

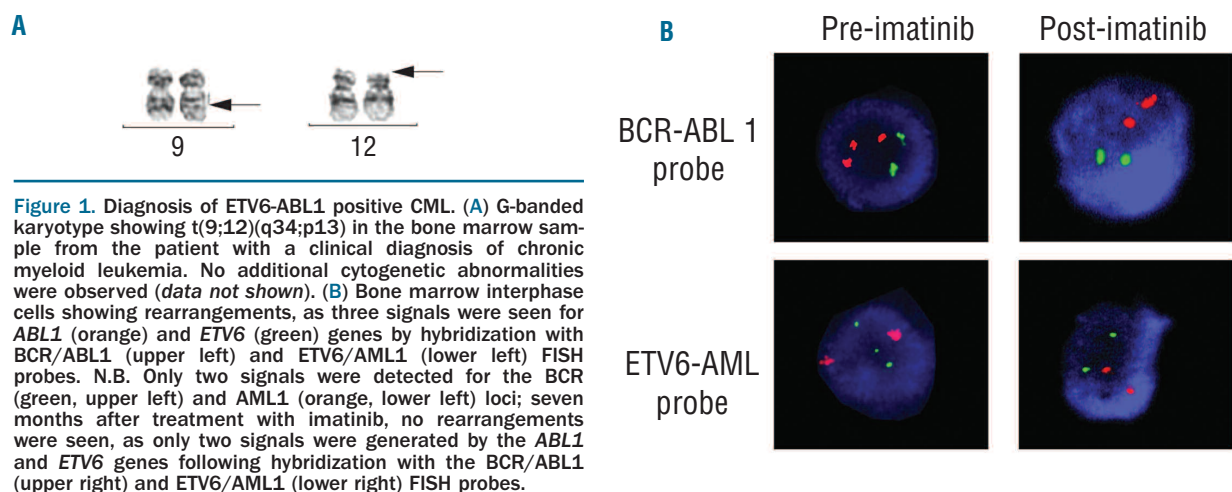
### ETV6-ABL1-positive “chronic myeloid leukemia”: clinical and molecular response to tyrosine kinase inhibition

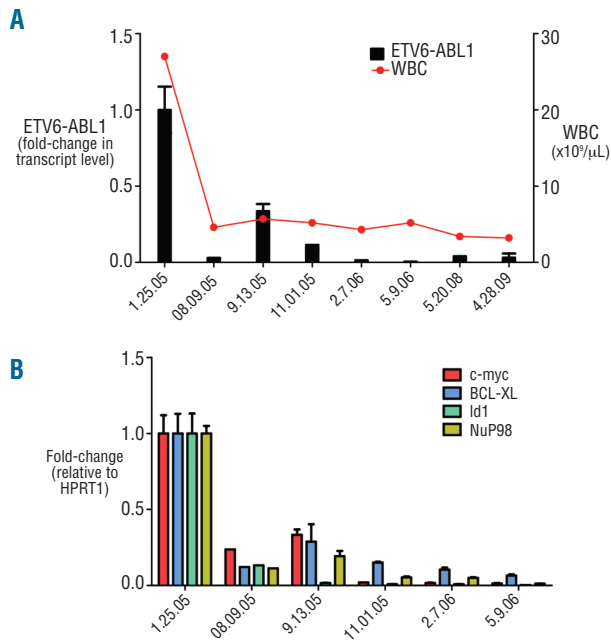
While great progress has been achieved in the clinical management and molecular understanding of Ph<sup>+</sup> chronic myeloid leukemia (CML), little is known about the optimal approach to monitor and treat patients with ETV6-ABL1<sup>+</sup> myeloproliferative neoplasms. Nine BCR-ABL1-negative CML patients with a variant *ABL1* gene (9q34) rearrangement, involving fusion to ETV6 aka TEL (12p13) have been reported thus far.<sup>1-9</sup> The ETV6-ABL1 fusion gene has also been identified in 3 patients with chronic myeloproliferative neoplasms other than “chronic myeloid leukemia” (cMPN) as well as 7 patients with BCR-ABL1 negative acute lymphoblastic leukemia and 4 patients with acute myeloid leukemia.<sup>10,11</sup> Of the 9 cases of ETV6-ABL1<sup>+</sup> chronic myeloid leukemia, only 2 were treated with a TKI in chronic phase<sup>4,5</sup> and only one of these reached a complete remission with a modest follow up of seven months (no molecular monitoring was performed).<sup>4</sup> Among the other published ETV6-ABL1<sup>+</sup> reports, one patient was treated with a second generation TKI for a relapsed cMPN.<sup>12</sup> We provide the first molecular documentation of sustained remission of ETV6-ABL1<sup>+</sup> chronic phase “chronic myeloid leukemia” (CML requires a BCR-ABL1 fusion according to the most recent 2008 WHO classification) to TKI. Molecular monitoring for the ETV6-ABL1 transcript is important given the fact that conventional karyotyping frequently fails to detect a cryptic translocation, i.e. the t(9;12). We provide evidence that treatment of ETV6-ABL1<sup>+</sup> “chronic myeloid leukemia” with imatinib results in downregulation of C-MYC, BCL-XL, ID1 and NUP98, mediators of BCR-ABL1 transforming activity. Moreover, we found no associated mutations in *UTX*, *ASXL1*, *EZH2*, *TET2* and *IDH1/2* suggesting that ETV6-ABL1<sup>+</sup> “chronic myeloid leukemia” may be as tyrosine kinase focused as BCR-ABL1 driven disease.

The patient is a 36-year male who was found to have splenomegaly and a total white blood cell (WBC) count

of  $55 \times 10^9/L$  (57% neutrophils, 6% lymphocytes, 1% monocytes, 3% eosinophils, 2% basophils, 7% metamyelocytes, 24% myelocytes). Lactate dehydrogenase was elevated at 653 IU/L. Bone marrow biopsy revealed myeloid hyperplasia suggestive of a myeloproliferative disorder. qRT-PCR for BCR-ABL1 translocation was negative. No BCR-ABL1 fusion signal was observed in interphase FISH analysis using BCR (22q11.2) and ASS-ABL1 (9q34) probes. Instead, 80% of interphase nuclei showed a variant signal pattern consisting of two signals for BCR and three signals for ASS-ABL1 consistent with rearrangement of ABL1 at 9q34 but not BCR at 22q11. Cytogenetic G-banding analysis and FISH showed t(9;12)(q34;p13) in an otherwise normal karyotype (Figure 1A and B). RT-PCR detected the ETV6-ABL1 translocation and the patient was diagnosed with ETV6-ABL1<sup>+</sup> CML-like disorder. Given the persistence of night sweats and fevers, and the persistent disease despite hydroxyurea at 1,000mg daily (WBC decreased to  $7 \times 10^9$  cells/L after one month of hydroxyurea) imatinib mesylate, 400mg daily, was initiated. The patient tolerated imatinib and achieved a complete hematological remission after three months of treatment (WBC  $5.6 \times 10^9$  cells/L). FISH testing after three months of imatinib revealed no evidence of rearrangement at the BCR or ETV6 loci (Figure 1B).

To follow the patient’s response to therapy more sensitively, qRT-PCR was performed using primers for ETV6 and ABL1 (*Online Supplementary Appendix*). Quantification of the ETV6-ABL1 transcript level in peripheral blood cells one month prior to initiation of imatinib revealed  $2,160 \times 10^3$  ETV6-ABL1 copies/ $\mu$ g RNA. After one month of treatment, ETV6-ABL1 transcript level dropped to  $495 \times 10^3$  copies/ $\mu$ g RNA. The ETV6-ABL1 transcript became undetectable by seven months of treatment, indicating a complete and rapid molecular response. The patient’s molecular response closely mirrored normalization of the WBC count (Figure 2A). We also evaluated the expression of BCR-ABL1 target genes *C-MYC*, *BCL-XL*, *ID1* and *NUP98*, pre- and post-imatinib treatment (Figure 2B). The expression of these genes closely mirrored ETV6-ABL1 expression: imatinib downregulated their expression. The patient has continued to





**Figure 2.** Response to Imatinib in an ETV6-ABL1 positive chronic myeloid leukemia. (A) White blood cell count and peripheral blood qRT-PCR analysis of ETV6-ABL1 transcript levels pre-imatinib and throughout imatinib treatment. (B) Concomitant with decreases in ETV6-ABL1 transcript levels, decreases in C-MYC, ID1, BCL-XL, and NUP-98 transcripts were also seen. The patient remains in hematological remission with no identifiable ETV6-ABL1 transcripts in the peripheral blood after approximately five years of imatinib.

do well on imatinib 400mg/day with no evidence of ETV6-ABL1 transcript by qRT-PCR for the past five years.

Somatic mutations in *UTX*, *ASXL1*, and *TET2* have been reported in chronic myeloid leukemia and mutations in *EZH2* and *IDH1/2* in myeloid malignancies other than CML. We found no somatic alterations in these genes in the DNA extracted from whole blood prior to imatinib treatment, nor when the patient was in a molecular remission.

Our studies indicate that ETV6-ABL1<sup>+</sup> “chronic myeloid leukemia” can be sensitive to imatinib and there is significant overlap of molecular targets of ETV6-ABL1 with those of BCR-ABL1, suggesting that the ETV6-ABL1 fusion protein may trigger similar oncogenic cascades as BCR-ABL1. Finally, we were able to exclude mutations in any of the recently identified “myeloid” genes including *UTX*, *ASXL1*, *EZH2*, *TET2* and *IDH1/2* suggesting that the pathogenesis of ETV6-ABL1<sup>+</sup> “chronic myeloid leukemia” may be as tyrosine kinase focused as BCR-ABL1 driven disease.

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## References

- Andreasson P, Johansson B, Carlsson M, Jarlsfelt I, Fioretos T, Mitelman F, et al. BCR/ABL-negative chronic myeloid leukemia with ETV6/ABL fusion. *Genes Chromosomes Cancer*. 1997;20(3):299-304.
- Barbouti A, Ahlgren T, Johansson B, Hoglund M, Lassen C, Turesson I, et al. Clinical and genetic studies of ETV6/ABL1-positive chronic myeloid leukaemia in blast crisis treated with imatinib mesylate. *Br J Haematol*. 2003;122(1):85-93.
- Brunel V, Lafage-Pochitaloff M, Alcalay M, Pelicci PG, Birg F. Variant and masked translocations in acute promyelocytic leukemia. *Leuk Lymphoma*. 1996;22(3-4):221-8.
- Kawamata N, Dashti A, Lu D, Miller B, Koeffler HP, Schreck R, et al. Chronic phase of ETV6-ABL1 positive CML responds to imatinib. *Genes Chromosomes Cancer*. 2008;47(10):919-21.
- Kelly JC, Shahbazi N, Scheerle J, Jahn J, Suchen S, Christacos NC, et al. Insertion (12;9)(p13;q34q34): a cryptic rearrangement involving ABL1/ETV6 fusion in a patient with Philadelphia-negative chronic myeloid leukemia. *Cancer Genet Cytogenet*. 2009;192(1):36-9.
- Keung YK, Beaty M, Steward W, Jackle B, Pettinati M. Chronic myelocytic leukemia with eosinophilia, t(9;12)(q34;p13), and ETV6-ABL gene rearrangement: case report and review of the literature. *Cancer Genet Cytogenet*. 2002;138(2):139-42.
- Lin H, Guo JQ, Andreeff M, Arlinghaus RB. Detection of dual TEL-ABL transcripts and a Tel-Abl protein containing phosphotyrosine in a chronic myeloid leukemia patient. *Leukemia*. 2002;16(2):294-7.
- Tirado CA, Sebastian S, Moore JO, Gong JZ, Goodman BK. Molecular and cytogenetic characterization of a novel rearrangement involving chromosomes 9, 12, and 17 resulting in ETV6 (TEL) and ABL fusion. *Cancer Genet Cytogenet*. 2005;157(1):74-7.
- Van Limbergen H, Beverloo HB, van Drunen E, Janssens A, Hahlen K, Poppe B, et al. Molecular cytogenetic and clinical findings in ETV6/ABL1-positive leukemia. *Genes Chromosomes Cancer*. 2001;30(3):274-82.
- Janssen JW, Ridge SA, Papadopoulos P, Cotter F, Ludwig WD, Fonatsch C, et al. The fusion of TEL and ABL in human acute lymphoblastic leukaemia is a rare event. *Br J Haematol*. 1995;90(1):222-4.
- Zuna J, Zaliava M, Muzikova K, Meyer C, Lizcova L, Zemanova Z et al. Acute leukemias with ETV6/ABL1 (TEL/ABL) fusion: poor prognosis and prenatal origin. *Genes Chromosomes Cancer*. 2010; 49(10):873-84.
- Nand R, Bryke C, Kroft SH, Divgi A, Bredeson C, Atallah E. Myeloproliferative disorder with eosinophilia and ETV6-ABL gene rearrangement: efficacy of second-generation tyrosine kinase inhibitors. *Leuk Res*. 2009;33(8):1144-6.

## Cytogenetically complex SEC31A-ALK fusions are recurrent in ALK-positive large B-cell lymphomas

Fusion tyrosine kinases involving anaplastic lymphoma kinase (ALK) are central to the pathogenesis of numerous malignancies, in which they represent impor-