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Hematopoietic recovery and immune reconstitution after axicabtagene ciloleucel in patients with large B-cell lymphoma

Running title: Immune reconstitution after axi-cel in lymphoma


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Abstract: 200; Text: 2007; Tables: 2; Figures: 2; References: 25
Abstract

Chimeric antigen receptor (CAR) T-cell therapy targeting CD19 may be associated with long-term adverse effects such as cytopenia and immune deficiency. To characterize these late events, we analyzed 31 patients with relapsed or refractory large B-cell lymphoma treated with axicabtagene ciloleucel at our institution on 2 clinical trials, ZUMA-1 (NCT02348216) and ZUMA-9 (NCT03153462). Complete blood counts, lymphocyte subsets, and immunoglobulin levels were measured serially until month 24 or progression. Fifteen (48%) patients had grade 3-4 cytopenia, anemia (5 [16%]), neutropenia (9 [29%]), or thrombocytopenia (13 [42%]) at day 30. Cytopenia at day 30 was not significantly associated with later diagnosis of myelodysplasia. Among patients with ongoing remission, grade 3-4 cytopenias was observed in 1/9 (11%) at 2 years. While peripheral CD8$^+$ T cells recovered early, CD4$^+$ T cell recovery was delayed with a count of <200/μL in 3/9 (33%) patients at 1 year and 2/7 (29%) at 2 years. Immunoglobulin G levels normalized in 5/9 (56%) patients at 2 years. Thirteen (42%) patients developed grade 3-4 infectious complications, including herpes zoster and Pneumocystis jiroveci pneumonia. These results suggest the need for prolonged monitoring and prophylaxis against opportunistic infections in these patients to improve the long-term safety of axicabtagene ciloleucel therapy.

Clinicaltrials.gov:

ZUMA-1 (NCT02348216)

ZUMA-9 (NCT03153462)
**Introduction**

Chimeric antigen receptor (CAR) T-cell therapies targeting CD19, such as axicabtagene ciloleucel, have significantly improved the outcome of patients with refractory large B-cell lymphoma (LBCL). While acute toxicities, such as cytokine release syndrome and neurotoxicity, have been well reported, late adverse effects, such as cytopenia and immune deficiency, have not been well characterized.

Cytopenias lasting beyond day 30 post infusion have been observed with most CAR T-cell products, suggesting a class effect rather than a complication of conditioning chemotherapy. In addition, prolonged B-cell aplasia is a known on-target off-tumor effect of anti-CD19 CAR T-cell therapies. It is unclear at this time how long it takes for cytopenias to resolve and at what point these patients should be worked up for myelodysplastic syndrome (MDS). It is also unknown what the timing of T-cell immune reconstitution is after axi-cel therapy and whether these patients are at short or long-term risk of infectious complications.
Methods

Patient selection and evaluation

This is a post-hoc analysis of patients with refractory LBCL treated with axi-cel on the pivotal ZUMA-1 study (NCT02348216; N=24) or ZUMA-9 (NCT03153462; N=7); the latter was an expanded access trial for patients not eligible for ZUMA-1 study (cohort 1) or whose commercial product was out-of-specification (OOS)(cohort), and 2 patients from cohort 2 were included in this analysis. Patients were treated at MD Anderson Cancer Center between 01/2015 and 03/2018 (data cut-off: 12/19/2018). The studies were approved by the Institutional Review Board and conducted in accordance with institutional guidelines and the principles of the Declaration of Helsinki.

Conditioning chemotherapy and axi-cel infusion were administered as previously described.(1, 2) Clinical and laboratory features were collected prospectively in all patients who had a complete blood count (CBC) performed 30 days (+/- 5 days) after axi-cel infusion. Pre-conditioning laboratory values were available for CBC but not for lymphocyte subsets. Patients who received treatment for progression before day 30 were excluded. Infectious complications and diagnosis of MDS were reported from axi-cel infusion until last follow-up, progression, subsequent therapy or death. Cytokine release syndrome and neurotoxicity were prospectively graded according to Lee 2014 criteria and CTCTAE v4.03, respectively.(11) Response status was determined by 2007 revised response criteria for malignant lymphoma.(12)

Hematopoietic recovery and immune reconstitution assessment
To assess blood count and immune recovery, we performed complete blood count, lymphocyte subsets analysis by flow cytometry, and immunoglobulin G (IgG) levels at months 1, 3, 6, 9, 12, 15, 18 and 24, or until disease progression.

Statistical considerations

Categorical and continuous variables were evaluated using Chi square ($\chi^2$) or Fisher exact tests, or the Mann-Whitney test, as appropriate, to describe differences in baseline characteristics, response and toxicity between patients groups. Association between continuous variables were assessed using the bivariate Pearson correlation. A p-value of $\leq0.05$ (two-tailed) was considered statistically significant (IBM SPSS 23).
Results

Patient baseline characteristics

Grade 3-4 cytopenias were observed in 15 (48%) of 31 patients at day 30 (D30) after axi-cel infusion, and included neutropenia in 9 (29%) patients, anemia in 5 (16%) and thrombocytopenia in 13 (42%) patients.

On univariate analysis, baseline characteristics significantly associated with D30 grade 3-4 cytopenia were ECOG performance status 1 (p=0.03), >3 prior therapies (p=0.03), and low absolute lymphocyte count (ALC)(p=0.007) (Table 1).

Hematopoietic recovery and immune reconstitution

Fifteen patients included in this analysis had ongoing remission at 1 year and 10 at 2 years (1 did not have evaluable CBC). While these included both patients from ZUMA-1 and ZUMA-9, the 2 patients who received OOS product in cohort 2 of ZUMA-9 progressed before year 1 assessment. Among patients with ongoing remission, persistent grade 3-4 cytopenias was present in 4/15 (27%) patients at 1 year and in 1/9 (11%) patients at 2 years (Figure 1A-C). Among these patients, samples for flow cytometry assessment were available in 9 patients at 1 year, and 7 patients at 2 years. While recovery of CD8+ T cells (median 597 cells/µL, range 143-1492 cells/µL) and CD56+ NK cells (median 118 cells/µL, range 38-236 cells/µL) occurred in 9/9 (100%) patients by 1 year, reconstitution of CD4+ T cells was delayed, with normalization in 6/9 (67%) patients at 1 year (median 225 cells/µL, range 108-593 cells/µL) and 5/7 (71%) patients assessed at 2 years (median 263 cells/µL, range 83-1166 cells/µL); of interest, age of patients who did not experience CD4+ T cell normalization at 1 and 2 years ranged from 40 to 67 years.
IgG levels were normal in 7/13 (54%) patients at 1 year (median 646 mg/dL, range 200-1299 mg/dL) and 5/9 (56%) patients assessed at 2 years (median 552 mg/dL, range 366-1147 mg/dL) (Figure 1D-H). When limiting the analysis to patients who did not receive intravenous immunoglobulin G (IVIG) support, 4/4 had normalization of IgG levels at 1 year. Interestingly, a positive and significant association was observed between platelet count and CD56 cell count in 10 patients with available samples for flow cytometry assessment ($r=+0.64$, $p<0.001$) (Figure 1I).

Risk of myelodysplastic syndrome

When comparing the 15 patients with grade 3-4 D30 cytopenia to the 16 patients without grade 3-4 D30 cytopenia, the former received significantly higher number of platelet transfusions (11/15 [73%] vs 4/16 [25%], $p=0.01$), but there was no significant difference in red blood cell transfusions (13/15 [87%] vs 10/16 [63%], $p=0.22$). Four cases of MDS were diagnosed, after a median of 13.5 months (range, 4-26 months) from axi-cel infusion, all attributed to previous chemotherapies: median number of previous systemic therapies in these 4 patients was 5 (range 4-7), including autologous stem cell transplant in 1 patient. No significant difference in the incidence of MDS was observed between patients with and without grade 3-4 D30 cytopenia (3/15 [20%] vs 1/16 [6%], $p=0.33$).

Infectious complications

All 31 patients received granulocyte colony stimulating factor (GCSF) support for neutropenia; 13 (42%) received prophylactic IVIG and 4 (13%) therapeutic IVIG; 2 (6%) patients received
antibacterial prophylaxis, 13 (42%) Pneumocystis jiroveci pneumonia (PJP) prophylaxis, and 22 (71%) antiviral prophylaxis, while no patient received anti-fungal prophylaxis. Overall, among patients with ongoing response, 24 (77%) developed infectious complications with grade ≥3 in 13 (42%) (Figure 2). There was a trend for association between grade 3-4 D30 cytopenia and grade ≥3 infectious complications (9/15 [60%] vs 4/16 [25%], p=0.07), which was statistically significant when limiting the analysis to grade 3-4 D30 lymphopenia (9/14 [64%] vs 4/17 [24%], p=0.03). Due to small sample size, the association between D30 CD4 count and infections could not be assessed. Among all 31 patients, seventy-one infectious events of any grade were reported, with no isolated organism in 40 (56%) cases (Table 2). For these 71 infectious events, the most common etiology was viral (17/71 [24%]), followed by bacterial (10/71 [14%]) and fungal (4/71 [6%]), whereas in the remaining cases an organism could not be isolated. Among viral infections, herpes zoster skin infection was the most common event, reported in 5 patients (16%), none of whom were on antiviral prophylaxis. Median time from axi-cel infusion to onset of herpes zoster infection was 79 days (range, 49-723 days), and all cases resolved with valacyclovir therapy. A patient who was not on prophylaxis developed PJP 18 days after axi-cel infusion, and was successfully treated with sulfamethoxazole-trimethoprim.

Toxicity and efficacy

Among all 31 patients, 30 (97%) patients developed CRS of any grade, G3-4 in 3 (10%), and 20 (65%) patients neurotoxicity of any grade, G3-4 in 15 (48%). No significant difference in incidence of G3-4 CRS (13% vs 6%, p=0.60) and incidence of G3-4 neurotoxicity (53% vs 44%,
p=0.72) was observed when comparing patients who developed D30 G3-4 cytopenia to those who did not.

In order to manage either CRS or neurotoxicity, 12 (39%) patients required corticosteroids, and 21 (68%) required tocilizumab. No difference in use of corticosteroids (53% vs 25%, p=0.15) or use of tocilizumab (73% vs 63%, p=0.70) was observed when comparing patients who developed D30 G3-4 cytopenia to those who did not.

Overall, 26 (84%) patients achieved a response, and 19 (61%) a complete response (CR). No statistically significant difference in overall response rate (87% vs 81%, p=1) or CR rate (60% vs 63%, p=1) was observed when comparing patients with D30 G3-4 cytopenia to those without. No statistically significant difference in median baseline ALC was observed when comparing patients who achieved a response to those who did not (0.52 vs 0.34 \times 10^9/L; p=0.37), or patients who achieved a CR to those who did not (0.55 vs 0.39 \times 10^9/L; p=0.34).
Discussion

In summary, our results indicate that grade 3-4 cytopenia beyond D30 occur after CAR T-cell therapy in a subset of patients. While cytopenias resolve in most patients, it can persist in one third of evaluable patients at 1 year, highlighting the importance of long-term monitoring. The comparable incidence between our study and other reports suggests that the MDS was most likely due to prior therapies. Unfortunately, myeloid driver mutations were not tested in these patients before lymphodepleting chemotherapy, to further corroborate this hypothesis. While previous chemotherapies (and subsequent marrow reserve) and conditioning therapy may contribute to the prolonged myelosuppression, the observation of severe cytopenias in patients who have not received it suggests the intriguing hypothesis that they may instead signify intra-bone marrow CAR T-cell activity. To this regard, in our study grade 3-4 D30 cytopenia was more common in heavily pretreated patients with low baseline ALC, which may indicate a weaker recipient immune system due to prior therapy, and a potentially easier product engraftment. A trend for higher baseline ferritin levels in these patients also suggests a potential role for chronic inflammation in its etiology. Consistent with prior studies in patients with B-acute lymphoblastic leukemia (B-ALL) treated with anti-CD19 CAR T-cell therapy, we observed that viral infections, especially herpes zoster, were a later event, as compared to other infections. This is likely explained by the delayed recovery of CD4+ T cells, a well-established risk factor for the development of both common and opportunistic infections, which in this series occurred independently of age. These data suggest that effective long-term monitoring and prophylaxis against opportunistic infections needs to be considered in a subset of patients receiving CAR T-cell therapy. Although
the baseline characteristics of patients enrolled in the ZUMA-9 study mirrored those of patients treated in the real world setting, the subset of patients included in this analysis had an apparently higher incidence of grade ≥ 3 infections as compared to the 2-year follow-up of the ZUMA-1 trial (42% vs 28%). It is important to highlight that this may be due to differences in baseline characteristics and co-morbid health conditions of the subset of patients selected for this single institution analysis, as compared to the previously reported multi-center results. While pre-conditioning IgG levels were not available in this study, and prior treatment with anti-CD20 monoclonal antibodies and/or chemotherapy may have affected their levels before CAR T-cell infusion, hypogammaglobulinemia was observed in a subset of patients early after axi-cel therapy. Similar to what was reported with the use of tisagenlecleucel, recovery of IgG to normal levels was observed in more than half the patients in this series. In contrast to children where prophylactic IVIG is recommended in patients with IgG <400 mg/dL, in adults it is usually used in patients with hypogammaglobulinemia who also have frequent or severe infections. Consistent with this, we did not observe significant differences in infections between patients receiving prophylactic IVIG or not (p=0.10).

Finally, a strong and positive correlation between platelet count and CD56 cell count was observed in our study. NK cells are among the first hematopoietic cells that recover during bone marrow reconstitution based on data from stem cell transplant recipients. The association observed here between platelet count and NK cell numbers might reflect the dynamics of bone marrow recovery, although a mechanistic relation may still be possible and is under evaluation at our institution.
In conclusion, our study suggests that patients with prolonged cytopenias after CAR T-cell therapy may be managed conservatively with supportive care. Importantly, our results suggest the potential need for antimicrobial prophylaxis against opportunistic infections because of delayed CD4$^+$ T-cell recovery following axicabtagene ciloleucel therapy in a subset of patients.
Acknowledgments

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Authorship Contributions

PS designed the study, analyzed data, and wrote the paper; LN, JW, FH, NF, HJL, LEF, FS, SA, PK, VEM and EAH provided clinical care to patients and coauthored the paper; YC, SH, SA, AV, SA, and SJ collected clinical data and coauthored the paper; SN designed the study, analyzed the data, provided clinical care to patients, and wrote the paper.

Disclosure of Conflict of Interest

SSN reports honoraria and research support from Kite, a Gilead Company, Merck, Celgene, Allogene, and Unum Therapeutics; research support from Bristol-Myers Squibb, Poseida, Cellectis, Karus, and Acerta Pharma; and honoraria from Novartis, Pfizer, Precision Biosciences, Cell Medica, Calibr, Incyte, and Legend Biotech. FS reports honoraria from Celgene. LN reports honoraria from Celgene, Genentech, Gilead, Janssen, Juno, Novartis, Spectrum, TG Therapeutics and research support from Celgene, Genentech, Janssen, Karus Therapeutics, and Merck. NF reports honoraria from Celgene, Gilead Sciences, Pharmacyclics, Roche Pharma AG, research support from Celgene, Gilead Sciences, Pharmacyclics, and Roche Pharma AG.
References

Table 1. Baseline (day -5) characteristics by grade 3-4 cytopenia at day 30 after axi-cel infusion.

<table>
<thead>
<tr>
<th></th>
<th>Total (N=31)</th>
<th>Grade 3-4 cytopenia at 30 days (N=15)</th>
<th>No Grade 3-4 cytopenia at 30 days (N=16)</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>DLBCL</td>
<td>21 (68)</td>
<td>12 (80)</td>
<td>9 (56)</td>
<td>0.35</td>
</tr>
<tr>
<td>PMBCL</td>
<td>6 (19)</td>
<td>2 (13)</td>
<td>4 (25)</td>
<td></td>
</tr>
<tr>
<td>TFL</td>
<td>4 (13)</td>
<td>1 (7)</td>
<td>3 (19)</td>
<td></td>
</tr>
<tr>
<td>Age in years, median (range)</td>
<td>52 [23-76]</td>
<td>58 [23-76]</td>
<td>51 [28-75]</td>
<td>0.86</td>
</tr>
<tr>
<td>Male</td>
<td>23 (74)</td>
<td>9 (60)</td>
<td>14 (88)</td>
<td>0.11</td>
</tr>
<tr>
<td>Female</td>
<td>8 (26)</td>
<td>6 (40)</td>
<td>2 (12)</td>
<td></td>
</tr>
<tr>
<td>ECOG 0</td>
<td>19 (61)</td>
<td>6 (40)</td>
<td>13 (81)</td>
<td>0.03</td>
</tr>
<tr>
<td>ECOG 1</td>
<td>12 (39)</td>
<td>9 (60)</td>
<td>3 (19)</td>
<td></td>
</tr>
<tr>
<td>Ann Arbor Stage I-II</td>
<td>7 (23)</td>
<td>2 (13)</td>
<td>5 (48)</td>
<td>0.39</td>
</tr>
<tr>
<td>Ann Arbor Stage III-IV</td>
<td>24 (77)</td>
<td>13 (87)</td>
<td>11 (52)</td>
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<tr>
<td>No BM involvement</td>
<td>28 (90)</td>
<td>13 (87)</td>
<td>15 (94)</td>
<td>0.60</td>
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<tr>
<td>BM involvement</td>
<td>3 (10)</td>
<td>2 (13)</td>
<td>1 (6)</td>
<td></td>
</tr>
<tr>
<td>IPI score 0-2</td>
<td>18 (58)</td>
<td>6 (40)</td>
<td>12 (75)</td>
<td>0.07</td>
</tr>
<tr>
<td>IPI score 3-4</td>
<td>13 (42)</td>
<td>9 (60)</td>
<td>4 (25)</td>
<td></td>
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<tr>
<td>Previous therapies (n)</td>
<td>3 [1-11]</td>
<td>4 [2-8]</td>
<td>3 [1-11]</td>
<td>0.05</td>
</tr>
<tr>
<td>&gt; 3 previous therapies</td>
<td>14 (45)</td>
<td>10 (67)</td>
<td>4 (25)</td>
<td>0.03</td>
</tr>
<tr>
<td>Refractory disease</td>
<td>30 (97)</td>
<td>15 (100)</td>
<td>15 (94)</td>
<td>1</td>
</tr>
<tr>
<td>Previous ASCT</td>
<td>11 (35)</td>
<td>8 (53)</td>
<td>3 (19)</td>
<td>0.07</td>
</tr>
<tr>
<td>Cytopения at day -5</td>
<td>4 (13)</td>
<td>3 (20)</td>
<td>1 (6)</td>
<td>0.33</td>
</tr>
<tr>
<td>ANC (10⁹/L), median (range)</td>
<td>4 [0.8-24.8]</td>
<td>4 [1.1-24.9]</td>
<td>3.8 [0.8-12]</td>
<td>0.86</td>
</tr>
<tr>
<td>AMC (10⁹/L), median (range)</td>
<td>0.5 [0.1-1.6]</td>
<td>0.5 [0.1-1.2]</td>
<td>0.5 [0.3-1.6]</td>
<td>0.72</td>
</tr>
<tr>
<td>ALC (10⁹/L), median (range)</td>
<td>0.5 [0.2-3.6]</td>
<td>0.4 [0.2-3.6]</td>
<td>0.7 [0.2-1.6]</td>
<td><strong>0.007</strong></td>
</tr>
<tr>
<td>Hemoglobin (g/dL), median (range)</td>
<td>10.9 [7.4-14.2]</td>
<td>10.7 [7.4-12.6]</td>
<td>11.3 [9.3-14.2]</td>
<td>0.10</td>
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<tr>
<td>PLT count (10⁹/L), median (range)</td>
<td>157 [31-409]</td>
<td>105 [31-356]</td>
<td>184 [67-409]</td>
<td>0.21</td>
</tr>
<tr>
<td>Ferritin (ng/mL), median (range)</td>
<td>747 [209-9388]</td>
<td>2205 [230-9388]</td>
<td>450 [209-1554]</td>
<td>0.08</td>
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<tr>
<td>CRP (mg/L), median (range)</td>
<td>51 [2.7-352]</td>
<td>62.3 [2.7-352]</td>
<td>39.3 [3-268]</td>
<td>1</td>
</tr>
<tr>
<td>LDH (U/L), median (range)</td>
<td>703 [362-3426]</td>
<td>703 [362-3426]</td>
<td>681 [369-1667]</td>
<td>1</td>
</tr>
</tbody>
</table>

DLBCL, diffuse large B-cell lymphoma; PMBCL, primary mediastinal B-cell lymphoma; TFL, transformed follicular lymphoma; BM, bone marrow; IPI, international prognostic index; n, number; ASCT, autologous stem cell transplant; ANC, absolute neutrophil count; AMC, absolute monocyte count; ALC, absolute lymphocyte count; PLT, platelet; CRP, C-reactive protein; LDH, lactate dehydrogenase
Table 2. Infection types (all grades included)

<table>
<thead>
<tr>
<th>TOTAL (n= 71)</th>
<th>Number (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>NO ISOLATED ORGANISM (17 respiratory, 13 ENT, 3 skin, 4 GU, 2 GI, 1 sepsis)</td>
<td>40 (56)</td>
</tr>
<tr>
<td>VIRAL, total</td>
<td></td>
</tr>
<tr>
<td>Herpetic (5 VZV, 3 HSV, 2 CMV, 1 HHV6)</td>
<td>17 (24)</td>
</tr>
<tr>
<td>Respiratory</td>
<td>11 (15)</td>
</tr>
<tr>
<td>Enteric</td>
<td>3 (4)</td>
</tr>
<tr>
<td>Parvovirus</td>
<td>1 (1)</td>
</tr>
<tr>
<td>Hepatitis B</td>
<td>1 (1)</td>
</tr>
<tr>
<td>BACTERIAL, total</td>
<td>10 (14)</td>
</tr>
<tr>
<td>Gram negative (2 UTI, 2 GI, 1 bacteremia)</td>
<td>5 (7)</td>
</tr>
<tr>
<td>Gram positive (3 GI, 2 skin)</td>
<td>5 (7)</td>
</tr>
<tr>
<td>FUNGAL, total</td>
<td>4 (6)</td>
</tr>
<tr>
<td>Yeast (skin)</td>
<td>3 (4)</td>
</tr>
<tr>
<td>Pneumocystis (pneumonia)</td>
<td>1 (1)</td>
</tr>
</tbody>
</table>

ENT, ear nose throat; VZV, varicella zoster virus; CMV, cytomegalovirus; HSV, herpes simplex virus; HHV6, human herpes virus 6; UTI, urinary tract infection; GI, gastro-intestinal
Figure 1. Trends in hematopoietic and immune reconstitution after axi-cel therapy in patients with relapsed or refractory large B-cell lymphoma. A-C. Hematopoietic recovery (absolute neutrophil count, hemoglobin and platelet count; D-H. Reconstitution of ALC, CD4\(^+\), CD8\(^+\), CD56\(^+\) and immunoglobulin G up to 24 months after CART infusion. I. Correlation between CD56\(^+\) cell immune reconstitution and platelet recovery.

Red dotted line indicates lower limit of normal

Figure 2. Median time to infection (TTI)
Viral vs bacterial, $p=0.45$
Fungal vs bacterial, $p=0.11$

Proportion with infection vs Time from CART to infection (days)

- Viral: $N=17$, median TTI 79 days
- Bacterial: $N=10$, median TTI 25 days
- Fungal: $N=4$, median TTI 75 days