48⁻LSKs), (C) Fraction B (CD150⁺CD48⁺LSKs) and (D) Fraction C (CD150⁻CD48⁺LSKs) hematopoietic stem cells per 2.5×10^6 BM cells (*Cdk4^{Δ/Δ}*; *Cdk6^{Δ/Δ}*; *Cdk4/6^{fl/fl}*, $n \geq 5$ /genotype). (E) DAPI/Ki-67 cell cycle staining of Fraction A cells. Depicted are % cells G₀, G₁ or S/G₂/M per 2.5×10^6 BM cells ($n=3$ (*Cdk4^{Δ/Δ}*); $n=3$ (*Cdk6^{Δ/Δ}*); $n=6$ (*Cdk4/6^{fl/fl}*)). All comparisons were done with one-way ANOVA followed by Bonferroni's Multiple Comparison Test, * $P \leq 0.05$, ** $P \leq 0.01$, *** $P \leq 0.001$. (F) Analysis six weeks post final poly(I:C) injection: BM cellularity in both femur and tibia per mouse. Percentage of LSKs, Fraction A, Fraction B and Fraction C cells and hematopoietic stem cells (HSCs) (CD150⁺CD48⁻CD135⁻CD34⁻LSKs) per 2.5×10^6 BM cells. Comparisons were done with Kruskal-Wallis test followed by Dunn's Multiple Comparison Test, * $P \leq 0.05$.

A**B**



G**Analysis 6 weeks after poly(I:C) injection****Supplementary Figure 4: CDK4 and CDK6 drive the myeloid and lymphoid progenitor pool in opposite directions**

(A-F) Analysis was performed three weeks post final poly(I:C) injection. (A) Representative flow cytometry gating strategy of common lymphoid progenitors (CLPs), myeloid-primed progenitors (Lin-c-Kit+Sca-1; LKS⁻), common myeloid progenitors (CMPs), granulocyte/macrophage progenitors (GMPs) and megakaryocyte/erythroid progenitors (MEPs) in *Cdk4/6^{fl/fl}*, *Cdk4^{Δ/Δ}* and *Cdk6^{Δ/Δ}* bone marrow (BM). (B) Percentage of CMPs (LKS⁻CD34⁺CD16/32^{lo}), GMPs (LKS⁻CD34⁺CD16/32^{hi}) and MEPs (LKS⁻CD34⁻CD16/32^{lo}) per 1*10⁶ BM cells (*Cdk4^{Δ/Δ}*; *Cdk6^{Δ/Δ}*; *Cdk4/6^{fl/fl}*, n≥5/genotype). (C) Representative flow

cytometry gating strategy of CD11b⁺Gr1^{hi} cells, CD3⁺ T cells and CD19⁺ B cells in *Cdk4/6^{fl/fl}*, *Cdk4^{Δ/Δ}* and *Cdk6^{Δ/Δ}* BM. **(D)** CD19⁺ B cells in BM (2 femurs and tibiae; *Cdk4^{Δ/Δ}*; *Cdk6^{Δ/Δ}*; *Cdk4/6^{fl/fl}*, n≥8/genotype). **(E)** Left: Relative numbers of CD11b⁺Gr1⁺ myeloid cells in blood (gated on Ter119⁻ cells; *Cdk4^{Δ/Δ}*; *Cdk6^{Δ/Δ}*; *Cdk4/6^{fl/fl}*, n≥8/genotype). Right: Representative flow cytometry gating. **(F)** White blood cells (WBC) in peripheral blood (*Cdk4^{Δ/Δ}*; *Cdk6^{Δ/Δ}*; *Cdk4/6^{fl/fl}*, n≥15/genotype). All comparisons were done with one-way ANOVA followed by Bonferroni's Multiple Comparison Test, **P*≤0.05, ***P*≤0.01, ****P*≤0.001. Analyses shown in **(G)** Figure 4A-B, Supplementary Figure 4A-B, **(H)** Figure 4C-D and Supplementary Figure 4E were repeated six weeks after the final poly(I:C) injection. Comparisons were done with Kruskal-Wallis test followed by Dunn's Multiple Comparison Test, **P*≤0.05; ** *P*≤0.01.

Supplementary Methods

Genotyping and deletion PCR

***Cdk4*^{fl/fl}**: Genomic DNA was amplified with the following primers: *Cdk4*_{wt} (forward: TGCAGAATCTTCGGTGCAAACCCTG, reverse: ATGACCCGGCGGTAACAAAGGAACTC) resulting in a 244 bp product in mice harboring a wildtype allele, and *Cdk4*_{loxP} (forward: GCGCAACGCAATTAATGATAAC, reverse: GCAGTGACAACACTACAGCCTGCCAC) resulting in a 346 bp product in mice harboring a loxP-flanked *Cdk4* allele at the following conditions: 3 min 95 °C; 35 x 95 °C 15 sec., 60 °C 15 sec., 72 °C 20sec; 72 °C 1 min, hold at 8 °C. Using the *Cdk4*_{wt} forward and *Cdk4*_{loxP} reverse primer, the deletion band (*Cdk4*^{Δ/Δ}, ~ 400 bp) can be amplified.

***Cdk6*^{fl/fl}**: Genomic DNA was amplified with the following primers: *Cdk6*_{tm1c_wt} (forward: ATCCATGTTTGGAGCACCTTTGGAGAG, reverse: TGAGCCAACGTAAGCCCTAGCAATG) resulting in a 245 bp product in mice harboring a wildtype allele and a 430 bp product in mice harboring a loxP-flanked *Cdk6* allele at the following conditions: 3 min 95 °C; 35 x 95 °C 15 sec., 60 °C 15 sec., 72 °C 20sec; 72 °C 1 min, hold at 8 °C. By adding the *Cdk6*_{delta} primer (AACAGAAAGGGCCAGATGACCATGC), the deletion band (*Cdk6*^{Δ/Δ}, ~ 600 bp) can be amplified.

Hematocytometry

Peripheral blood was collected by *vena facialis* puncture into EDTA-coated tubes (Mini-Collect K3EDTA tubes, Greiner Bio-One). Automated complete blood counts were obtained from a hematology analyzer (scil Vet ABC). Composition of immune cells was analyzed by flow cytometry.

Immunoblotting

Whole-cell lysates of BM or spleen were lysed in 1x Laemmli buffer, incubated at 95°C for 5 min and sonicated for 15 min at RT. Protein concentrations were determined using Pierce™ BCA Protein Assay Kit (Thermo Scientific, Waltham, MA, USA). Proteins were resolved by a 10% sodium dodecyl sulphate polyacrylamide gel and transferred to Immobilon®-P Polyvinylidene difluoride membrane (Merck, Darmstadt, Germany). Membranes were blocked in 5% milk for 1 h and probed with the appropriate antibody overnight. Detection of bound antibodies was performed by incubation with horseradish peroxidase-conjugated anti-rabbit (Cell Signaling Technology, Danvers, MA, USA, 7074S) or anti-mouse (CST, 7076S) antibodies at room temperature for 1 h followed by chemiluminescent visualization

after incubation of the membranes with 20X LumiGLO® Reagent and 20X Peroxide (CST) using the ChemiDoc™ Imaging System (Bio-Rad, Hercules, CA, USA) according to manufacturer's protocol. Anti-CDK4 (C-22), anti-CDK6 (SPM 383) and α -TUBULIN (DM1A) were obtained from Santa Cruz (Dallas, Texas, USA). Immunoblots were quantified using Image Lab 5.2.1 software (Bio-Rad).