



Proposed diagnostic criteria for classical chronic myelomonocytic leukemia (CMM), CMM variants and pre-CMM conditions

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ABSTRACT

Chronic myelomonocytic leukemia (CMM) is a myeloid neoplasm characterized by dysplasia, abnormal production and accumulation of monocytic cells and an elevated risk of transforming into acute leukemia. Over the past two decades, our knowledge about the pathogenesis and molecular mechanisms in CMM has increased substantially. In parallel, better diagnostic criteria and therapeutic strategies have been developed. However, many questions remain regarding prognosis and optimal therapy. In addition, there is a need to define potential pre-phases of CMM and special CMM variants, and to separate these entities from each other and from conditions mimicking CMM. To address these unmet needs, an international consensus group met in a Working Conference in August 2018 and discussed open questions and issues around CMM, its variants, and pre-CMM conditions. The outcomes of this meeting are summarized herein and include diag-

Haematologica 2019
Volume 104(10):1935-1949

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Received: March 14, 2019.

Accepted: April 29, 2019.

Pre-published: May 2, 2019.

doi:10.3324/haematol.2019.222059

Check the online version for the most updated information on this article, online supplements, and information on authorship & disclosures: www.haematologica.org/content/104/10/1935

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nostic criteria and a proposed classification of pre-CMML conditions as well as refined minimal diagnostic criteria for classical CMML and special CMML variants, including oligomonocytic CMML and CMML associated with systemic mastocytosis. Moreover, we propose diagnostic standards and tools to distinguish between 'normal', pre-CMML and CMML entities. These criteria and standards should facilitate diagnostic and prognostic evaluations in daily practice and clinical studies in applied hematology.

Introduction

Chronic myelomonocytic leukemia (CMML) is a myeloid stem cell disease characterized by an abnormal production and accumulation of monocytic cells, often in association with other signs of myeloproliferation, substantial dysplasia in one or more hematopoietic cell lineages, and an increased risk of transformation into secondary acute myeloid leukemia (AML).^{1–5} As per definition, the Philadelphia chromosome and its related *BCR-ABL1* fusion gene are absent in CMML. Other disease-related drivers, such as the *JAK2* mutation V617F or the *KIT* mutation D816V, may be detected and may indicate a special variant of CMML, such as CMML associated with systemic mastocytosis (SM-CMML).^{6–8} However, most somatic mutations identified in CMML patients, such as mutations in *SRSF2*, *TET2*, or *RAS*, are not disease-specific, but are also detected in myelodysplastic syndromes (MDS), myeloproliferative neoplasms (MPN), or AML.^{8–11}

For many years, CMML was listed as a separate variant among the MDS in the classification of the French-American-British (FAB) working group.^{2,12} However, in 2001, the World Health Organization (WHO) reclassified CMML into a newly created MDS/MPN overlap group, defined by the presence of both MDS-related and MPN-related morphological and clinical features.¹³ Depending on the leukocyte count, CMML can be divided into a 'dysplastic' variant (leukocyte count $\leq 13 \times 10^9/L$) and a 'proliferative' variant (leukocyte count $> 13 \times 10^9/L$).² In 2001 and 2008, the WHO also proposed a split into CMML-1 and CMML-2, based on the percentage of blast cells in the blood and bone marrow (BM).^{13,14} In the most recent updates of the WHO 2016 classification, CMML is again listed amongst the MDS/MPN overlap disorders.^{15,16} Based on the percentage of blasts, CMML is now divided into CMML-0, CMML-1, and CMML-2.^{15–19} Moreover, contrasting the 2008 WHO classification, the diagnosis of CMML now requires both an absolute moncytosis ($\geq 1 \times 10^9/L$) and relative moncytosis ($\geq 10\%$ of leukocytes) in the peripheral blood (PB).^{15–19} In the 2008 and 2016 update of the WHO classification, CMML can only be diagnosed per definition when rearrangements in *PDGFRA*, *PDGFRB* or *FGFR1* genes have been excluded, and in the 2016 update, the *PCM1-JAK2* fusion gene was added as an excluding criterion.^{14–16,19} These molecular aberrations are commonly found in eosinophilia-associated neoplasms such as chronic eosinophilic leukemia.^{20,21} However, CMML is also listed as an underlying variant in these molecular 'entities' in the WHO classification system.^{20,21}

Over the past two decades, our knowledge about the molecular features and mechanisms in CMML has increased substantially.^{4–11,22–26} Moreover, new diagnostic criteria, prognostic markers, and therapeutic concepts have been developed.^{26–29} Nevertheless, a number of ques-

tions remain concerning basic diagnostic standards, prognostication, optimal management and therapeutic options. Furthermore, there is a need to define clinically relevant pre-phases of CMML and distinct CMML variants by clinical variables, histomorphological features, flow cytometric phenotypes, molecular markers and cytogenetic findings. It is also important to separate CMML and pre-CMML conditions from diverse mimickers. To address these unmet needs, an international consensus group discussed open questions and issues around CMML, its variants and pre-CMML entities in a Working Conference held in August 2018. The outcomes of this meeting are summarized in this article and include proposed diagnostic criteria and a classification of pre-CMML conditions as well as updated minimal diagnostic criteria for CMML and its variants. In addition, diagnostic standards and diagnostic algorithms are proposed. Details concerning the conference format, pre- and post-conference discussion and consensus-finding are described in the *Online Supplement*.

Definition of CMML and minimal diagnostic criteria

The diagnostic criteria of CMML, as defined by the WHO,^{15,16} are depicted in *Online Supplementary Table S1*. Our faculty is of the opinion that these criteria are valid in general for the classical form of CMML, but need adjustments for special variants of CMML. Based on consensus discussion, the following concept is proposed.

The classical form of CMML is defined by the following pre-requisite criteria: (i) persistent (at least 3 months) absolute PB moncytosis ($\geq 1 \times 10^9/L$) and relative moncytosis ($\geq 10\%$ of PB leukocytes); (ii) exclusion of *BCR-ABL1*⁺ leukemia, classical MPN and all other hematologic neoplasms that may serve as a primary source of moncytosis; and (iii) a blast cell count of 0–19% in PB and/or BM smears and exclusion of all (other) histopathological, morphological, phenotypic, molecular and cytogenetic signs that qualify as evidence of AML. In addition, morphological and/or histopathological evidence for diagnostic dysplasia in one or more of the three major BM cell lineages ($\geq 10\%$ of megakaryocytes and/or erythroid precursor cells and/or neutrophilic cells) must be present. If dysplasia is absent or not diagnostic (<10%), the presence of cytogenetic or molecular lesions (mutations) typically found in CMML and/or the presence of CMML-related flow cytometry abnormalities may be employed as co-criteria and may lead to the diagnosis of CMML, provided that the pre-requisite criteria listed above are fulfilled. Pre-requisite criteria and co-criteria of the classical form of CMML are presented in Table 1.

The exclusion of various reactive states producing moncytosis (and sometimes even dysplasia) was also discussed and regarded as being of great importance. However, these mimickers cannot *a priori* exclude the presence of a concomitant CMML, but may indeed occur

Table 1. Minimal diagnostic criteria for classical chronic myelomonocytic leukemia.***A. Prerequisite criteria (all must be fulfilled)**

- Persistent (3 months) peripheral blood moncytosis $\geq 1 \times 10^9/L$ and relative moncytosis of $\geq 10\%$ of circulating peripheral blood leukocytes
- Exclusion of *BCR-ABL1*⁺ leukemia, classical MPN and all other bone marrow neoplasms that could serve as a primary source of chronic persistent moncytosis
- Blast cell count of $<20\%$ in peripheral blood and bone marrow smears and exclusion of all other histopathological, morphological, molecular and cytogenetic features that count as evidence of the presence of acute myeloid leukemia**

B. Morphological criterion = Dysplasia

- Dysplasia in at least 10% of all cells in one of the following lineages in the bone marrow smear: erythroid; neutrophilic; megakaryocytic

C. Co-criteria (for patients fulfilling A but not B, and otherwise showing typical clinical features of CMML such as splenomegaly)

- Typical chromosome abnormalities by conventional karyotyping or FISH***
- Abnormal findings in histological and/or immunohistochemical studies of bone marrow biopsy sections supporting the diagnosis of CMML****
- Abnormal immunophenotype of bone marrow and blood cells by flow cytometry, with multiple CMML-associated phenotypic aberrancies indicating the presence of an abnormal/dysplastic population of monocytic and other myeloid cells*****
- Evidence of a clonal population of myeloid cells determined by molecular (sequencing) studies revealing CMML-related mutations*****

*The diagnosis of classical CMML can be established when all prerequisite criteria (A) and either morphological dysplasia (B) or one or more of the co-criteria (C) are fulfilled.

Examples: Auer rods, overt acute myeloid leukemia (AML) by histology and immunohistochemistry; presence of AML-specific diagnostic cytogenetic and/or molecular markers (e.g., inv16). *Typical cytogenetic abnormalities found in CMML (*Online Supplementary Table S6*). ****Leukemic infiltration of CD14+ monocytes and exclusion of AML.

*****Utilizing a cutoff value of $>94\%$ MO1 monocytes, phenotyping can identify CMML cases with a sensitivity of $>90\%$ and a specificity of $>95\%$, and a decrease in MO3 monocytes is even as diagnostic as an increase in circulating MO1 cells.^{127,129,131} *****Genes that are often mutated in the context of CMML/MDS include, among others, *TET2*, *SRSF2*, *ASXL1* and *SETBP1*. The minimal allele burden proposed to count as co-criterion: $\geq 10\%$. CMML: chronic myelomonocytic leukemia; MPN: myeloproliferative neoplasm(s); MDS: myelodysplastic syndrome(s); FISH: fluorescence *in situ* hybridization.

in CMML patients in the context of certain infections. Furthermore, most of these mimickers do not produce persistent moncytosis. Proof of clonality by molecular and cytogenetic studies, and other disease-specific parameters, together with global and specific laboratory (e.g., microbial screen) tests should easily lead to the conclusion that the patient is suffering from reactive moncytosis but not from (or also from) CMML.

The *a priori* exclusion of AML as a criterion should apply to both the classical and the special variants of CMML, whereas the *a priori* exclusion of other indolent hematopoietic neoplasms should only apply to the classical variant of CMML and oligomonocytic CMML but not to other special CMML variants. This is because several previous and more recent studies have shown that CMML may be accompanied by (or may accompany) other myeloid or lymphoid neoplasms, such as systemic mastocytosis. In several of these patients, the CMML clone is dominant and the additional sub-clone is smaller in size and usually not relevant clinically, even if these smaller clones express certain driver mutations, such as *KIT* D816V or a rearranged *PDGFRA* or *PDGFRB*. Rarely, a Philadelphia chromosome-positive chronic myeloid leukemia may develop as an additional small-sized (sub)clone in a patient with CMML. Our faculty is of the opinion that the presence of additional (chronic) myeloid, mast cell, or lymphoid neoplasms does not exclude a diagnosis of CMML, provided that diagnostic WHO criteria for CMML are fulfilled. Moreover, these concomitant neoplasms should not exclude a diagnosis of CMML even when the driver of the concomitant disease (e.g., *KIT* D816V) is detectable in CMML monocytes. Thus, whereas the occurrence of AML is always regarded as transformation of CMML, the occurrence of *indolent* myeloid, mast cell, or lymphoid neoplasms should be regarded as concomitant disorders. Co-existing myeloid neoplasms

and CMML may be derived from the same original founder clone.

There are also patients in whom a certain driver of another BM neoplasm is present, such as a mutated *JAK2*, *PDGFRA/B*, or *FGFR1*, but only the diagnostic criteria for CMML (not those of the other BM neoplasm) are fulfilled. Our faculty concludes that these cases should also be regarded and diagnosed as special variants of CMML. This strategy is in line with the current WHO classification. In fact, whereas the primary molecular diagnosis is often based on a mutated form of *JAK2*, *PDGFRA/B* or other classical driver, the underlying or additional diagnosis may well be CMML.^{20,21}

Grading of CMML

The grading system of CMML proposed by the WHO is regarded as standard in clinical hematology. Our faculty recommends the use of this grading system as the initial prognostic tool in classical CMML. In fact, classical CMML should be split into CMML-0, CMML-1 and CMML-2 based on the blast cell count (*Online Supplementary Table S2*).¹⁵⁻¹⁹ In addition, CMML can be divided into a dysplastic variant and a proliferative variant based on leukocyte counts (threshold: $13 \times 10^9/L$) (*Online Supplementary Table S2*). The resulting grading system defines six distinct CMML variants with variable clinical outcome.¹⁷ However, grading may sometimes be challenging. For example, blast cell counts obtained from BM smears may differ from those obtained in the PB so that the grade is questionable. Our faculty recommends that in patients in whom results from BM and PB smears would not fit into one distinct grade of CMML (e.g., BM blasts 4% and PB blasts 6%) grading should be based on the higher blast cell percentage (*Online Supplementary Table S2*). It is worth noting that initial prognostication by grading does not include all essential prognostic parameters.

Table 2. Overview of special variants of chronic myelomonocytic leukemia.

Special variant	Key diagnostic features that discriminate the variant from classical CMMML
Oligomonocytic CMMML	Absolute PB monocyte count <1x10 ⁹ /L
SM with concomitant CMMML = SM-CMMML	WHO criteria for SM fulfilled; in most patients CMMML monocytes exhibit KIT D816V
CMMML with a concomitant myeloid neoplasm* expressing a classical MPN-driver, such as JAK2 V617F, BCR-ABL1 or rearranged PDGFRA/B*** or FGFR1.	WHO criteria for a classical MPN, such as CML**, PMF, or a myeloid neoplasm with rearranged PDGFRA/B*** are fulfilled in addition to the criteria for CMMML.
CMMML with expression of a molecular MPN-driver – examples: CMMML with JAK2 V617F or CMMML with a rearranged PDGFRA/B or CMMML with rearranged FGFR1.	Molecular drivers of classical MPN, such as JAK2 V617F**** or rearranged PDGFRA/B*** are found but diagnostic criteria for such classical MPN are not fulfilled (only criteria for CMMML are met)
CMMML with a concomitant lymphoid/lymphoproliferative neoplasm	WHO criteria for a lymphoid neoplasm are fulfilled

*These conditions must be separated from MPN with concomitant monocytosis that do not fulfil the diagnostic criteria for CMMML. **Unlike in SM-CMMML, in which monocytes display KIT D816V or CMMML with rearranged PDGFRA, the CMMML monocytes must not express BCR-ABL1 in patients with CML plus CMMML. ***Several different translocations and fusion genes involving PDGFRA or PDGFRB may be detected, such as the t(5;12) associated with the TEL-PDGFRB fusion gene. ****JAK2 V617F itself counts as a feature of MPN; therefore, detection of JAK2 V617F can confirm the diagnosis of CMMML (as MPN/MDS overlap disease) when other signs of myeloproliferation are absent (e.g., no splenomegaly and no leukocytosis). CMMML: chronic myelomonocytic leukemia; PB: peripheral blood; SM: systemic mastocytosis; WHO: World Health Organization; MPN: myeloproliferative neoplasm; CML: chronic myeloid leukemia; PMF: primary myelofibrosis.

We therefore recommend that in each case, deeper (full) prognostication should follow using multiparametric scoring systems (see later). It should be noted, however, that grading of CMMML has only been validated in the classical form of CMMML, not in special CMMML variants. Therefore, although grading is also recommended for special CMMML entities, it is not regarded standard and the result must be interpreted with caution in these patients.

Special variants of CMMML: overview

As mentioned before, the classical form of CMMML meets all pre-requisite criteria, and no signs (including molecular features) of an additional, concomitant BM neoplasm are detected. The special variants of CMMML form a heterogeneous group of neoplasms comprising distinct clinical and biological entities. In one group of patients, the relative monocyte count ($\geq 10\%$) is fulfilled without resulting in an absolute count $\geq 1 \times 10^9/L$, precluding the diagnosis of ‘classical CMMML’. Most of these patients are diagnosed as having MDS or MPN/MDS-unclassified by WHO criteria. In another group of patients, a molecular signature suggestive of a different type of myeloid neoplasm is detected but only the criteria for CMMML (not those for the other neoplasm) are met. Such an example is CMMML with JAK2 V617F (without definitive evidence of a concomitant MPN). In a third group, CMMML co-exists with another BM neoplasm, such as MPN or mastocytosis. In these patients, additional blood count abnormalities (e.g., eosinophilia), an elevated serum tryptase level and/or BM fibrosis, may be detected.

All variants of CMMML (classical and special) can occur as a primary CMMML or as a secondary CMMML following a ‘mutagenic’ event, such as chemotherapy (therapy-related CMMML). In addition, our faculty is of the opinion, that the term secondary CMMML may also be appropriate for those patients who develop CMMML (months or years) after another indolent myeloid neoplasm, such as a MDS or systemic (indolent or aggressive) mastocytosis, had been diagnosed. In the following paragraphs, the clinical features and diagnostic criteria of special (atypical) variants of CMMML are proposed and discussed. An overview of the special variants of CMMML is provided in Table 2.

Oligomonocytic CMMML

Over the past few years, more and more cases of cytopenic patients exhibiting relative monocytosis ($\geq 10\%$) and moderately increased absolute blood monocytes not reaching the required threshold to diagnose classical CMMML ($1.0 \times 10^9/L$) have been described. These cases have recently been referred to as oligomonocytic CMMML.³⁰ According to the WHO classification most of these patients would be classified as having MDS (with monocytosis) or perhaps MPN/MDS-unclassifiable. However, most of these patients exhibit typical features of CMMML, including a typical morphology of PB and BM cells, splenomegaly, and CMMML-related molecular features (e.g. mutations in TET2 and SRSF2).³⁰⁻³² Some of these patients have prominent BM monocytosis without diagnostic PB monocytosis at diagnosis.^{30,32}

Whereas several of these cases remain stable without progression, the majority will develop ‘overt’ CMMML or, eventually, secondary AML during follow-up. Therefore, oligomonocytic CMMML may also be regarded as a potential pre-phase of classical CMMML. Our faculty is of the opinion that the term oligomonocytic CMMML should be used in clinical practice. Diagnostic pre-requisite criteria for oligomonocytic CMMML are: (i) persistent (lasting at least 3 months) absolute peripheral monocytosis of $0.5-0.9 \times 10^9/L$ and relative blood monocytosis ($\geq 10\%$ of blood leukocytes); (ii) exclusion of BCR-ABL1⁺ leukemia, classical MPN and all other myeloid neoplasms that can explain monocytosis; and (iii) a blast cell count of 0-19% in PB and/or BM smears and exclusion of all histopathological, morphological, phenotypic, molecular and cytogenetic signs that count as proof of AML. Diagnostic dysplasia in one or more of the three major BM lineages ($\geq 10\%$) must also be documented. If dysplasia is lacking or ‘sub-diagnostic’ (<10%), the presence of cytogenetic or molecular lesions (mutations) typically found in CMMML and/or the presence of CMMML-related flow cytometry abnormalities, may also lead to the conclusion that the patient has oligomonocytic CMMML provided that the other diagnostic criteria described above are fulfilled and all other myeloid neoplasms have been excluded. The proposed criteria for oligomonocytic CMMML are listed in

Table 3. Proposed minimal diagnostic criteria for oligomonocytic chronic myelomonocytic leukemia.***A. Prerequisite criteria (all must be fulfilled)**

- Persistent (3 months) peripheral blood moncytosis $0.5\text{--}0.9 \times 10^9/\text{L}$ and relative moncytosis of $\geq 10\%$ of circulating peripheral blood leukocytes
- Exclusion of *BCR-ABL1*⁺ leukemia, classical MPN and all other bone marrow neoplasms that could serve as a primary source of chronic persistent moncytosis
- Blast cell count $<20\%$ in peripheral blood and bone marrow smears and exclusion of all other histopathological, morphological, molecular and cytogenetic features that count as evidence of the presence of acute myeloid leukemia**

B. Morphological criterion = Dysplasia

Dysplasia in at least 10% of all cells in one of the following lineages in the bone marrow smear: erythroid; neutrophilic; megakaryocytic

C. Co-criteria (for patients fulfilling A but not B, and otherwise showing typical clinical features of CMML such as splenomegaly)

- Typical chromosome abnormalities by conventional karyotyping or FISH***
- Abnormal findings in histological and/or immunohistochemical studies of bone marrow biopsy sections supporting the diagnosis of CMML****
- Abnormal immunophenotype of bone marrow and blood cells by flow cytometry, with multiple CMML-associated phenotypic aberrancies indicating the presence of an abnormal/dysplastic population of monocytic (and other myeloid) cells*****
- Evidence of a clonal population of myeloid cells determined by molecular (sequencing) studies revealing CMML-related mutations*****

*The diagnosis of classical CMML can be established when all prerequisite criteria (A) and either morphological dysplasia (B) or one or more of the co-criteria (C) are fulfilled.

Examples: Auer rods, overt acute myeloid leukemia (AML) by histology and immunohistochemistry; presence of AML-specific diagnostic cytogenetic and/or molecular markers (e.g., inv16). *Typical cytogenetic abnormalities found in CMML (*Online Supplementary Table S6*). ****Leukemic infiltration of CD14+ monocytes and exclusion of AML.

*****Utilizing a cutoff value of $>94\%$ MO1 monocytes, phenotyping can identify CMML cases with a sensitivity of $>90\%$ and a specificity of $>95\%$, and a decrease in MO3 monocytes is even as diagnostic as an increase in circulating MO1 cells.^{127,129,131} *****Genes that are often mutated in the CMML/MDS context include, among other, *TET2*, *SRSF2*, *ASXL1* and *SETBP1*. Minimal allele burden proposed to count as a co-criterion: $\geq 10\%$. CMML: chronic myelomonocytic leukemia; MPN: myeloproliferative neoplasm(s); MDS: myelodysplastic syndrome(s); FISH: fluorescence *in situ* hybridization.

Table 3. Patients with oligomonocytic CMML should be managed and followed clinically in the same way as patients with classical CMML.

CMML associated with *KIT D816V*⁺ systemic mastocytosis

According to WHO criteria, systemic mastocytosis (SM) can be divided into: (i) indolent SM (ISM), which is associated with a normal life expectancy; (ii) smoldering SM (SSM), in which signs of BM dysplasia, myeloproliferation and/or splenomegaly are found but survival and prognosis are still favorable; and (iii) advanced SM, defined by a poor prognosis.^{33–36} Advanced SM is further divided into aggressive SM (ASM), SM with an associated hematologic neoplasm (SM-AHN) and mast cell leukemia (MCL).^{33–36} The most frequent AHN detected in patients with SM-AHN is CMMI.^{6–8,36} In these patients the SM component of the diseases may present as ISM, ASM or, rarely, as MCL. Our faculty concludes that diagnostic WHO criteria for SM and diagnostic criteria for classical CMML (except exclusion of SM) must be fulfilled to diagnose SM-CMML.

Patients with SM may present with moncytosis resembling oligomonocytic CMML. However, the clinical features of SSM and advanced SM overlap largely with those found in patients with oligomonocytic CMML. Especially in SSM, myeloproliferation, dysplasia and splenomegaly are diagnostic criteria.^{33–35} Therefore, our faculty is of the opinion that such patients should be classified as having ISM, SSM or ASM with moncytosis rather than SM with oligomonocytic CMML.

In patients with CMML, a concomitant SM is often overlooked, especially when the disease does not present with cutaneous lesions. In other patients, CMML is diag-

nosed long before SM is detected by chance or after *KIT D816V* is identified: even though it is tempting to call these conditions CMML-SM, our faculty agreed that the classical terminology should be SM-CMML which is also in line with the WHO classification^{34,35} and that the subtype of SM and of CMML should be defined in the final diagnosis (e.g., ISM-CMML-1 or ASM-CMML-2) with recognition that in the SM-context, CMML is always regarded as a secondary neoplasm.^{6,36} Furthermore our faculty is of the opinion that it should be standard practice to examine BM and blood leukocytes for the presence of *KIT D816V* in all patients with (suspected) CMML. In almost all patients with SM-CMML, neoplastic monocytes display *KIT D816V*.⁷ In these monocytes, mutated *KIT* is not expressed on the cell surface but acts as a cytoplasmic driver. In line with this hypothesis drugs targeting *KIT D816V* can sometimes induce a major decrease in monocyte counts in patients with ASM-CMML.³⁷

Therapy of SM-CMML should be based on a bi-directional strategy: in fact the SM component of the disease should be treated as if no CMML was diagnosed and CMML should be treated as if no SM had been found, with recognition of drug-drug interactions and the possibility of drug-induced anaphylaxis.^{33–35} In many cases (ISM-CMML) the SM component of the disease is only treated symptomatically.^{33–35}

CMML associated with mutated *JAK2*, rearranged *PDGFRA/B* or other drivers

Patients with CMML may present with the *JAK2* mutation V617F, a rearranged *PDGFRA* or *PDGFRB*, often in the context of hypereosinophilia, or other drivers related to distinct hematopoietic neoplasms as defined by the WHO.^{5,9–11,38–43}

CMM^L with rearranged *PDGFRA*, *PDGFRB*, *FGFR1* or *PCM1-JAK2*

In these patients, persistent substantial monocytosis ($\geq 1.0 \times 10^9/L$) is detected and all other consensus criteria for classical CMM^L (see previous paragraphs) are also met, except the following specific exclusion criteria: CMM^L to be excluded in the presence of a well-characterized diagnosis of myeloid/lymphoid neoplasm with rearranged *PDGFRA*, *PDGFRB*, *FGFR1* or *PCM1-JAK2* (Table 2). Except for neglecting the above-mentioned criteria, our proposal is otherwise fully in agreement with all of the other tenets postulated by the WHO classification.^{14,15} In relation to neoplasms with rearranged *PDGFRA/B*, *FGFR1* or *PCM1-JAK2*, the WHO's definition of 'myeloid/lymphoid neoplasms' is too generic and there is a clinical need to know whether the underlying myeloid neoplasm is an aggressive disease, like AML, or a chronic neoplasm such as CMM^L or chronic eosinophilic leukemia.^{20,21} Our faculty is of the opinion that (unlike in previous times) the presence of one criterion-confirmed myeloid neoplasm should not *a priori* exclude the presence of another (second concomitant) myeloid or lymphoid neoplasm. Hence, when CMM^L is encountered in the context of another molecularly defined myeloid/lymphoid neoplasm (as a final diagnosis), it should be delineated as a specific subtype of the myeloid/lymphoid neoplasm with eosinophilia along with the specific associated gene rearrangement (*PDGFRA/B* or *FGFR1* or *PCM1-JAK2*).

CMM^L with *JAK2 V617F*

In these patients the situation is different. First, *JAK2 V617F* itself may be considered as a criterion of myeloproliferation in MDS/MPN, e.g. in cases with MDS/MPN with ring sideroblasts and thrombocytosis. In the context of CMM^L, the *JAK2* mutation is also typically associated with other signs of myeloproliferation (including BM fibrosis) and with the 'myeloproliferative variant' of CMM^L.^{39,42,43} Therefore, our faculty concludes that *JAK2 V617F* should also count as a molecular co-criterion of MDS/MPN and thus for CMM^L. Second, the presence of a *JAK2*-mutated MPN does not exclude the presence of a concomitant CMM^L if diagnostic criteria for both neoplasms are fulfilled. If this is not the case because the size of the MPN-like clone carrying *JAK2 V617F* is too small and/or other MPN features are clearly missing, the final diagnosis will be CMM^L with *JAK2 V617F*. On the other hand, in patients in whom the *JAK2* allelic burden is high and clinical and laboratory features argue for an overt MPN rather than CMM^L (e.g., polycythemia and/or BM fibrosis without dysplasia and without molecular or flow cytometry-based signs of CMM^L) the final diagnosis will be *JAK2 V617F*⁺ MPN with monocytosis.⁴³ In a third group of patients, diagnostic criteria for both a distinct MPN and CMM^L are fulfilled and the mutation status confirms the presence of an overt *JAK2*-mutated MPN (usually with high allelic burden). These patients are suffering from both MPN and CMM^L or from a gray zone disease displaying hybrid features between MPN and CMM^L.^{44,45} Our faculty concludes that it is therefore important to measure the *JAK2 V617F* allele burden in all patients with CMM^L.^{39,42,43} Other drivers, such as *BCR-ABL1*, are rarely found in patients with CMM^L. However, although in classical CMM^L, the presence of *BCR-ABL1* must be excluded, it may be detected in rare patients, suggesting the existence of a special variant of CMM^L (defined by a co-exist-

ing chronic myeloid leukemia). In some of these cases, the chronic myeloid leukemia clone may be small. In other patients, however, the chronic myeloid leukemia may even mask the CMM^L at the initial diagnosis.⁴⁶

The management and therapy of patients with special variants of CMM^L depend on the subtype of the disease and the molecular driver involved, e.g., *FIP1L1/PDGFRA*, other gene abnormalities involving *PDGFRA* or *PDGFRB*, *KIT D816V* or *JAK2 V617F*. Therefore, it is of crucial importance to screen for all these drivers in all patients with CMM^L. The type of therapy to consider in these patients depends on clinical features, the histopathological diagnosis, the size of the mutated clone(s) and the type of driver. The type of driver is of considerable importance since novel treatments directed against these drivers, are often extremely effective.⁴⁷⁻⁵⁰ For example, imatinib can induce long-lasting molecular and hematologic complete remissions in patients with *FIP1L1/PDGFRA*-rearranged myeloid neoplasms with features of CMM^L or MPN.⁴⁷⁻⁴⁹ Even in patients who develop CMM^L and secondary AML in the context of *FIP1L1/PDGFRA*, the disease may respond to imatinib.⁵⁰ It is, therefore, important to diagnose all patients based on molecular markers and to define the major drivers and therapeutic targets expressed by malignant cells in order to provide optimal management and therapy.

CMM^L associated with lymphoid neoplasms

In a small group of patients with CMM^L, a co-existing lymphoproliferative neoplasm is diagnosed, such as a lymphocytic leukemia, non-Hodgkin lymphoma or multiple myeloma.⁵¹⁻⁶⁰ In most patients, the lymphoid neoplasm is detected first, and CMM^L is considered to develop as treatment-induced, secondary, leukemia.^{51,57} In other patients, CMM^L is diagnosed first and later a lymphoid neoplasm is detected during follow-up.⁵²⁻⁵⁶ It is worth noting that in patients with CMM^L, polyclonal hypergammaglobulinemia is often recorded: this must be distinguished from the monoclonal gammopathy of concomitant myeloma, monoclonal gammopathies of undetermined significance and both low-count and high-count monoclonal B lymphocytoses which represent pre-malignant conditions.

The management and treatment of lymphoid neoplasms presenting with concomitant (secondary) CMM^L is a clinical challenge. In non-transplantable cases, both diseases require separate treatment plans. Because of the high risk of transformation to AML, allogeneic hematopoietic stem cell transplantation should be considered in young, fit patients, especially when it can be expected that the lymphoid neoplasm will also be eradicated by this approach.

Treatment-related CMM^L and other secondary forms of CMM^L

Our faculty concluded that both the classical form of CMM^L and the special variants of CMM^L should be divided into primary (*de novo*) CMM^L and secondary CMM^L. The latter group includes patients who (i) received chemotherapy and/or radiation therapy in the past (therapy-related CMM^L) or (ii) have a history of a preceding MDS, MPN or another indolent myeloid or mast cell neoplasm prior to their diagnosis of CMM^L.^{51,57,58,61-64} Recent data suggest that patients with therapy-related secondary CMM^L may have shorter overall survival compared to that of patients with primary (*de*

novo) CMML.⁶⁵ Although progression-free survival may not be different in these patients compared to those with *de novo* CMML, some of these patients progress rapidly to secondary AML. It is also worth noting that patients with therapy-related secondary CMML have a higher frequency of karyotypic abnormalities compared to patients with *de novo* CMML.⁶⁶ Eligible patients in this group should be offered allogeneic hematopoietic stem cell transplantation.

Potential pre-phases of CMML

During the past few years evidence has accumulated suggesting that hematopoietic neoplasms, including MDS, MPN and MDS/MPN, develop in a step-wise manner. In the earliest phases of clonal development, patients present without overt signs or symptoms of a hematopoietic neoplasm but their leukocytes carry one or more somatic mutations, usually (early, passenger-type) mutations otherwise also found in overt myeloid neoplasms (for example *TET2* mutations).⁶⁷⁻⁷⁰ In the context of MDS and other myeloid neoplasms, these cases have been referred to as clonal hematopoiesis of indeterminate potential (CHIP), or, when accompanied by cytopenia, as clonal cytopenia of unknown significance (CCUS).⁶⁹⁻⁷³ Since these mutations are frequently detected in older individuals, the condition is also called age-related clonal hematopoiesis (ARCH).^{70,73} In a few healthy individuals, *bona fide* oncogenic drivers (such as *BCR-ABL1*) are detected in a small subset of leukocytes. Because of the oncogenic potential of these drivers, these conditions are termed clonal hematopoiesis with oncogenic potential (CHOP).^{71,73} CHIP, CCUS and CHOP may also be the earliest clonal conditions preceding CMML. For these cases, the definitions recently proposed for CHIP, CCUS and CHOP should apply.^{69,71,73}

Apart from somatic mutations, other factors, such as epigenetic modifications, chronic inflammation or aging-related processes, may also trigger the selection and expansion of pre-malignant neoplastic clones in myeloid neoplasms including CMML.⁷⁴⁻⁷⁶ Some of these conditions may present with persistent monocytosis without signs of an overt myeloid neoplasm and may represent pre-phases of overt CMML. In other patients, however, no or another hematopoietic neoplasm develops during follow-up. Therefore, our faculty concluded that this pre-phase should be termed idiopathic monocytosis of unknown significance, provided that the following criteria are met: (i) persistent (at least 3 months) relative ($\geq 10\%$) and absolute ($>0.5 \times 10^9/L$) monocytosis; (ii) no diagnostic dysplasia and no signs of myeloproliferation; (iii) no signs and criteria of a myeloid or other hematopoietic neoplasm fulfilled; (iv) no flow cytometric abnormalities or somatic mutations related to a myeloid, mast cell or lymphoid neoplasm detected in leukocytes; and (v) no reactive condition that would explain reactive monocytosis is detected (Table 4 and *Online Supplementary Table S3*). If CHIP-like mutations are found in such patients, but no hematopoietic neoplasm can be diagnosed using the WHO criteria, the final diagnosis changes to clonal monocytosis of unknown significance (*Online Supplementary Table S3*). It is also worth noting that idiopathic cytopenias of unknown significance can precede CMML.^{64,77-79} Especially in patients with idiopathic thrombocytopenia of unknown significance, a CMML may be detected upon deeper investigations or during follow-up.⁷⁷⁻⁷⁹ Finally, as mentioned before, oligomonocytic CMML, although proposed as a special variant of CMML, must also be regarded as a potential pre-phase of classical CMML. In this regard it is important to note that these patients should have a regular follow-up

Table 4. Overview of non-clonal and clonal conditions that may precede chronic myelomonocytic leukemia.

Feature	IMUS	ICUS	Pre-CMML conditions and comparison to classical CMML	CMUS	O-CMML	CMML	
			CCUS	CHIP/CHOP			
Absolute moncytosis ($\geq 0.5 \times 10^9/L$)	+	+/-	+/-	+/-	+	+	+
Substantial moncytosis ($\geq 1 \times 10^9/L$)	+/-	-	-	-	+/-	-	+
Relative moncytosis (>10% of leukocytes)	+	-	-	-	+	+	+
Dysplasia*	-	-	-	-	-	+	+
Cytopenia(s)**	-	+	+	-	-	+/-	+/-
BM blasts	<5%	<5%	<5%	<5%	<5%	<20%	<20%
Flow abnormalities	-	-	+/-	+/-	-	++	++
Cytogenetic abnormality (≥ 1)	-***	-***	+/-	+/-	-***	++	++
Molecular aberration/s****	-	-	+	+	++++	++	++

*At least 10% of all cells in a given lineage (erythroid, neutrophilic, or megakaryocytic) are dysplastic. **Persistent cytopenia(s) recorded over a time-period of at least 4 months.

In a subset of cases, a small clone is detectable by FISH. *A molecular aberration is defined by CMML/MDS-related mutations and an allele burden of $\geq 2\%$. The working definition for pre-CMML conditions is also $\geq 2\%$ allele burden, whereas the minimal allele burden to count as a co-criterion for CMML is 10%. In most patients with overt CMML, multiple gene mutations/aberrations are found. ****Here a CHIP-like mutation is detected – if more than one CHIP-like mutations are found the question is whether the final diagnosis changes to O-CMML. CMML: chronic myelomonocytic leukemia; IMUS: idiopathic monocytosis of unknown (undetermined) significance; ICUS: idiopathic cytopenia of undetermined significance; CCUS: clonal cytopenia of undetermined significance; CHIP: clonal hematopoiesis of indeterminate potential; CCUS: clonal cytopenia of undetermined significance; CMUS: clonal monocytosis of unknown (undetermined) significance; O-CMML: oligomonocytic CMML; MDS: myelodysplastic syndrome; BM: bone marrow; FISH: fluorescence *in situ* hybridization.

with repeated investigations of all disease-related parameters. A summary of non-clonal and clonal conditions potentially preceding CMML is shown in Table 4. With regard to criteria delineating non-clonal pre-diagnostic conditions, like idiopathic cytopenia of undetermined significance from the clonal conditions described above (CHIP, CCUS, CHOP), we refer the reader to the pertinent literature.^{69,71,73}

Peripheral blood and bone marrow smears: proposed standards and recommendations

As in other myeloid neoplasms, a thorough examination of appropriately prepared and stained BM and PB smears is a crucial diagnostic approach in suspected CMML. It is standard to examine and count at least 100 leukocytes in the PB film and 200–500 nucleated cells in well-prepared thin BM films. BM cellularity, the erythroid-to-myeloid (E:M) ratio, and the percentage of blast cells (including monoblasts and promonocytes), monocytes, mast cells, and other myeloid cells must be recorded (reported) in each case. As in patients with MDS, at least 10% of cells in one of the major BM lineages (erythroid and/or neutrophilic and/or megakaryocytic) must be dysplastic to meet the dysplasia criterion for CMML.^{13–18} It is also standard to study well-prepared and appropriately stained PB smears in CMML and to report the percentage of circulating monocytes, including normal (mature) and abnormal (immature) monocytes, blast cells, other immature myeloid cells, dysplastic (hypogranulated) neutrophils and other cell types in the PB. Overall, the same standards and recommendations that count for the evaluation of MDS by morphology (BM and PB stains)^{12,80–83} also apply in cases with (suspected) CMML.^{13–18} An important point is the classification of blast cells and monocytic cells in CMML (Table 5).^{16,84} Blast cell types detectable in CMML include myeloblasts, monoblasts and also promonocytes (even if not named blast cells) (Table 5). Monocytes should be classified as normal (mature) or abnormal (immature).^{16,84} The morphological criteria used to distinguish between these cell types are presented in Table 5. Together with morphology, cytochemical staining for non-specific esterase can also assist in the cytological delineation between monocytes, monoblasts and promonocytes.¹⁶ An important aspect is that in many patients, megakaryocyte dysplasia is

better documented and quantified in BM histology sections than in BM smears. Therefore, megakaryocyte dysplasia should only be recorded in BM smears when a sufficient number of these cells can be detected. Finally, the morphology of mast cells, when detected, should always be reported using established criteria and standards.⁸⁵

Bone marrow histology and immunohistochemistry in CMML

A thorough investigation of an appropriately processed and stained BM biopsy section by histology and immunohistochemistry is standard in all cases with known or suspected CMML or a suspected pre-CMML condition.^{14–16,30,86} Notably, BM histology and immunohistochemistry are essential approaches to confirm the diagnosis of CMML and to exclude AML and other CMML-mimickers. Moreover, BM histology and immunohistochemistry may provide important additional information, including that on BM fibrosis, focal accumulations of blast cells, increased angiogenesis, atypical (dysplastic) megakaryocytes, a hypocellular BM or concomitant mastocytosis (*Online Supplementary Table S4*).^{33–35,86} The evaluation and enumeration of CD14⁺ monocytes, CD34⁺ progenitor cells and CD117⁺/KIT⁺ cells (progenitors and mast cells) by immunohistochemistry in BM biopsy sections represent an integral part of the diagnostic assessment. These approaches can also prevent diagnostic errors. For example, when the smear is of suboptimal quality, a preliminary diagnosis of CMML may change to AML based on BM histology and CD34 immunohistochemistry.

BM biopsy specimens are usually taken from the iliac crest and should be of adequate length (≥2 cm). The specimen should be fixed in neutral formalin (or alternative standard fixation), decalcified in EDTA (for at least 8 h) or by alternative standard decalcification, and embedded in paraffin-wax. Ideally 2–3 μm thin sections should be prepared. Routine stains include hematoxylin-eosin, Giemsa, Prussian blue, AS-D chloroacetate esterase, toluidine blue and silver impregnation (Gömöri's stain). BM cellularity should be measured and reported according to published standards.^{87,88} For routine purposes, the pathologist should determine the cellularity as 'normocellular', 'hypocellular', or 'hypercellular', based on an age-adapted estimate.⁸⁹ The presence of variable degrees of BM fibrosis

Table 5. Classification of blast cells and monocytes in patients with chronic myelomonocytic leukemia.

Cell type	Nuclear shape	Chromatin	Cytoplasm	Size relative to mature monocytes
Blast cells:				
Myeloblast	Round/oval	Fine with nucleoli	Basophilic, rare or no granules	Smaller
Monoblast	Round/oval	Delicate/lace-like, nucleoli	Basophilic, rare azurophilic granules	Large (20–30 μM)
Promonocyte	Convoluted/indented*	Delicate/lace-like, nucleoli	Variably basophilic, variably azurophilic granules	Large
Monocytes:				
Abnormal/immature monocyte	Convoluted/indented	More condensed, rare nucleoli	Intermediate basophilic**	Smaller
Mature monocyte	Lobulated/indented	Condensed, no nucleoli	Gray or pinkish with occasional azurophilic granules and vacuoles	=

*The most important feature discriminating promonocytes from monoblasts. **Less basophilic than promonocytes and more basophilic than mature monocytes.

(usually mild to moderate) has been reported in CMML cases, with several recent studies attempting to determine the prognostic value of this finding.^{42,90,91} Indeed, although the data are not yet conclusive, the presence of marrow fibrosis in CMML seems to be of prognostic importance.^{42,90,91}

The application of immunohistochemical markers is recommended in all patients with (suspected) CMML. The minimal immunohistochemistry panel includes CD14 (monocytes), CD34 (progenitors), CD117/KIT (progenitors and mast cells), tryptase (mast cells), and a megakaryocyte marker (CD41, CD42 or CD61) (*Online Supplementary Table S5*).^{86,92,93} In unclear cases or when a co-existing (second) BM neoplasm is suspected, additional lineage-specific antibodies such as CD3, CD20, or CD25 (suspected mastocytosis) should be applied (*Online Supplementary Table S4*). When employing CD34 as a progenitor-related immunohistochemical marker, it is important to know that endothelial cells also express this antigen. Another important point is that blasts may sometimes be CD34-negative. In such cases, KIT/CD117 is applied as an alternative marker (*Online Supplementary*

Table S4). For the immunohistochemical detection of monocytic cells, CD14 is a preferred antigen.^{71,86} Tryptase and CD117 are useful immunohistochemistry markers for detecting and quantifying mast cells.^{92,93} When spindle-shaped mast cells form compact clusters in the BM and express CD25, these cells usually also display KIT D816V – in these cases the final diagnosis is always SM-CMML.⁹³ In other cases, the pathologist will ask for JAK2 V617F, based on an abnormal morphology and distribution of megakaryocytes. As in MDS, megakaryocytes may also express CD34 in patients with CMML.

Karyotyping in CMML: current recommendations and standards

Clonal cytogenetic abnormalities are detected in 20–30% of all patients with CMML. The most frequently identified aberrations are trisomy 8, abnormalities of chromosome 7 (especially monosomy 7 and deletion of 7q), and loss of the Y chromosome (-Y) (*Online Supplementary Table S6*).⁹⁴⁻⁹⁷ Compared to MDS, isolated del(5q) and complex abnormal karyotypes are rarely detected in CMML. Our faculty is of the opinion that conventional karyotyping of BM cells should be performed in all patients with known or suspected CMML or a suspected pre-CMML condition. At least 20 metaphases should be examined.⁹⁸ In the case of a clear-cut result, even 10–20 metaphases may be sufficient to define the karyogram. Reporting of karyotypes should be performed using the International System for Human Cytogenetic Nomenclature (ISCN) guidelines.⁹⁹ A clone is defined by two or more metaphases showing the same gain or structural rearrangement (deletion, inversion, translocation) of chromosomal material or at least three metaphases showing a monosomy of the same chromosome.⁹⁹ Several of the cytogenetic anomalies in CMML may be difficult to detect by conventional karyotyping. Therefore, we are of the opinion that fluorescence *in situ* hybridization (FISH) should be performed in all patients with (suspected) CMML, at least in those in whom no karyotype anomaly was detected by conventional karyotyping. The FISH probes should cover all relevant regions, including 5q31, 7q31, 20q, 7p11, 7p13, and p53. Special consideration should be directed to cryptic deletions of TET2 (in 4q24), NF1 (17q11), and ETV6 (12p13) which can occur in up to 10% of CMML patients¹⁰ and are only detectable by interphase FISH (*Online Supplementary Table S6*). It is worth noting that NF1 deletions may occur during progression/karyotype evolution in CMML. The limitation of FISH is that it does not detect all karyotypic abnormalities. In some patients with CMML, clonal evolution is found. Subclones are defined by additional chromosomal defects (apart from the primary chromosomal defect) in at least two cells (or 3 cells for monosomies) and the absence of these additional chromosomal defects in the other clonal cells.⁹⁹ A complex karyotype is defined by at least three chromosome defects in one clone.⁹⁹ As in MDS, a complex karyotype in CMML is indicative of a poor prognosis. Overall, cytogenetic studies are of prognostic significance in CMML and have been used to optimize prognostic scoring systems.^{97,100-102} In some patients with CMML, clonal evolution is observed over time and may then also be an adverse prognostic sign. Therefore, we recommend that chromosome analyses are performed each time a BM investigation is done in the follow-up in order to detect (or exclude) clonal evolution.

Table 6. Commonly mutated genes detectable in patients with classical chronic myelomonocytic leukemia.

Gene name abbreviation	Gene class and function	Relative frequency in CMML	Clinical impact
ASXL1	Epigenetic regulation Histone modification	40%*	Poor prognosis** CHIP/ARCH***
EZH2	Epigenetic regulation Histone modification	5%	
TET2	Epigenetic regulation DNA methylation	60%*	CHIP/ARCH***
DNMT3A	Epigenetic regulation DNA methylation	5%	Poor prognosis** CHIP/ARCH***
IDH1	Epigenetic regulation	1%	Drug target
IDH2	Epigenetic regulation	5-10%	Drug target
CBL	Signaling	15%	RAS pathway
NRAS	Signaling	15%	Poor prognosis** RAS pathway
KRAS	Signaling	10%	RAS pathway
PTPN11	Signaling	5%	RAS pathway
FLT3	Signaling	<5%	AML-related Drug target
SRSF2	Pre-mRNA splicing	50%*	
SF3B1	Pre-mRNA splicing	5-10%	
U2AF1	Pre-mRNA splicing	5-10%	
ZRSR2	Pre-mRNA splicing	5%	
RUNX1	Gene transcription	15%	Poor prognosis** AML-related
SETBP1	Gene transcription	15%	Poor prognosis**
TP53	DNA damage	1%	Poor prognosis**
PHF6	Chromatin adaptor	5%	

*These mutations can be regarded as CMML-related mutations, but only SRSF2 mutations do not, in addition, also count as classical CHIP/ARCH mutations. **Mutations in these genes are independent adverse prognostic factors regarding survival in CMML. ***These genes are frequently detected in individuals with clonal hematopoiesis of indeterminate potential (CHIP) also known as age-related clonal hematopoiesis (ARCH). Therefore, the diagnostic impact of these mutations may be regarded as somehow lower compared to that of other (CMML-related and other) mutations. CMML: chronic myelomonocytic leukemia; AML: acute myeloid leukemia.

Mutation profiles in CMML: current standards and limitations

Somatic mutations are detectable in the vast majority of patients with CMML.^{8,11,103-106} The clonal architecture, clone sizes and clonal evolution patterns vary from patient to patient.¹⁰⁶⁻¹⁰⁸ In some cases, initially small clones expand over time. It is, therefore, standard to apply next-generation sequencing assays with sufficient sensitivity to identify *bona fide* somatic mutations associated with CMML. The most frequently detected somatic mutations in CMML are mutations in *TET2* (60%), *SRSF2* (50%), and *ASXL1* (40%) (Table 6).^{31,103-110} The presence of a *SRSF2* mutation, particularly in combination with mutated *TET2*, correlates strongly with a CMML phenotype.^{31,109,110} It is also worth noting that two of these mutations (*TET2*, *ASXL1*) are also known as CHIP/ARCH-related mutations. However, only mutated *ASXL1* has been associated with a poor prognosis in CMML.^{104,109} An overview of somatic mutations recurrently detected in CMML is provided in Table 6. Somatic mutations with independent prognostic impact include several RAS-pathway mutations as well as mutations in *ASXL1*, *RUNX1* and *SETBP1* (Table 6).^{31,103-111} RAS-pathway mutations trigger cell signaling and proliferation and have been associated with cytokine-independent growth of CMML progenitor cells, the proliferative variant of CMML, AML transformation and poor survival.^{10,22,23,112-116} Other driver mutations involved in cell signaling, such as *JAK2* V617F or *KIT* D816V, are also major triggers of cellular differentiation (Online Supplementary Table S7). These drivers alone cannot induce transformation, but they may act together with other (e.g., ‘RAS pathway’) mutations to cause disease progression. Whereas *JAK2* V617F is a strong indicator of MPN-like differentiation, the presence of *KIT* D816V is almost always associated with concomitant mast cell differentiation and mastocytosis (SM-CMML).^{6-8,32-36,39,42,43} The other mutations found in CMML act as modulators of epigenetic events and transcription (e.g., *ASXL1*) or DNA methylation (e.g., *TET2*), as regulators of the spliceosome machinery (e.g., *SRSF2*), or as modulators of the DNA damage response, such as *TP53* (Table 6). During progression of CMML to secondary AML and especially during therapy, the mutational landscape(s) and clonal architecture(s) may change.¹⁰⁹⁻¹¹³ For example initially small clones may expand and may be selected because of resistance-mediating molecular features. It is worth noting that several mutated gene products also serve as potential targets of therapy (Table 6).

Our faculty recommends that next-generation sequencing studies should be regarded as a standard approach in all patients with suspected or known CMML as well as in patients with idiopathic monocytosis of unknown significance and in those with persistent reactive monocytosis (in order to exclude an additional clonal component). When a CMML-related mutation is found in an individual with idiopathic monocytosis of unknown significance or reactive monocytosis, the diagnosis may change to clonal monocytosis of unknown significance or oligomonocytic CMML, depending on additional findings.

Our faculty also recommends that the next-generation sequencing assay should have sufficient sensitivity (to detect 2-5% clonal cells) and should cover all relevant lesions shown in Table 6. In the context of CHIP/ARCH, a cutoff variant allele frequency of 2% is considered diagnostic,⁶⁹ whereas in the context of CMML, we propose 10% as the variant allele frequency diagnostic cut off and thus marker to count as a co-criterion of CMML when, for example, no diagnostic morphological dysplasia can be documented (Tables 1 and 3), similar to the definition in MDS.^{71,73} Determining the variant allele frequency is also useful for documenting the clinical impact of certain driver lesions in special CMML variants (e.g., with *JAK2* V617F or *KIT* D816V) and clone expansion during follow-up. Therefore, our faculty recommends that molecular studies in CMML should report variant allele frequencies with sufficient precision and sufficient sensitivity – in the same way as in MDS.^{71,73} Finally, our faculty recommends that molecular markers should increasingly be used to optimize prognostic scoring systems in CMML.¹¹⁷⁻¹²⁰

Flow cytometry in CMML: standards and limitations

Flow cytometry studies are an essential diagnostic tool in patients with (suspected) classical CMML, pre-CMML conditions and special CMML variants.¹²¹⁻¹³² Therefore, our faculty is of the opinion that it is standard practice to perform multi-color flow cytometry (MFC) in the PB and BM in all cases with suspected or known CMML or a suspected pre-CMML condition. MFC studies are helpful to confirm the monocyte and blast cell counts in these patients and to exclude AML. In addition, MFC is useful to confirm the presence of distinct monocyte populations. Monocytes are defined as CD14⁺ cells in these analyses. Based on the expression of CD14 and CD16, monocytes are further divided into classical (MO1) monocytes (CD14^{bright}/CD16⁻), intermediate (MO2) monocytes (CD14^{bright}/CD16⁺) and non-classical (MO3) monocytes (CD14^{dim}/CD16⁺) (Table 7).^{127,128,132} Compared to age-matched healthy donors¹³³ and patients with reactive monocytosis, but also myeloid neoplasms other than CMML (even MDS), the percentages of MO1 monocytes in the PB are higher and the percentage of MO3 monocytes is lower in patients with CMML.^{127,131,132} When the absolute monocyte count is increased in the PB, a cutoff value of >94% MO1 monocytes, based on their

Table 7. Phenotypic classification of monocytes and distribution of monocyte subsets in patients with chronic myelomonocytic leukemia and in controls.*

Monocyte-subset	Defining phenotype	CMMML	Typical relative frequency in: MDS or MPN	Reactive BM
Classical (MO1)	CD14 ^{bright} /CD16 ⁻	≥94%	70-97%	<94%
Intermediate (MO2)	CD14 ^{bright} /CD16 ⁺	<20%	5-20%	5-15%
Non-classical (MO3)	CD14 ^{dim} /CD16 ⁺	<5%	5-10%	5-20%

*Data refer to published results presented in references #125 through #132. CMMML: chronic myelomonocytic leukemia; MDS: myelodysplastic syndrome; MPN: myeloproliferative neoplasm; BM: bone marrow.

immunophenotype, can identify CMML with a sensitivity of >90% and a specificity of >95%.^{127,129,131} Moreover, during successful therapy, the distribution of MO1, MO2, and MO3 monocytes changes back to near normal or normal.¹²⁸ Therefore, our faculty recommends that the percentages of MO1 monocytes are quantified in the PB by MFC in all cases with suspected or known CMML at diagnosis and during follow-up.

In many cases with CMML, neoplastic monocytes aberrantly display CD2, CD5, CD10, CD23, and/or CD56.¹²¹⁻¹²⁴ Of all aberrantly expressed surface markers, CD56 is most commonly detected on CMML monocytes.¹²¹⁻¹²⁴ CD5 is only (very) weakly expressed on neoplastic monocytes in most cases with CMML. The most frequently underexpressed antigens may be CD14 and CD15. Overall, however, the use of decreased expression of these markers as a diagnostic test in CMML is limited by a relatively low sensitivity. An abnormal monocyte immunophenotype is also seen in other myeloid neoplasms, including MDS. On the other hand, phenotypically aberrant monocytes (as described above) are typically neoplastic cells (unless the patient has been treated with growth factors). Therefore, our faculty recommends that MFC studies in patients with (suspected) CMML employ antibodies directed against aberrantly expressed surface markers, including CD2 and CD56. Additionally, as mentioned before, several surface markers are ‘under-expressed’ on CMML monocytes compared to their levels on normal blood monocytes. These antigens include, among others, CD13, CD14, CD15, CD33, CD38, CD45, and CD64.^{121-124,129,131}

Other cell types may also express aberrant markers detectable by MFC in CMML. For example, myeloid progenitor cells may express CD56 in CMML and often exhibit the same phenotypic abnormalities as in MDS; this also holds true for neutrophils and erythroid cells (*Online Supplementary Table S8*). Other cell types that may show aberrant phenotypes are dendritic cells and mast cells. Mast cells are of particular importance as these cells may be indicative of the presence of a concomitant mastocytosis (SM-CMML). In these cases, mast cells almost invariably express CD25 in MFC analyses (*Online Supplementary Table S8*).¹³⁴ Overall, our faculty is of the opinion that MFC studies should be performed on monocyte subsets, myeloid progenitors, neutrophils, erythroid cells and mast cells in (suspected) CMML. An overview of immunophenotypic aberrancies detectable in CMML is given in *Online Supplementary Table S8*.

Differential diagnoses of CMML: reactive and clonal mimickers

A number of conditions can mimick CMML and must be taken into account when patients with unexplained monocytosis are evaluated. Reactive disorders mimicking CMML include certain chronic bacterial infections (examples: tuberculosis or subacute endomyocarditis), fungal infections, chronic auto-immune processes and non-hematologic neoplasms. There are also hematologic malignancies that may present as a CMML-like disease. For example, Philadelphia chromosome-positive chronic myeloid leukemia usually presents with (absolute) monocytosis and can also show signs of dysplasia. Particularly high monocyte counts are recorded in chronic myeloid leukemia cases expressing *BCR-ABL1*_{p190}. When cryptic variants of *BCR-ABL1* are expressed by leukemic cells, it

can be difficult to exclude CMML. Myeloid neoplasms (MDS or MPN) in progression and myelomonocytic or monocytic AML may also resemble CMML. The reactive and clonal mimickers of CMML are listed in *Online Supplementary Table S9*.

Scoring systems in CMML: recommended standards

Although several prognostic variables have been identified in CMML regarding survival and AML evolution, accurate prediction of the clinical course and survival remains a clinical challenge. A first step in prognostication is grading into CMML-0, CMML-1 and CMML-2. To delineate the prognosis in CMML more accurately, a number of scoring systems have been developed in the past.^{29,117-121,155-158} Until 2012, the International Prognostic Scoring System (IPSS) served as the gold standard for prognostication in MDS and (dysplastic) CMML.¹³⁵

However, a number of more specific scoring systems taking CMML-related features into account have also been proposed.^{117-120,136-138} During the past few years, researchers have successfully started to integrate cytogenetic and molecular variables into these scoring models.¹¹⁷⁻¹²¹ Our faculty concludes that these novel approaches should be followed and developed into clinical application.

Management strategies and therapeutic options in CMML

Several new treatment strategies for CMML have been developed during the past 15 years. A detailed description of therapeutic options is beyond the scope of this article. The reader is referred to a series of excellent published review articles.¹³⁹⁻¹⁴⁶ A disappointing fact is that all drug therapies are still non-curative. The only curative therapy in CMML remains allogeneic hematopoietic stem cell transplantation.^{147,148} For most young and eligible patients with acceptable transplant-related risk, allogeneic hematopoietic stem cell transplantation is therefore recommended. All other forms of treatment are cytoreductive, experimental or palliative in nature. Some of these drugs, such as the hypomethylating agents (5-azacytidine, decitabine) may induce long-term disease control in a subset of patients with classical CMML.¹³⁹⁻¹⁴⁵ In general, cytoreductive and palliative drugs should be used according to available recommendations provided by major societies.^{145,148} Similarly, treatment response assessment should be performed in line with available (accepted) guidelines.^{146,150}

Specific therapy may work in those patients who suffer from a special variant of CMML. For example, in CMML patients with a transforming *PDGFRA/B* mutation, treatment with imatinib or other similar tyrosine kinase inhibitor usually induces major responses or even long-lasting remissions.^{47-49,151} In patients with SM-CMML, midostaurin may result in disease control, especially when the CMML portion of the disease exhibits *KIT D816V*. However, in many cases, relapses occur. Treatment options in CMML and its variants are summarized in *Online Supplementary Table S10*.

Concluding remarks and future perspectives

CMML is a unique and rare hematopoietic neoplasm with a complex biology and pathology. In the past 10

years, several different pre-CMML conditions and sub-variants of CMML have been defined. In the current article, we propose minimal diagnostic criteria for classical CMML and for special CMML variants. These criteria should help in the diagnosis of pre-CMML conditions, classical CMML, special CMML variants, and conditions that mimick CMML. In addition, we propose standards and tools for the diagnosis, prognostication and management of CMML. Contemporary assays define all major histopathological, molecular, cytogenetic and flow cytometry-based features of neoplastic cells, and thereby cover all CMML variants, including oligomonocytic CMML and CMML associated with certain drivers or a concomitant myeloid neoplasm, such as mastocytosis. Different aberration profiles may also be found, resulting in a quite heterogeneous clinical picture and a variable clinical course. Although the course is often unpre-

dictable, initial grading and consecutive application of CMML-directed prognostic scores are standard tools that support the prognostication of patients with CMML concerning survival and AML evolution. The application of criteria, tools and standards proposed herein should assist in the diagnosis, prognostication and management of patients with CMML.

Acknowledgments

We thank Sabine Sonnleitner, Sophia Rammel, Susanne Gampel, Emir Hadzijusofovic and all other members of Peter Valent's research group involved for their excellent support in the organization of the Working Conference. This study was supported by Austrian Science Fund (FWF) grants F4701-B20 and F4704-B20, the Ludwig Boltzmann Institute for Hematology and Oncology (LBI HO) and a Stem Cell Research grant from the Medical University of Vienna.

References

1. Storniolo AM, Moloney WC, Rosenthal DS, Cox C, Bennett JM. Chronic myelomonocytic leukemia. Leukemia. 1990;4(11):766-770.
2. Bennett JM, Catovsky D, Daniel MT, et al. The chronic myeloid leukaemias: guidelines for distinguishing chronic granulocytic, atypical chronic myeloid, and chronic myelomonocytic leukaemia. Proposals by the French-American-British Cooperative Leukaemia Group. Br J Haematol. 1994;87(4):746-754.
3. Bennett JM. Chronic myelomonocytic leukemia. Curr Treat Options Oncol. 2002;3(3):221-223.
4. Patnaik MM, Parikh SA, Hanson CA, Tefferi A. Chronic myelomonocytic leukaemia: a concise clinical and pathophysiological review. Br J Haematol. 2014;165(3):273-286.
5. Itzykson R, Duchmann M, Lucas N, Solaro E. CMML: clinical and molecular aspects. Int J Hematol. 2017;105(6):711-719.
6. Sperr WR, Horny HP, Lechner K, Valent P. Clinical and biologic diversity of leukemias occurring in patients with mastocytosis. Leuk Lymphoma. 2000;37(5-6):473-486.
7. Sotlar K, Fridrich C, Mall A, et al. Detection of c-kit point mutation Asp-816 --> Val in microdissected pooled single mast cells and leukemic cells in a patient with systemic mastocytosis and concomitant chronic myelomonocytic leukemia. Leuk Res. 2002;26(11):979-984.
8. Patnaik MM, Rangit Vallapureddy, Lasho TL, et al. A comparison of clinical and molecular characteristics of patients with systemic mastocytosis with chronic myelomonocytic leukemia to CMML alone. Leukemia. 2018;32(8):1850-1856.
9. McCullough KB, Patnaik MM. Chronic myelomonocytic leukemia: a genetic and clinical update. Curr Hematol Malig Rep. 2015;10(3):292-302.
10. Kohlmann A, Grossmann V, Klein HU, et al. Next-generation sequencing technology reveals a characteristic pattern of molecular mutations in 72.8% of chronic myelomonocytic leukemia by detecting frequent alterations in TET2, CBL, RAS, and RUNX1. J Clin Oncol. 2010;28(24):3858-3865.
11. Patnaik MM, Tefferi A. Cytogenetic and molecular abnormalities in chronic myelomonocytic leukemia. Blood Cancer J. 2016;6:e393.
12. Bennett JM, Catovsky D, Daniel MT, et al. Proposals for the classification of the myelodysplastic syndromes. Br J Haematol. 1982;51(2):189-199.
13. Vardiman JW, Imbert M, Pierre R, et al. Chronic myelomonocytic leukemia. In: World Health Organization Classification of Tumours – Pathology & Genetics: Tumours of the Haematopoietic and Lymphoid Tissues: Eds. Jaffe ES, Harris NL, Stein H, Vardiman JW. IARC Press Lyon 2001, pp 49-52.
14. Orazi A, Bennett JM, Germing U, Brunning RD, Bain B, Thiele J. Chronic myelomonocytic leukemia. In: WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues: Eds. Swerdlow SH, Campo E, Harris NL, et al. International Agency for Research on Cancer - IARC Press Lyon 2008, pp 76-79.
15. Arber DA, Orazi A, Hasserjian R, et al. The 2016 revision to the World Health Organization classification of myeloid neoplasms and acute leukemia. Blood. 2016;127(20):2391-2405.
16. Orazi A, Bain B, Bennett JM, et al. Chronic myelomonocytic leukemia. In: WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues: Eds. Swerdlow SH, Campo E, Harris NL, et al. International Agency for Research on Cancer - IARC Press Lyon 2017, pp 82-86.
17. Schuler E, Schroeder M, Neukirchen J, et al. Refined medullary blast and white blood cell count based classification of chronic myelomonocytic leukemias. Leuk Res. 2014;38(12):1413-1419.
18. Bennett JM. Changes in the updated 2016: WHO classification of the myelodysplastic syndromes and related myeloid neoplasms. Clin Lymphoma Myeloma Leuk. 2016;16(11):607-609.
19. Moon Y, Kim MH, Kim HR, et al. The 2016 WHO versus 2008 WHO criteria for the diagnosis of chronic myelomonocytic leukemia. Ann Lab Med. 2018;38(5):481-488.
20. Bain BJ, Gilliland DG, Horny HP, Vardiman JW. Myeloid and lymphoid neoplasms with eosinophilia and abnormalities of PDGFRA, PDGFRB or FGFR1. In: WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues: Eds. Swerdlow SH, Campo E, Harris NL, et al. International Agency for Research on Cancer - IARC Press Lyon 2008, pp 68-73.
21. Bain BJ, Horny HP, Arber DA, Tefferi A, Hasserjian RP. Myeloid/lymphoid neoplasms with eosinophilia and rearrangement of PDGFRA, PDGFRB or FGFR1, or with PCM1-JAK2. In: WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues: Eds. Swerdlow SH, Campo E, Harris NL, et al. International Agency for Research on Cancer - IARC Press Lyon 2017, pp 72-79.
22. Parikh C, Subrahmanyam R, Ren R. Oncogenic NRAS rapidly and efficiently induces CMML- and AML-like diseases in mice. Blood. 2006;108(7):2349-2357.
23. Gelsi-Boyer V, Trouplin V, Adélaïde J, et al. Genome profiling of chronic myelomonocytic leukemia: frequent alterations of RAS and RUNX1 genes. BMC Cancer. 2008; 8:299.
24. Reinig E, Yang F, Traer E, et al. Targeted next-generation sequencing in myelodysplastic syndrome and chronic myelomonocytic leukemia aids diagnosis in challenging cases and identifies frequent spliceosome mutations in transformed acute myeloid leukemia. Am J Clin Pathol. 2016;145(4): 497-506.
25. Benton CB, Nazha A, Pemmaraju N, Garcia-Manero G. Chronic myelomonocytic leukemia: forefront of the field in 2015. Crit Rev Oncol Hematol. 2015;95(2):222-242.
26. Sallman DA, Padron E. Transformation of the clinical management of CMML patients through in-depth molecular characterization. Clin Lymphoma Myeloma Leuk. 2015;15S:S50-55.
27. Patnaik MM, Tefferi A. Chronic myelomonocytic leukemia: 2016 update on diagnosis, risk stratification, and management. Am J Hematol. 2016;91(6):631-642.
28. Onida F. Models of prognostication in chronic myelomonocytic leukemia. Curr Hematol Malig Rep. 2017;12(6):513-521.
29. Nazha A, Patnaik MM. Making sense of prognostic models in chronic myelomonocytic leukemia. Curr Hematol Malig Rep. 2018;13(5):341-347.

30. Geyer JT, Tam W, Liu YC, et al. Oligomonocytic chronic myelomonocytic leukemia (chronic myelomonocytic leukemia without absolute monocytosis) displays a similar clinicopathologic and mutational profile to classical chronic myelomonocytic leukemia. *Mod Pathol.* 2017;30(9):1213-1222.
31. Malcovati L, Papaemmanuil E, Ambaglio I, et al. Driver somatic mutations identify distinct disease entities within myeloid neoplasms with myelodysplasia. *Blood.* 2014;124(9):1513-1521.
32. Schuler E, Frank F, Hildebrandt B, et al. Myelodysplastic syndromes without peripheral monocytosis but with evidence of marrow monocytosis share clinical and molecular characteristics with CMML. *Leuk Res.* 2018;65:1-4.
33. Valent P, Horny HP, Escrivano L, et al. Diagnostic criteria and classification of mastocytosis: a consensus proposal. *Leuk Res.* 2001;25(7):603-625.
34. Valent P, Akin C, Metcalfe DD. Mastocytosis: 2016 updated WHO classification and novel emerging treatment concepts. *Blood.* 2017;129(11):1420-1427.
35. Valent P, Akin C, Hartmann K, et al. Advances in the classification and treatment of mastocytosis: current status and outlook toward the future. *Cancer Res.* 2017;77(6):1261-1270.
36. Sperr WR, Horny HP, Valent P. Spectrum of associated clonal hematologic non-mast cell lineage disorders occurring in patients with systemic mastocytosis. *Int Arch Allergy Immunol.* 2002;127(2):140-142.
37. Gotlib J, Kluin-Nelemans HC, George TI, et al. Efficacy and safety of midostaurin in advanced systemic mastocytosis. *N Engl J Med.* 2016;374(26):2530-2541.
38. Tefferi A, Gilliland DG. Oncogenes in myeloproliferative disorders. *Cell Cycle.* 2007;6(5):550-566.
39. Pich A, Riera L, Sismondi F, et al. JAK2V617F activating mutation is associated with the myeloproliferative type of chronic myelomonocytic leukaemia. *J Clin Pathol.* 2009;62(9):798-801.
40. Bacher U, Haferlach T, Schnittger S, Kreipe H, Kröger N. Recent advances in diagnosis, molecular pathology and therapy of chronic myelomonocytic leukaemia. *Br J Haematol.* 2011;153(2):149-167.
41. Bell GC, Padron E. Detection of a PDGFRB fusion in refractory CMML without eosinophilia: a case for broad spectrum tumor profiling. *Leuk Res Rep.* 2015;4(2):70-71.
42. Gur HD, Loghavi S, Garcia-Manero G, et al. Chronic myelomonocytic leukemia with fibrosis is a distinct disease subset with myeloproliferative features and frequent JAK2 p.V617F mutations. *Am J Surg Pathol.* 2018;42(6):799-806.
43. Hu Z, Ramos CB, Medeiros LJ, et al. Utility of JAK2 V617F allelic burden in distinguishing chronic myelomonocytic leukemia from primary myelofibrosis with monocytosis. *Hum Pathol.* 2019;85:209-298.
44. Chapman J, Geyer JT, Khanlari M, et al. Myeloid neoplasms with features intermediate between primary myelofibrosis and chronic myelomonocytic leukemia. *Mod Pathol.* 2018;31(3):429-441.
45. Patnaik MM, Pophali PA, Lasho TL, et al. Clinical correlates, prognostic impact and survival outcomes in chronic myelomonocytic leukemia patients with the JAK2V617F mutation. *Haematologica.* 2019;104(6):e236-239.
46. Khorashad JS, Tantravahi SK, Yan D, et al. Rapid conversion of chronic myeloid leukemia to chronic myelomonocytic leukemia in a patient on imatinib therapy. *Leukemia.* 2016;30(11):2275-2279.
47. Magnusson MK, Meade KE, Nakamura R, Barrett J, Dunbar CE. Activity of ST1571 in chronic myelomonocytic leukemia with a platelet-derived growth factor beta receptor fusion oncogene. *Blood.* 2002;100(3):1088-1091.
48. Apperley JF, Gardembas M, Melo JV, et al. Response to imatinib mesylate in patients with chronic myeloproliferative diseases with rearrangements of the platelet-derived growth factor receptor beta. *N Engl J Med.* 2002;347(7):481-487.
49. Reiter A, Walz C, Cross NC. Tyrosine kinases as therapeutic targets in BCR-ABL negative chronic myeloproliferative disorders. *Curr Drug Targets.* 2007;8(2):205-216.
50. Shah S, Loghavi S, Garcia-Manero G, Khoury JD. Discovery of imatinib-responsive FIP1L1-PDGFRα mutation during refractory acute myeloid leukemia transformation of chronic myelomonocytic leukemia. *J Hematol Oncol.* 2014;7:26.
51. Ueki K, Sato S, Tamura J, et al. Three cases of multiple myeloma developing into melphalan-related chronic myelomonocytic leukemia. *J Med.* 1991;22(3):157-161.
52. Kouides PA, Bennett JM. Transformation of chronic myelomonocytic leukemia to acute lymphoblastic leukemia: case report and review of the literature of lymphoblastic transformation of myelodysplastic syndrome. *Am J Hematol.* 1995;49(2):157-162.
53. Yamamoto M, Nakagawa M, Ichimura N, et al. Lymphoblastic transformation of chronic myelomonocytic leukemia in an infant. *Am J Hematol.* 1996;52(3):212-214.
54. Gaulier A, Jary-Bourguignat L, Serna R, Pulik M, Davi F, Raphaël M. Occurrence of angioimmunoblastic T cell lymphoma in a patient with chronic myelomonocytic leukemia features. *Leuk Lymphoma.* 2000;40(1-2):197-204.
55. Robak T, Urbańska-Ryś H, Smolewski P, et al. Chronic myelomonocytic leukemia coexisting with B-cell chronic lymphocytic leukemia. *Leuk Lymphoma.* 2003;44(11):2001-2008.
56. Menter T, Schlageter M, Bastian L, Haberthür R, Rätz Bravo AE, Tzankov A. Development of an Epstein-Barr virus-associated lymphoproliferative disorder in a patient treated with azacitidine for chronic myelomonocytic leukaemia. *Hematol Oncol.* 2014;32(1):47-51.
57. Pemmaraju N, Shah D, Kantarjian H, et al. Characteristics and outcomes of patients with multiple myeloma who develop therapy-related myelodysplastic syndrome, chronic myelomonocytic leukemia, or acute myeloid leukemia. *Clin Lymphoma Myeloma Leuk.* 2015;15(2):110-114.
58. Hágihara M, Inoue M, Kodama K, Uchida T, Hua J. Simultaneous manifestation of chronic myelomonocytic leukemia and multiple myeloma during treatment by prednisolone and eltrombopag for immune-mediated thrombocytopenic purpura. *Case Rep Hematol.* 2016;2016: 4342820.
59. Saillard C, Guermouche H, Derriue C, et al. Response to 5-azacytidine in a patient with TET2-mutated angioimmunoblastic T-cell lymphoma and chronic myelomonocytic leukaemia preceded by an EBV-positive large B-cell lymphoma. *Hematol Oncol.* 2017;35(4):864-868.
60. Soriano PK, Stone T, Baqai J, Sana S. A case of synchronous bone marrow chronic myelomonocytic leukemia (CMML) and nodal marginal zone lymphoma (NMZL). *Am J Case Rep.* 2018;19:1135-1139.
61. Wang SA, Galili N, Cerny J, et al. Chronic myelomonocytic leukemia evolving from preexisting myelodysplasia shares many features with de novo disease. *Am J Clin Pathol.* 2006;126(5):789-797.
62. Breccia M, Cannella L, Frustaci A, Stefanizzi C, D'Elia GM, Alimena G. Chronic myelomonocytic leukemia with antecedent refractory anemia with excess blasts: further evidence for the arbitrary nature of current classification systems. *Leuk Lymphoma.* 2008;49(7):1292-1296.
63. Ahmed F, Osman N, Lucas F, et al. Therapy related CMML: a case report and review of the literature. *Int J Hematol.* 2009;89(5):699-703.
64. Singh ZN, Post GR, Kiwan E, Maddox AM. Cytopenia, dysplasia, and monocytosis: a precursor to chronic myelomonocytic leukemia or a distinct subgroup? Case reports and review of literature. *Clin Lymphoma Myeloma Leuk.* 2011;11(3):293-297.
65. Subari S, Patnaik M, Alfakara D, et al. Patients with therapy-related CMML have shorter median overall survival than those with de novo CMML: Mayo Clinic long-term follow-up experience. *Clin Lymphoma Myeloma Leuk.* 2015;15(9):546-549.
66. Patnaik MM, Vallapureddy R, Yalniz FF, et al. Therapy related-chronic myelomonocytic leukemia (CMML): molecular, cytogenetic, and clinical distinctions from de novo CMML. *Am J Hematol.* 2018;93(1):65-73.
67. Busque L, Patel JP, Figueroa ME, et al. Recurrent somatic TET2 mutations in normal elderly individuals with clonal hematopoiesis. *Nat Genet.* 2012;44:1179-1181.
68. Genovese G1, Kähler AK, Handsaker RE, et al. Clonal hematopoiesis and blood-cancer risk inferred from blood DNA sequence. *N Engl J Med.* 2014;371(26):2477-2487.
69. Steensma DP, Bejar R, Jaiswal S, et al. Clonal hematopoiesis of indeterminate potential and its distinction from myelodysplastic syndromes. *Blood.* 2015;126(1):9-16.
70. Jaiswal S, Fontanillas P, Flannick J, et al. Age-related clonal hematopoiesis associated with adverse outcomes. *N Engl J Med.* 2014;371(26):2488-2498.
71. Valent P, Orazi A, Steensma DP, et al. Proposed minimal diagnostic criteria for myelodysplastic syndromes (MDS) and potential pre-MDS conditions. *Oncotarget.* 2017;8(43):73483-73500.
72. Gibson CJ, Lindsley RC, Tchekmedyan V, et al. Clonal hematopoiesis associated with adverse outcomes after autologous stem cell transplantation for lymphoma. *J Clin Oncol.* 2017;35(14):1598-1605.
73. Valent P, Akin C, Arock M, et al. Proposed terminology and classification of pre-malignant neoplastic conditions: a consensus proposal. *EBioMedicine.* 2017;26:17-24.
74. Elbæk MV, Sørensen AL, Hasselbalch HC. Chronic inflammation and autoimmunity as risk factors for the development of chronic myelomonocytic leukemia? *Leuk Lymphoma.* 2016;57(8):1793-1799.
75. Grignano E, Mekinian A, Braun T, et al. Autoimmune and inflammatory diseases associated with chronic myelomonocytic leukemia: a series of 26 cases and literature review. *Leuk Res.* 2016;47:136-141.
76. Deininger MWN, Tyner JW, Solary E. Turning the tide in myelodysplastic/myelo-

- proliferative neoplasms. *Nat Rev Cancer*. 2017;17(7):425-440.
77. Mainwaring CJ, Shutt J, James CM. Not all cases of idiopathic thrombocytopenic purpura (correction of pupura) are what they might first seem. *Clin Lab Haematol*. 2002;24(4):261-262.
 78. Cai Y, Teng R, Lin Z, Zhang Y, Liu H. Chronic myelomonocytic leukemia presenting as relapsing thrombotic thrombocytopenic purpura. *Aging Clin Exp Res*. 2013;25(3):349-350.
 79. Hadjadj J, Michel M, Chauveheid MP, Godeau B, Papo T, Sacre K. Immune thrombocytopenia in chronic myelomonocytic leukemia. *Eur J Haematol*. 2014;93(6):521-526.
 80. Valent P, Horny HP, Bennett JM, et al. Definitions and standards in the diagnosis and treatment of the myelodysplastic syndromes: consensus statements and report from a working conference. *Leuk Res*. 2007;31(6):727-736.
 81. Mufti GJ, Bennett JM, Goasguen J, et al. Diagnosis and classification of myelodysplastic syndrome: International Working Group on Morphology of Myelodysplastic Syndrome (IWGM-MDS) consensus proposals for the definition and enumeration of myeloblasts and ring sideroblasts. *Haematologica*. 2008;93(11):1712-1717.
 82. Germing U, Strupp C, Giagounidis A, et al. Evaluation of dysplasia through detailed cytomorphology in 3156 patients from the Düsseldorf Registry on myelodysplastic syndromes. *Leuk Res*. 2012;36(6):727-734.
 83. Della Porta MG, Travaglino E, Boveri E, et al. Minimal morphological criteria for defining bone marrow dysplasia: a basis for clinical implementation of WHO classification of myelodysplastic syndromes. *Leukemia*. 2015;29(1):66-75.
 84. Goasguen JE, Bennett JM, Bain BJ, Vallespi T, Brunning R, Mufti GJ; International Working Group on Morphology of Myelodysplastic Syndrome. Morphological evaluation of monocytes and their precursors. *Haematologica*. 2009;94(7):994-997.
 85. Sperr WR, Escrivano L, Jordan JH, et al. Morphologic properties of neoplastic mast cells: delineation of stages of maturation and implication for cytological grading of mastocytosis. *Leuk Res*. 2001;25(7):529-536.
 86. Orazi A, Chiu R, O'Malley DP, et al. Chronic myelomonocytic leukemia: the role of bone marrow biopsy immunohistology. *Mod Pathol*. 2006;19(12):1536-1545.
 87. Tuzuner N, Bennett JM. Reference standards for bone marrow cellularity. *Leuk Res*. 1994;18(8):645-647.
 88. Tuzuner N, Cox C, Rowe JM, Bennett JM. Bone marrow cellularity in myeloid stem cell disorders: impact of age correction. *Leuk Res*. 1994;18(8):559-564.
 89. Schemenau J, Baldus S, Anlauf M, et al. Cellularity, characteristics of hematopoietic parameters and prognosis in myelodysplastic syndromes. *Eur J Haematol*. 2015;95(3):181-189.
 90. Petrova-Drus K, Chiu A, Margolskee E, et al. Bone marrow fibrosis in chronic myelomonocytic leukemia is associated with increased megakaryopoiesis, splenomegaly and with a shorter median time to disease progression. *Oncotarget*. 2017;8(61):103274-103282.
 91. Khan M, Muzzafar T, Kantarjian H, et al. Association of bone marrow fibrosis with inferior survival outcomes in chronic myelomonocytic leukemia. *Ann Hematol*. 2018;97(7):1183-1191.
 92. Horny HP, Sotlar K, Valent P. Diagnostic value of histology and immunohistochemistry in myelodysplastic syndromes. *Leuk Res*. 2007;31(12):1609-1616.
 93. Horny HP, Sotlar K, Sperr WR, Valent P. Systemic mastocytosis with associated clonal haematological non-mast cell lineage diseases: a histopathological challenge. *J Clin Pathol*. 2004;57(6):604-608.
 94. Fugazza G, Bruzzone R, Dejana AM, et al. Cytogenetic clonality in chronic myelomonocytic leukemia studied with fluorescence in situ hybridization. *Leukemia*. 1995;9(1):109-114.
 95. Haase D, Germing U, Schanz J, et al. New insights into the prognostic impact of the karyotype in MDS and correlation with subtypes: evidence from a core dataset of 2124 patients. *Blood*. 2007;110(13):4385-4395.
 96. Wassie EA, Itzykson R, Lasho TL, et al. Molecular and prognostic correlates of cytogenetic abnormalities in chronic myelomonocytic leukemia: a Mayo Clinic-French Consortium Study. *Am J Hematol*. 2014;89(12):1111-1115.
 97. Palomo L, Xicoy B, Garcia O, et al. Impact of SNP array karyotyping on the diagnosis and the outcome of chronic myelomonocytic leukemia with low risk cytogenetic features or no metaphases. *Am J Hematol*. 2016;91(2):185-192.
 98. Steidl C, Steffens R, Gassmann W, et al. Adequate cytogenetic examination in myelodysplastic syndromes: analysis of 529 patients. *Leuk Res*. 2005;29(9):987-993.
 99. ISCN: an International System for Human Cytogenetic Nomenclature (2016). Eds. McGowan-Jordan J, Simons A, Schmid M. Karger Basel, New York, 2016.
 100. Such E, Cervera J, Costa D, et al. Cytogenetic risk stratification in chronic myelomonocytic leukemia. *Haematologica*. 2011;96(3):375-383.
 101. Nomdedeu M, Calvo X, Pereira A, et al.; Spanish Group of Myelodysplastic Syndromes. Prognostic impact of chromosomal translocations in myelodysplastic syndromes and chronic myelomonocytic leukemia patients. A study by the Spanish Group of Myelodysplastic Syndromes. *Genes Chromosomes Cancer*. 2016;55(4):322-327.
 102. Hirsch-Ginsberg C, LeMaistre AC, Kantarjian H, et al. RAS mutations are rare events in Philadelphia chromosome-negative/bcr gene rearrangement-negative chronic myelogenous leukemia, but are prevalent in chronic myelomonocytic leukemia. *Blood*. 1990;76(6):1214-1219.
 103. Kohlmann A, Grossmann V, Haferlach T. Integration of next-generation sequencing into clinical practice: are we there yet? *Semin Oncol*. 2012;39(1):26-36.
 104. Smith AE, Mohamedali AM, Kulasekararaj A, et al. Next-generation sequencing of the TET2 gene in 355 MDS and CMML patients reveals low-abundance mutant clones with early origins, but indicates no definite prognostic value. *Blood*. 2010;116(19):3923-3932.
 105. Patnaik MM, Tefferi A. Chronic myelomonocytic leukemia: 2018 update on diagnosis, risk stratification and management. *Am J Hematol*. 2018;93(6):824-840.
 106. Jankowska AM, Makishima H, Tiu RV, et al. Mutational spectrum analysis of chronic myelomonocytic leukemia includes genes associated with epigenetic regulation: UTX, EZH2, and DNMT3A. *Blood*. 2011;118(14):3932-3941.
 107. Itzykson R, Kosmider O, Renneville A, et al. Clonal architecture of chronic myelomonocytic leukemias. *Blood*. 2013;121(12):2186-2198.
 108. Patel BJ, Przychodzen B, Thota S, et al. Genomic determinants of chronic myelomonocytic leukemia. *Leukemia*. 2017;31(12):2815-2823.
 109. Gelsi-Boyer V, Trouplin V, Roquain J, et al. ASXL1 mutation is associated with poor prognosis and acute transformation in chronic myelomonocytic leukaemia. *Br J Haematol*. 2010;151(4):365-375.
 110. Federmann B, Abele M, Rosero Cuesta DS, et al. The detection of SRSF2 mutations in routinely processed bone marrow biopsies is useful in the diagnosis of chronic myelomonocytic leukemia. *Hum Pathol*. 2014;45(12):2471-2479.
 111. Bally C, Adès L, Renneville A, et al. Prognostic value of TP53 gene mutations in myelodysplastic syndromes and acute myeloid leukemia treated with azacitidine. *Leuk Res*. 2014;38(7):751-755.
 112. Padua RA, Guinn BA, Al-Sabah AI, et al. RAS, FMS and p53 mutations and poor clinical outcome in myelodysplasias: a 10-year follow-up. *Leukemia*. 1998;12(6):887-892.
 113. Ricci C, Ferro E, Corti S, et al. RAS mutations contribute to evolution of chronic myelomonocytic leukemia to the proliferative variant. *Clin Cancer Res*. 2010;16(8):2246-2256.
 114. Wang J, Liu Y, Li Z, et al. Endogenous oncogenic Nras mutation promotes aberrant GM-CSF signaling in granulocytic/monocytic precursors in a murine model of chronic myelomonocytic leukemia. *Blood*. 2010;116(26):5991-6002.
 115. Padron E, Painter JS, Kunigal S, et al. GM-CSF-dependent pSTAT5 sensitivity is a feature with therapeutic potential in chronic myelomonocytic leukemia. *Blood*. 2013;121(25):5068-5077.
 116. Geissler K, Jäger E, Barna A, et al. Chronic myelomonocytic leukemia patients with RAS pathway mutations show high in vitro myeloid colony formation in the absence of exogenous growth factors. *Leukemia*. 2016;30(11):2280-2281.
 117. Itzykson R, Kosmider O, Renneville A, et al. Prognostic score including gene mutations in chronic myelomonocytic leukemia. *J Clin Oncol*. 2013;31(19):2428-2436.
 118. Elena C, Galli A, Such E, et al. Integrating clinical features and genetic lesions in the risk assessment of patients with chronic myelomonocytic leukemia. *Blood*. 2016;128(10):1408-1417.
 119. Palomo L, Garcia O, Arnan M, et al. Targeted deep sequencing improves outcome stratification in chronic myelomonocytic leukemia with low risk cytogenetic features. *Oncotarget*. 2016;7(35):57021-57035.
 120. Onida F. Models of Prognostication in chronic myelomonocytic leukemia. *Curr Hematol Malig Rep*. 2017;12(6):513-521.
 121. Xu Y, McKenna RW, Karandikar NJ, Pildain AJ, Kroft SH. Flow cytometric analysis of monocytes as a tool for distinguishing chronic myelomonocytic leukemia from reactive monocytosis. *Am J Clin Pathol*. 2005;124(5):799-806.
 122. Lacronique-Gazaille C, Chaury MP, Le Guyader A, Faucher JL, Bordessoule D, Feuillard J. A simple method for detection of major phenotypic abnormalities in myelodysplastic syndromes: expression of CD56 in CMML. *Haematologica*. 2007;92(6):859-860.
 123. Kern W, Bacher U, Haferlach C, Schnittger S, Haferlach T. Acute monoblastic/monocytic leukemia and chronic myelomonocytic leukemia share common immunophenotypic features but differ in the extent of aber-

- rantly expressed antigens and amount of granulocytic cells. *Leuk Lymphoma.* 2011;52(1):92-100.
124. Shen Q, Ouyang J, Tang G, et al. Flow cytometry immunophenotypic findings in chronic myelomonocytic leukemia and its utility in monitoring treatment response. *Eur J Haematol.* 2015;95(2):168-176.
125. Selimoglu-Buet D, Wagner-Ballon O, Saada V, et al; Francophone Myelodysplasia Group. Characteristic repartition of monocyte subsets as a diagnostic signature of chronic myelomonocytic leukemia. *Blood.* 2015;125(23):3618-3626.
126. Harrington AM, Schelling LA, Ordobazari A, Olteanu H, Hosking PR, Kroft SH. Immunophenotypes of chronic myelomonocytic leukemia (CMML) subtypes by flow cytometry: a comparison of CMML-1 vs CMML-2, myeloproliferative vs dysplastic, de novo vs therapy-related, and CMML-specific cytogenetic risk subtypes. *Am J Clin Pathol.* 2016;146(2):170-181.
127. Selimoglu-Buet D, Badaoui B, Benayoun E, et al; Groupe Francophone des Myélodysplasies. Accumulation of classical monocytes defines a subgroup of MDS that frequently evolves into CMML. *Blood.* 2017;130(6):832-835.
128. Picot T, Aanei CM, Flandrin Gresta P, et al. Evaluation by flow cytometry of mature monocyte subpopulations for the diagnosis and follow-up of chronic myelomonocytic leukemia. *Front Oncol.* 2018;8:109.
129. Hudson CA, Burack WR, Leary PC, Bennett JM. Clinical utility of classical and nonclassical monocyte percentage in the diagnosis of chronic myelomonocytic leukemia. *Am J Clin Pathol.* 2018;150(4):293-302.
130. Feng R, Bhatt VR, Fu K, Pirruccello S, Yuan J. Application of immunophenotypic analysis in distinguishing chronic myelomonocytic leukemia from reactive monocytosis. *Cytometry B Clin Cytom.* 2018;94(6):901-909.
131. Hudson CA, Burack WR, Bennett JM. Emerging utility of flow cytometry in the diagnosis of chronic myelomonocytic leukemia. *Leuk Res.* 2018;73:12-15.
132. Talati C, Zhang L, Shaheen G, et al. Monocyte subset analysis accurately distinguishes CMML from MDS and is associated with a favorable MDS prognosis. *Blood.* 2017;129(13):1881-1883.
133. Damasceno D, Teodosio C, van den Bossche WBL, et al, on behalf of the TiMaScan Study Group. Distribution of subsets of blood monocytic cells throughout life. *J Allergy Clin Immunol.* 2019;144(1):320-323.e5.
134. Escribano L, Garcia Montero AC, Núñez R, Orfao A; Red Española de Mastocitosis. Flow cytometric analysis of normal and neoplastic mast cells: role in diagnosis and follow-up of mast cell disease. *Immunol Allergy Clin North Am.* 2006;26(3):535-547.
135. Greenberg P, Cox C, LeBeau MM, et al. International scoring system for evaluating prognosis in myelodysplastic syndromes. *Blood.* 1997;89(6):2079-2088.
136. Onida F, Kantarjian HM, Smith TL, et al. Prognostic factors and scoring systems in chronic myelomonocytic leukemia: a retrospective analysis of 213 patients. *Blood.* 2002;99(3):840-809.
137. Such E, Germing U, Malcovati L, et al. Development and validation of a prognostic scoring system for patients with chronic myelomonocytic leukemia. *Blood.* 2013;121(15):3005-3015.
138. Padron E, Garcia-Manero G, Patnaik MM, et al. An international data set for CMML validates prognostic scoring systems and demonstrates a need for novel prognostication strategies. *Blood Cancer J.* 2015;5:e333.
139. Padron E, Komrokji R, List AF. The clinical management of chronic myelomonocytic leukemia. *Clin Adv Hematol Oncol.* 2014;12(3):172-178.
140. Pleyer L, Germing U, Sperr WR, et al. Azacitidine in CMML: matched-pair analyses of daily-life patients reveal modest effects on clinical course and survival. *Leuk Res.* 2014;38(4):475-483.
141. Padron E, Steensma DP. Cutting the cord from myelodysplastic syndromes: chronic myelomonocytic leukemia-specific biology and management strategies. *Curr Opin Hematol.* 2015;22(2):163-170.
142. Solary E, Itzykson R. How I treat chronic myelomonocytic leukemia. *Blood.* 2017;130(2):126-136.
143. Moyo TK, Savona MR. Therapy for chronic myelomonocytic leukemia in a new era. *Curr Hematol Malig Rep.* 2017;12(5):468-477.
144. Hunter AM, Zhang L, Padron E. Current management and recent advances in the treatment of chronic myelomonocytic leukemia. *Curr Treat Options Oncol.* 2018;19(12):67.
145. Diamantopoulos PT, Kotsianidis I, Symeonidis A, et al; Hellenic MDS Study Group. Chronic myelomonocytic leukemia treated with 5-azacytidine - results from the Hellenic 5-Azacytidine Registry: proposal of a new risk stratification system. *Leuk Lymphoma.* 2018;14:1-10.
146. Itzykson R, Fenaux P, Bowen D, et al, on behalf of the European Hematology Association, the European LeukemiaNet. Diagnosis and treatment of chronic myelomonocytic leukemias in adults: recommendations from the European Hematology Association and the European LeukemiaNet. *HemaSphere.* 2018;2(6):e150.
147. Eissa H, Gooley TA, Sorror ML, et al. Allogeneic hematopoietic cell transplantation for chronic myelomonocytic leukemia: relapse-free survival is determined by karyotype and comorbidities. *Biol Blood Marrow Transplant.* 2011;17(6):908-915.
148. de Witte T, Bowen D, Robin M, et al. Allogeneic hematopoietic stem cell transplantation for MDS and CMML: recommendations from an international expert panel. *Blood.* 2017;129(13):1753-1762.
149. Onida F, Barosi G, Leone G, et al. Management recommendations for chronic myelomonocytic leukemia: consensus statements from the SIE, SIES, GITMO groups. *Haematologica.* 2013;98(9):1344-1352.
150. Savona MR, Malcovati L, Komrokji R, et al; MDS/MPN International Working Group. An international consortium proposal of uniform response criteria for myelodysplastic/ myeloproliferative neoplasms (MDS/ MPN) in adults. *Blood.* 2015;125(12): 1857-1865.
151. Drechsler M, Hildebrandt B, Kündgen A, Germing U, Royer-Pokora B. Fusion of H4/D10S170 to PDGFRbeta in a patient with chronic myelomonocytic leukemia and long-term responsiveness to imatinib. *Ann Hematol.* 2007;86(5):353-354.