

### High prevalence of relapse in children with Philadelphia-like acute lymphoblastic leukemia despite risk-adapted treatment

Acute lymphoblastic leukemia (ALL) remains a leading cause of cancer-related death in children and young adults. Since the 1960s, improvements in the treatment of children with ALL have led to 10-year survival rates now exceeding 85%.<sup>1</sup> Philadelphia-like (Ph-like) ALL is characterized by a gene expression profile similar to that of *BCR-ABL1* positive (Ph+) ALL but lacking the *BCR-ABL1* oncogene and similarly, patients experience poor outcome.<sup>2-4</sup> Ph-like ALL is associated with a range of genetic alterations, particularly rearrangements, which activate cytokine receptor and kinase signalling.<sup>2-4</sup> In 2011, the Australian and New Zealand Children's Haematology/Oncology Group (ANZ-CHOG) completed enrolment of patients to a minimal residual disease (MRD) intervention clinical trial, known as ALL8 (*clinicaltrials.gov Identifier: ACTRN12607000302459*). Children were stratified to high-risk regimens based on several criteria, including treatment failure or high MRD at day 79.<sup>5</sup> Overall, 46 children with precursor B-ALL relapsed, and surprisingly 72% (33/46) of these patients were classified as medium-risk.<sup>5</sup> In this retrospective study of ALL8, the frequency of patients with Ph-like ALL and their attendant genomic lesions were studied, and clinical outcomes were compared to those of non Ph-like B-ALL

patients. The incidence of Ph-like ALL was 11.7%, where the majority of these children were reported to be of Caucasian ethnicity and stratified as being either standard- or medium-risk. Significantly, 57.8% (11/19) of Ph-like ALL patients subsequently relapsed compared to 16% (26/143) who were not Ph-like, with significantly inferior event-free and overall survival ( $P < 0.0001$  and  $P = 0.003$ , respectively).

Six hundred and fifty-six patients aged between one and eighteen years were evaluated for eligibility on the ANZ-CHOG ALL8 trial from 2002-2011. Ethical approval was obtained from each institutional Human Research Ethics Committee and parents or legal guardians gave written, informed consent. Two hundred and forty-five patients, selected on the basis of sample accessibility, were available for Ph-like ALL screening (*Online Supplementary Figure S1*). The mean age at diagnosis (6.4 yrs vs. 5.6 yrs,  $P = 0.02$ ) and higher white cell counts (WCC) ( $P < 0.001$ ), were significantly different between those available for analysis and those patients excluded, with the studied group also having a higher number of patients over ten years of age (23% vs. 16%) (*Online Supplementary Table S1*). Risk groups were similar in each cohort and, overall, event-free and relapse-free survival were not significantly different between both groups (*Online Supplementary Figure S2*).

The criteria for stratification to the high-risk group, based upon Berlin-Frankfurt-Münster (BFM) protocols, were the presence of *BCR-ABL1* or *MLL t(4;11)* translocation; poor prednisolone response at day eight; failure to achieve

**Table 1A.** Age, MRD, risk stratification, relapse status (A), rearrangements and variants of patients (B) identified with Ph-like ALL and *P2RY8-CRLF2*.

| Patient code | Age at diagnosis | Final risk stratification | MRD at day 79        | Relapsed | Relapse free survival (years) | SCT |
|--------------|------------------|---------------------------|----------------------|----------|-------------------------------|-----|
| A2489        | 5.2              | High                      | 2x10 <sup>2</sup>    | Y        | 1.77                          | Y   |
| A5258        | 14.9             | High                      | 6x10 <sup>3</sup>    | N        | 4.56                          | Y   |
| A1781        | 3.5              | Medium                    | Negative             | Y        | 4.34                          | N   |
| A5243        | 16.3             | Medium                    | Pos<10 <sup>-4</sup> | Y        | 2.56                          | N   |
| A1516        | 8.2              | Medium                    | Pos<10 <sup>-4</sup> | N*       | 1.91                          | N   |
| A4513        | 13.2             | Medium                    | Negative             | N        | 5.46                          | N   |
| A1725        | 12.7             | Medium                    | Pos<10 <sup>-4</sup> | N        | 5.07                          | N   |
| A2497        | 16.7             | Medium                    | Pos<10 <sup>-4</sup> | Y        | 1.92                          | Y   |
| A3019        | 14.9             | Medium                    | Negative             | Y        | 0.96                          | N   |
| A5164        | 6.2              | Medium                    | Negative             | Y        | 2.26                          | Y   |
| A1702        | 3                | Medium                    | Negative             | N        | 10.86                         | N   |
| A1747        | 3.1              | Medium                    | Pos<10 <sup>-4</sup> | Y        | 2.18                          | Y   |
| A2173        | 8.2              | Medium                    | Negative             | Y        | 2.15                          | N   |
| A3100        | 15               | Medium                    | Negative             | Y        | 1.06                          | N   |
| A5416        | 5.5              | Medium                    | Negative             | Y        | 2.12                          | Y   |
| A3086        | 11               | Medium                    | Negative             | N        | 8.07                          | N   |
| A5428        | 1.5              | Medium                    | Negative             | Y        | 2.54                          | Y   |
| A2481        | 5.9              | Standard                  | Negative             | N        | 2.94                          | N   |
| A2005        | 13.4             | Medium                    | Pos<10 <sup>-4</sup> | N        | 5.78                          | N   |
| A2273        | 5                | Medium                    | Negative             | Y        | 3.54                          | Y   |
| A2426        | 10.7             | Standard                  | Negative             | Y        | 4.51                          | N   |
| A2517        | 4.9              | Standard                  | Negative             | Y        | 2.35                          | Y   |
| A3239        | 2.1              | Medium                    | Negative             | N        | 5.15                          | N   |
| A3700        | 2.7              | Medium                    | Negative             | N        | 6.90                          | N   |
| A4964        | 12.9             | High                      | 5x10 <sup>-4</sup>   | N**      | 1.76                          | Y   |

remission by day 33 or high MRD ( $>5 \times 10^{-4}$ ) at day 79 (Table 1A). Standard- and medium-risk patients received the same standard BFM four-drug induction chemotherapy regimen including a prednisolone pre-phase and intrathecal methotrexate. In addition to the four-drug protocol, high-risk patients received a further three novel intensive blocks of chemotherapy followed by stem cell transplant (SCT) in most cases.<sup>5</sup> All *BCR-ABL1* positive patients also received imatinib.

Determination of Ph-like ALL has differed between cohorts. European studies have favored the term *BCR-ABL1*-like ALL and have used hierarchical clustering (HC) of an Affymetrix gene expression array based on a probe set of 110 genes designed to detect major pediatric ALL subtypes.<sup>3</sup> In contrast, US studies have used a TaqMan Low Density Arrays (TLDA) based approach consisting of either eight or fifteen genes selected by Prediction Analysis for Microarrays (PAM) analysis.<sup>7,8</sup> While there is overlap, the HC model identifies a greater proportion of patients as having a *BCR-ABL1*-like signature, but this approach does not directly identify causative fusions.<sup>9,10</sup> Based on the US approach, we have designed a custom TLDA using nine genes to identify patients with Ph-like ALL.<sup>7,11</sup>

The TLDA was used according to manufacturer's instructions (Thermo Fisher Scientific, MA, USA) to deter-

mine Ph-like status. Genes were selected based upon prior reports<sup>3</sup> with *CRLF2*, *PDGFRB*, *ABL1*, *ABL2* and *EPOR* also included to aid identification of potential fusions (*Online Supplementary Methods*). Reverse transcription polymerase chain reaction (RT-PCR) followed by Sanger sequencing was performed using a panel of 30 known fusions on all TLDA positive cases and those with high *CRLF2* gene expression.<sup>4</sup> Cases with high *CRLF2* expression were also subjected to fluorescent *in situ* hybridization (FISH) to confirm *IGH-CRLF2* fusions. Illumina TruSeq stranded library preparation for messenger ribonucleic acid sequencing (mRNA seq) on the Illumina NextSeq or HiSeq platforms was performed on all TLDA positive and high *CRLF2* cases, with the sole exception being that of a case with low RNA quality (*Online Supplementary Methods*). Patients were classified as having Ph-like ALL if a sample was TLDA positive.

Cytokine or kinase activating lesions have been identified in the majority of childhood and adolescent/young adults (AYA) with Ph-like ALL.<sup>12</sup> Of the 245 childhood B-ALL patients evaluated, eight patients were identified as being *BCR-ABL1* positive and 75 had an *ETV6-RUNX1* fusion, leaving 162 available for Ph-like screening. Nineteen patients (11.7%) were identified as having Ph-like ALL, as determined by TLDA. Rearrangements were identified in 17/19 patients (Table 1B).

**Table 1B.** Age, MRD, risk stratification, relapse status (A), rearrangements and variants of patients (B) identified with Ph-like ALL and *P2RY8-CRLF2*.

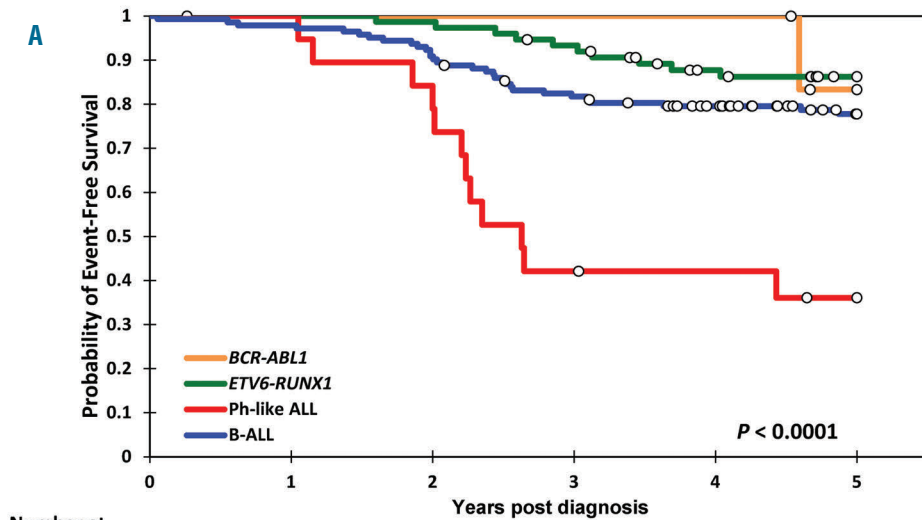
| Patient code | Rearrangement      | TLDA | mRNA sequencing | Variants detected by mRNA and/or Sanger sequencing   | <i>IKZF1</i> deletion |
|--------------|--------------------|------|-----------------|--|-----------------------|
| A2489        | <i>EBF1-PDGFRB</i> | Pos  | Y               | no variants detected   | del 4-7               |
| A5258        | <i>EBF1-PDGFRB</i> | Pos  | Y               | no variants detected   | del 4-7               |
| A1781        | <i>SSBP2-JAK2</i>  | Pos  | Y               | no variants detected   | del 4-8               |
| A5243        | <i>PAX5-JAK2</i>   | Pos  | Y               | no variants detected   | del 2-7               |
| A1516        | <i>IGH-EPOR</i>    | Pos  | Y               | <i>SREBF1</i> pV580M   | del 2-7               |
| A4513        | <i>IGH-EPOR</i>    | Pos  | Y               | <i>ABL2</i> pP608S, <i>SREBF1</i> pV580M, <i>TYK2</i> pG363S, <i>RUNX1</i> pL29S   | del 4-7               |
| A1725        | <i>IGH-CRLF2</i>   | Pos  | Y               | <i>CRLF2</i> pF232C, <i>CDKN2A</i> pA148T  | del 2-8               |
| A2497        | <i>IGH-CRLF2</i>   | Pos  | Y               | <i>CREBBP</i> pN1940S  | del 2-8               |
| A3019        | <i>IGH-CRLF2</i>   | Pos  | Y               | <i>JAK2</i> pR683S   | del 4-7               |
| A5164        | <i>IGH-CRLF2</i>   | Pos  | Y               | <i>CRLF2</i> pF232C  | del 4-7               |
| A1702        | <i>P2RY8-CRLF2</i> | Pos  | Y               | <i>JAK2</i> pI682F   | del 4-7               |
| A1747        | <i>P2RY8-CRLF2</i> | Pos  | Y               | <i>JAK2</i> pT875N, <i>FLT3</i> pD324N   | del 2-7               |
| A2173        | <i>P2RY8-CRLF2</i> | Pos  | Y               | <i>JAK2</i> pR683S   | del 4-7               |
| A3100        | <i>P2RY8-CRLF2</i> | Pos  | Y               | <i>NRAS</i> pG13D, <i>JAK3</i> pP132T  | del 4-7               |
| A5416        | <i>P2RY8-CRLF2</i> | Pos  | Y               | <i>JAK2</i> pR683S, <i>SET2D</i> pM1080I   | del 2-8               |
| A3086        | <i>PSMG1-ERG</i>   | Pos  | Y               | <i>PAX5</i> pG266E, <i>BRAF</i> pA31V  | None                  |
| A5428        | <i>PAX5-ZNF521</i> | Pos  | Y               | <i>ABL1</i> pS972L, <i>ABL2</i> pK909R, <i>CREBBP</i> pV1243I, <i>SREBF1</i> pV580M, <i>TYK2</i> pI684S, <i>RUNX1</i> pE395A | del 4-7               |
| A2481        | Unknown            | Pos  | Y               | no variants detected   | None                  |
| A2005        | Unknown            | Pos  | Y               | <i>IKZF1</i> pN159Y, <i>PTK2B</i> pT65R, <i>BRAF</i> pD594G  | None                  |
| A2273        | <i>P2RY8-CRLF2</i> | Neg  | N               | Poor RNA   | del 4-7               |
| A2426        | <i>P2RY8-CRLF2</i> | Neg  | Y               | <i>KRAS</i> pA146P   | None                  |
| A2517        | <i>P2RY8-CRLF2</i> | Neg  | Y               | <i>TYK2</i> pR568W, <i>RUNX1</i> pL29S   | del 4-7               |
| A3239        | <i>P2RY8-CRLF2</i> | Neg  | Y               | <i>CREBBP</i> pN1940S  | None                  |
| A3700        | <i>P2RY8-CRLF2</i> | Neg  | Y               | no variants detected   | None                  |
| A4964        | <i>P2RY8-CRLF2</i> | Neg  | Y               | <i>NRAS</i> pG12D, <i>SET2D</i> pM1080I  | del 2-8               |

\*Secondary malignancy (AML) at 1.91 years; \*\*died in remission. MRD: minimal residual disease; TLDA: Taqman low density array; mRNA: messenger ribonucleic acid; Pos: positive result; Neg: negative result; Y: yes; N: no; del: deletion of exons; SCT: stem cell transplant.

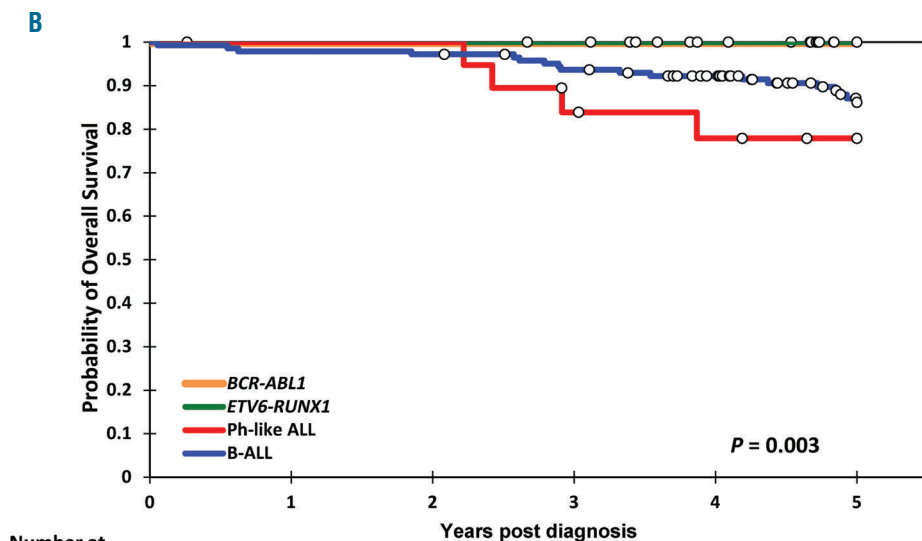
Previous reports have suggested 27-60% of Ph-like ALL patients harbor rearrangements of *CRLF2*, with 50% of these demonstrating concomitant mutations in *JAK1* or *JAK2*.<sup>4,13</sup> Similarly, the majority of ALL8 Ph-like ALL cases (9/19, 47%) harbored *CRLF2* rearrangements (*CRLF2r*), with 77.7% (7/9) demonstrating concomitant *JAK2* or *CRLF2* mutations. While there is some conjecture, the majority of studies have demonstrated *CRLF2* overexpression is significantly associated with poor outcome.<sup>14,15</sup> Importantly, 77.7% (7/9) of ALL8 Ph-like ALL patients with a *CRLF2r*, subsequently relapsed. Interestingly, a further six TLDA negative patients harbored *CRLF2r*. Of these

patients three relapsed, and two subsequently underwent SCT. A fourth patient received a SCT but died in remission. The remaining identified fusions included *EBF1-PDGFRB* and *IGH-EPOR* (n=2), *PAX5-JAK2*, *SSBP2-JAK2*, *PAX5-ZNF521* and *PSMG1-ERG* in one patient each (Table 1B).

*IKZF1* deletions are shown to be significantly associated with relapse risk in both Ph+ and Ph-like ALL, with the frequency reported to be between 27% and 69% in Ph-like ALL cases.<sup>3,12,13</sup> Of note, the ALL8 cohort included B-ALL patients in all risk stratifications, whereas other studies only included high-risk patients, potentially limiting comparisons between groups.<sup>3,12,13</sup> Herein, *IKZF1* deletions



| Number at risk    | 0   | 1   | 2   | 3   | 4   | 5  |
|-------------------|-----|-----|-----|-----|-----|----|
| <i>BCR-ABL1</i>   | 8   | 8   | 8   | 8   | 8   | 4  |
| <i>ETV6-RUNX1</i> | 75  | 75  | 75  | 70  | 60  | 51 |
| Ph-like ALL       | 19  | 19  | 16  | 9   | 8   | 5  |
| B-ALL             | 143 | 141 | 132 | 116 | 105 | 84 |



| Number at risk    | 0   | 1   | 2   | 3   | 4   | 5  |
|-------------------|-----|-----|-----|-----|-----|----|
| <i>BCR-ABL1</i>   | 8   | 8   | 8   | 8   | 8   | 5  |
| <i>ETV6-RUNX1</i> | 75  | 75  | 75  | 75  | 69  | 59 |
| Ph-like ALL       | 19  | 19  | 19  | 16  | 14  | 11 |
| B-ALL             | 143 | 141 | 140 | 133 | 123 | 95 |

Figure 1. Children with Ph-like ALL have inferior survival outcomes compared to other B-ALL patients. Kaplan-Meier analysis with log-rank statistic of A) event free survival and B) overall survival at five years post diagnosis. Children with Ph-like ALL are shown in red, *BCR-ABL1*+ patients in orange, *ETV6-RUNX1* in green and the remaining B-ALL children in blue. The number of patients at risk for the different B-ALL subtypes is shown below the graph at each year.

were significantly associated with Ph-like ALL (84% vs. 14%,  $P < 0.0001$ ). One Ph-like ALL patient, for whom a fusion was not identified, harbored an *IKZF1* p.N159Y mutation detected by mRNA seq and validated in genomic DNA by PCR and Sanger sequencing.

Most studies of patients with Ph-like ALL have demonstrated significantly inferior outcomes, which may be improved with treatment intensification.<sup>3,4,12</sup> In contrast to patients enrolled on Total Therapy XV, a study of risk-directed therapy based on MRD wherein no significant differences in outcome were reported,<sup>13</sup> the ALL8 Ph-like ALL cohort demonstrated significantly inferior event-free ( $P < 0.0001$ ) and overall survival ( $P = 0.003$ ; Figure 1).

On the ALL8 protocol, only two patients with Ph-like ALL had a final high-risk classification (both *EBF1-PDGFRB*) as a result of high MRD at day 79. One non Ph-like *P2RY8-CRLF2* patient was also re-stratified to high-risk as a result of day 79 MRD. All three cases had *IKZF1* deletions; one relapsed and a second died in remission. On ALL8, 72% (8/11) of patients classified as Ph-like ALL relapsed within six months of completing their two years of maintenance therapy (average time to relapse 2.1 years). At five years, the overall survival of Ph-like cases was 78% (15/19), indicating that many patients were salvaged by further therapy or SCT,<sup>5</sup> but their survival rate was still significantly inferior to other B-ALL sub-groups. Similar to that observed in Total Therapy XV, ALL8 patients with Ph-like disease were twice as likely to undergo SCT.<sup>13</sup>

Herein, we demonstrate that despite a risk adjusted treatment approach, there remained a high rate of relapse among children in the ANZCHOG ALL8 study who were retrospectively identified as Ph-like. Of note, the MRD risk stratification used in this protocol did not identify all Ph-like ALL cases as high-risk. Finally, rapid identification of Ph-like disease may guide therapeutic intervention with rationally targeted therapies based on patient specific driving genomic lesions. Tyrosine kinase inhibitors are increasingly utilized in patients with ABL-class fusions, with current and future trials likely to inform drug efficacy in the case of other targets.

Susan L. Heatley,<sup>1,2,3</sup> Teresa Sadras,<sup>1,2</sup> Chung H. Kok,<sup>1,2</sup> Eva Nievergall,<sup>1,2</sup> Kelly Quek,<sup>1</sup> Phuong Dang,<sup>1</sup> Barbara McClure,<sup>1</sup> Nicola Venn,<sup>4</sup> Sarah Moore,<sup>5</sup> Jeffrey Suttle,<sup>5</sup> Tamara Law,<sup>4</sup> Anthea Ng,<sup>3,6</sup> Walter Muskovic,<sup>4</sup> Murray D. Norris,<sup>4</sup> Tamas Revesz,<sup>2,3,7</sup> Michael Osborn,<sup>3,7,8</sup> Andrew S. Moore,<sup>3,9,10</sup> Ram Suppiah,<sup>11</sup> Chris Fraser,<sup>3,9</sup> Frank Alvaro,<sup>3,12</sup> Timothy P. Hughes,<sup>1,2,13</sup> Charles G. Mullighan,<sup>14</sup> Glenn M. Marshall,<sup>3,4,6,8,15</sup> Luciano Dalla Pozza,<sup>3,6</sup> David T. Yeung,<sup>1,2,13</sup> Rosemary Sutton<sup>3,4,8,15</sup> and Deborah L. White<sup>1,2,3,8,16</sup>

<sup>1</sup>Cancer Theme, South Australian Health & Medical Research Institute (SAHMRI), Adelaide, SA, Australia; <sup>2</sup>Discipline of Medicine, University of Adelaide, SA, Australia; <sup>3</sup>Australian and New Zealand Children's Haematology/Oncology Group (ANZCHOG); <sup>4</sup>Molecular Diagnostics, Children's Cancer Institute, Sydney, NSW, Australia; <sup>5</sup>Genetics and Molecular Pathology, SA Pathology, Adelaide, SA, Australia; <sup>6</sup>Sydney Children's Hospital Network, Sydney, NSW, Australia; <sup>7</sup>Women's & Children's Hospital, Adelaide, SA, Australia; <sup>8</sup>Australian Genomic Health Alliance (AGHA); <sup>9</sup>Oncology Services Group, Children's Health Queensland Hospital and Health Service, Brisbane, QLD, Australia; <sup>10</sup>The University of Queensland Diamantina Institute and UQ Child Health Research Centre, The University of Queensland, Brisbane, QLD, Australia; <sup>11</sup>Mater Children's Hospital Brisbane, QLD, Australia; <sup>12</sup>John Hunter Children's Hospital, Newcastle, NSW, Australia; <sup>13</sup>Division of

Haematology, SA Pathology, Adelaide, SA, Australia; <sup>14</sup>Department of Pathology, St Jude Children's Research Hospital, Memphis, TN, USA; <sup>15</sup>School of Women and Children's Health, University of New South Wales, Sydney, NSW, Australia and <sup>16</sup>Discipline of Paediatrics, University of Adelaide, SA, Australia

*Acknowledgments:* we thank the staff at the Tissue Bank of the Children's Research Institute for their assistance.

*Funding:* National Health and Medical Research Council, Australia (APP1057746, APP1044884); Channel 7 Children's Research Fund, Adelaide, SA, Australia; Leukaemia Foundation, Australia; Cancer Council of South Australia, Adelaide, SA, Australia; Beat Cancer, Adelaide, SA, Australia.

*Correspondence:* deborah.white@sahmri.com  
doi:10.3324/haematol.2016.162925

*Information on authorship, contributions, and financial & other disclosures was provided by the authors and is available with the online version of this article at [www.haematologica.org](http://www.haematologica.org).*

## References

- Pui CH, Pei D, Campana D, et al. A revised definition for cure of childhood acute lymphoblastic leukemia. *Leukemia*. 2014; 28(12):2336-2343.
- Mullighan CG, Su X, Zhang J, et al. Deletion of *IKZF1* and prognosis in acute lymphoblastic leukemia. *N Engl J Med*. 2009;360(5):470-480.
- Den Boer ML, van Slegtenhorst M, De Menezes RX, et al. A subtype of childhood acute lymphoblastic leukaemia with poor treatment outcome: a genome-wide classification study. *Lancet Oncol*. 2009; 10(2):125-134.
- Roberts KG, Morin RD, Zhang J, et al. Genetic alterations activating kinase and cytokine receptor signaling in high-risk acute lymphoblastic leukemia. *Cancer Cell*. 2012;22(2):153-166.
- Marshall GM, Dalla Pozza L, Sutton R, et al. High-risk childhood acute lymphoblastic leukemia in first remission treated with novel intensive chemotherapy and allogeneic transplantation. *Leukemia*. 2013;27(7):1497-1503.
- Karsa M, Dalla Pozza L, Venn NC, et al. Improving the identification of high risk precursor B acute lymphoblastic leukemia patients with earlier quantification of minimal residual disease. *PLoS One*. 2013; 8(10):e76455.
- Reshmi SC, Harvey RC, Roberts KG, et al. Targetable kinase gene fusions in high-risk B-ALL: a study from the Children's Oncology Group. *Blood*. 2017;129(25):3352-3361.
- Harvey RC, Kang H, Roberts KG, et al. Development and validation of a highly sensitive and specific gene expression classifier to prospectively screen and identify B-Precursor Acute Lymphoblastic Leukemia (ALL) patients with a Philadelphia chromosome-like ("Ph-like" or "BCR-ABL1-Like") signature for therapeutic targeting and clinical intervention. *Blood*. 2013;122(21):826.
- Boer JM, Marchante JR, Evans WE, et al. BCR-ABL1-like cases in pediatric acute lymphoblastic leukemia: a comparison between DCOG/Erasmus MC and COG/St. Jude signatures. *Haematologica*. 2015;100(9):e354-357.
- Boer JM, Steeghs EM, Marchante JR, et al. Tyrosine kinase fusion genes in pediatric BCR-ABL1-like acute lymphoblastic leukemia. *Oncotarget*. 2017;8(3):4618-4628.
- Roberts KG, Gu Z, Payne-Turner D, et al. High frequency and poor outcome of Philadelphia chromosome-like acute lymphoblastic leukemia in adults. *J Clin Oncol*. 2017;35(4):394-401.
- Roberts KG, Li Y, Payne-Turner D, et al. Targetable kinase-activating lesions in Ph-like acute lymphoblastic leukemia. *N Engl J Med*. 2014; 371(11):1005-1015.
- Roberts KG, Pei D, Campana D, et al. Outcomes of children with BCR-ABL1-like acute lymphoblastic leukemia treated with risk-directed therapy based on the levels of minimal residual disease. *J Clin Oncol*. 2014;32(27):3012-3020.
- Harvey RC, Mullighan CG, Chen IM, et al. Rearrangement of *CRLF2* is associated with mutation of *JAK* kinases, alteration of *IKZF1*, Hispanic/Latino ethnicity, and a poor outcome in pediatric B-progenitor acute lymphoblastic leukemia. *Blood*. 2010;115(26):5312-5321.
- van der Veer A, Waanders E, Pieters R, et al. Independent prognostic value of BCR-ABL1-like signature and *IKZF1* deletion, but not high *CRLF2* expression, in children with B-cell precursor ALL. *Blood*. 2013;122(15):2622-2629.